

Antimicrobial activity of silver nanoparticles synthesised using Solanum Xanthocarpum extract against oral microbes

Running title:Antimicrobial activity of Solanum Xanthocarpum induced silver nanoparticles

SWATHI.UB¹, SINDHU RAMESH², S.RAJESHKUMAR³, KEERTHANA T⁴, SINDHU RAMESH⁵,

*¹Department of Conservative Dentistry and Endodontics,
Saveetha Dental College and Hospitals,
Saveetha Institute of Medical and Technical Sciences,
Saveetha University,
Chennai, India*

*²Professor, Department of Conservative Dentistry and Endodontics
Saveetha Dental College and Hospitals,
Saveetha Institute of Medical and Technical Sciences,
Saveetha University,
Chennai, India*

*³Department of Pharmacology,
Saveetha Dental College and Hospitals,
Saveetha Institute of Medical and Technical Sciences,
Saveetha University,
Chennai, India*

*⁴Department of Conservative Dentistry and Endodontics,
Saveetha Dental College and Hospitals,
Saveetha Institute of Medical and Technical Sciences,
Saveetha University,
Chennai, India*

Corresponding Author

*⁵Professor, Department of Conservative Dentistry and Endodontics,
Saveetha Dental College and Hospitals,
Saveetha Institute of Medical and Technical Sciences ,
Saveetha University,
162 , PH Road , Chennai 600077,
TamilNadu , India*

ABSTRACT

Introduction

In the branch of nanotechnology, the synthesis of nanoparticles and their development play a significant role because of their wide applications. They are used as antimicrobial agents and prevent the development of multidrug resistant bacteria. All parts of Solanum xanthocarpum plant, including the stem, flowers, fruits and roots have proved to have medicinal properties.

Aim

This study aimed to evaluate antimicrobial activity of silver nanoparticles synthesised using Solanum Xanthocarpum extract against oral microbes.

Materials and Methods

For this study about 25 grams of dried fruit powder of *Solanum Xanthocarpum* was mixed with 250 ml of water, placed in a rotating shaker at room temperature for 48 hours, filtered, air dried in room temperature and stored at 4 degree C. 25 ml of plant filtrate was added into 225 ml of aqueous solution of 1mM silver nitrate for reduction of silver nitrate into Ag⁺ ions and kept at room temperature for 24 hours in a rotating shaker at 28degree C.

Results

Solanum xanthocarpum induced with silver nanoparticles showed good antibacterial activity against oral pathogens.

Conclusion

The use of fruits for the synthesis of silver nanoparticles has many advantages such as, ease with which the process can be scaled up, economic viability and to obtain smaller particle size.

Clinical Significance

The capability of other plant parts such as fruit and root as a capping and reducing agent is not tested and not well defined. In the present study, it was found that fruits were a good source for the synthesis of silver nanoparticles.

Keywords

Antimicrobial Activity; Silver nanoparticles; *Solanum xanthocarpum*; Oral microbes

INTRODUCTION

Nanotechnology occupies tremendous applications in diverse fields such as chemistry, cosmetics, catalysis, energy, plasmonics, opto-electronics and medicine. In the branch of nanotechnology, the synthesis of nanoparticles and their development play a significant role because of their wide applications.

Nanoparticles are used as antimicrobial agents and they are highly effective and acquire huge attention as they satisfy the requirements where antibiotics fail to prevent the development of Multi-Drug Resistant (MDR) mutants. Many studies have explained the advantages of introducing a material with better properties, including the use of nanotechnology to fabricate antimicrobial agents.

Plant mediated biological synthesis of silver nanoparticles is of importance due to its ecofriendliness and its simplicity. Biosynthesis of silver nanoparticles from plants such as *Euphorbia hirtaki*, (Elumalai et al. 2010) *Svensonia hyderabadensis*, (Rao and Savithramma 2011) *Trianthena decandre*, (Geethalakshmi and Sarada 2010) *Shorea tumbuggaia* (Venkateswarlu et al. 2010) have been reported previously.

Solanum Xanthocarpum of the Family Solanaceae is a prickly, perennial, diffuse, patch forming herb, flowering and fruiting throughout the year and it is most commonly found in Southeast Asia, Malaysia, and in all districts of Tamil Nadu, India. (K. M. Mathew 1983) This plant is also known as Yellow berried nightshade/ Indian nightshade, Other names of the plant include *Kandangathiri/ Katakari, Kateli/ Berkateli*. (Pandey 2004) All parts of the plant, including the stem, flowers, fruits and roots have medicinal properties. In India, entire dried plant is used for treating diseases like leprosy, dropsy and cough. (Prempeh and Mensah-Attipoe 2008)

Pharmacological activities of the plant include antibacterial, antifungal, antinociceptive, antioxidant, hypoglycemic and larvicidal properties. (Samiei et al. 2013) According to a study that was done by Pandey et al, seed fumes of this

plant were useful in the relieving tooth pain and pain from gingival swellings and also useful in treatment of fever, rheumatism, pneumonia and other respiratory troubles.(Sheeba 1970)

Previously our team has a rich experience in working on various research projects across multiple disciplines (L. Govindaraju and Gurunathan 2017; A. Christabel et al. 2016; Soh and Narayanan 2013; Mehta et al. 2019; Ezhilarasan, Apoorva, and Ashok Vardhan 2019; Campeau et al. 2014; Kumar and S 2016; S. L. Christabel 2015; Kumar and Rahman 2017; Sridharan, Ramani, and Patankar 2017; Ramesh et al. 2016; Thamaraiselvan et al. 2015; Thangaraj et al. 2016; Ponnulakshmi et al. 2019; “Fluoride, Fluoridated Toothpaste Efficacy and Its Safety in Children - Review” 2018) Now the growing trend in this area motivated us to pursue this project.

This study aimed to evaluate antimicrobial activity of silver nanoparticles synthesised using Solanum Xanthocarpum extract against oral microbes.

MATERIALS AND METHODS

Collection of pathogens

The pathogens used for antimicrobial activity were collected from Tamil Nadu, India. The pathogens used for the study were *E.fecalis*, *Candida Albicans*, *S.Mutans*, *S.Aureus*.

Collection of plant materials

Fruit of Solanum xanthocarpum was collected from Tamilnadu, India. The collected plant materials were brought to the laboratory for plant extraction and for synthesis of silver nanoparticles.

Processing of plant materials

The fruit of Solanum Xanthocarpum was collected and washed thoroughly. The fruit was then cut into smaller pieces for quick drying. Cleaned fruits were shade dried for 10-15 days and the dried plant materials were crushed into fine powder with the help of an electric grinder. The fine powder obtained was stored in an airtight container at room temperature.

Preparation of Solanum xanthocarpum fruit extract

For preparation of Solanum Xanthocarpum fruit extract, 25 grams of dried fruit powder was mixed with 250 ml of distilled water. The solution was stirred for proper mixing. Then the solution was placed in a rotating shaker (100rpm) at room temperature for 48 hours. After incubation, the extract was filtered in Whatman No: 1 filter paper. Finally the filtrate was allowed to air dry at room temperature and dried powder was stored at 4 degree C until it was used.

Synthesis of silver nanoparticles from fruit extract

25 ml of solanum xanthocarpum filtrate was added into 225 ml of aqueous solution of 1mM of silver nitrate for the reduction of silver nitrate into silver ions and kept at room temperature for 24 hours in a rotating shaker at a temperature 28degree C. The solution was kept in the dark to avoid other biological changes. By this process silver nanoparticles were produced by the reduction of silver ions to metallic silver. Silver nanoparticles were determined by the change in colour of the reaction mixture.

Analysis of silver nanoparticles in Solanum Xanthocarpum fruit extract

UV-vis Spectrophotometer analysis

UV-vis spectrophotometer was used for the Spectrophotometer analysis. The reduction of Ag⁺ ions was monitored by measuring the UV-vis spectrum of the reaction medium. The wavelength of the spectrophotometer was taken between 300-550 nm. 2.5 ml of plant filtrate extracts and 22.5 ml of water was prepared as blank for UV-vis Spectrophotometer analysis.

Antibacterial activity

The Muller Hinton agar plates and Rose Bengal agar plates were swabbed with bacterial pathogens and filled with 10-50µl (200-1000µg) of plant extract. The plates were incubated at a temperature of 37OC for 48 hours and after

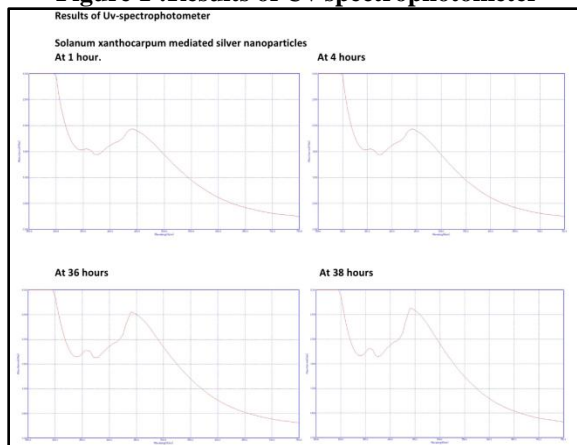
the incubation period, the diameters of zone of inhibition produced by the sample with different organisms in different plates were measured and was recorded.

RESULTS AND DISCUSSION

Analysis of silver nanoparticles in plant extract

Reduction of silver ions to silver nanoparticles were analysed by observing the colour change and UV-Vis spectroscopy.(Figure 1 and Figure 2)

Figure 1 :Results of Uv spectrophotometer



Colour change

The synthesized silver nanoparticles were confirmed by naked observation. Production of silver nanoparticles takes place by the reduction of silver ions during exposure to the plant followed by color change. Within 2 hours the silver ions gets reduced and it exhibits colourless to dark reddish brown colour. This colour change is due to the Surface Plasmon Resonance (SPR) phenomenon.(Figure 2)

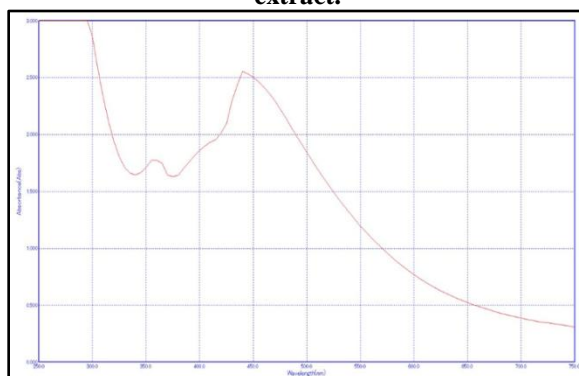
Figure 2 :Colour change observed before and after addition of silver nanoparticles.



UV-VIS Spectra analysis

Wavelength between 400-450nm the formation of silver nanoparticles reach the peak maximum. The specific characteristic peak for silver nanoparticles was due to the SPR. The UV-Visible spectrum shows the formation of silver nanoparticles of aqueous fruit extract as the peak maxima in 440 nm. This is characteristic to silver nanoparticles and the broadening of peak indicated that the particles were polydispersed.(Figure 3)

Figure 3:UV-vis spectrograph of Ag nanoparticle synthesized from Solanum xanthocarpum aqueous fruit extract.



Antibacterial activity of aqueous fruit extract of Solanum xanthocarpum induced with silver nanoparticles against pathogens.

The activity was limited in *Candida albicans* spp (14 mm). The best activity was found in *E.fecalis*(35 mm),followed by *S.mutans* (23mm), and *S.aureus* sp. (20 mm) in 100 µg concentration. (Figure 4)(Figure 5)

Figure 4: Zone of inhibition against various oral pathogens.

Organisms	Concentration 25 ul	Concentration 50 ul	Concentration 100 ul	Concentration AB
S.Mutans	16	19	23	24
Candida Albicans	13	15	14	12
S.Aureus	16	18	20	25
E.fecalis	29	31	35	45

Figure 5:Solanum Xanthocarpum induced silver nanoparticles against various oral pathogens.



The synthesis of nanoparticles is now emerging field because of its eco friendly route.(Chen, Lin, and Ma 2003; Ingle et al. 2008)In the present study, Ag nanoparticles from aqueous extract of fruits of *Solanum xanthocarpum* were studied.Ankanna et al (Ankanna et al. 2010)say by the aqueous extract it gives colour change from colourless to dark yellowish brown in colour. In the present study, the extract changed its colour from colourless to dark reddish brown.The formation of Ag nanoparticles was confirmed by UV-Vis spectral analysis. According to a study

done by Govindaraju et al., (K. Govindaraju et al. 2010) had explained the same family Solanaceae and it was found that the plant *Solanum torvum* got a maximum peak at around 434 nm. The study done by Udayakumar et al., reported (Udayakumar et al. 2003) that the leaf extract of *Solanum xanthocarpum* inhibits *Klebsiella pneumoniae* (18mm) and *E.coli* (7mm) in 100 µl concentrations. According to the study done by Satyavani et al., (Ramanathan, Satyavani, and Gurudeeban 2011) suggested that leaf aqueous extract of silver nanoparticles of *Citrullus colocynthis* showed maximum activity against *E. coli*, *B. subtilis*, *P. aeruginosa* and *Streptococcus pyogenes* but it showed no activity against *P. mirabilis*, *Salmonella enteritis* and *Staphylococcus aureus*. One of the main reasons for endodontic failure is the presence of some species of bacteria inside the root canal system such as *Enterococcus faecalis*. These bacteria are more resistant to disinfection agents, causing a persistent intra-radicular or extra-radicular infection. (Alghamdi and Shakir 2020) *E. faecalis* has the capacity for growing as a biofilm on root canal walls without synergistic support from other bacteria, has high resistance to antimicrobial agents and is a very resistance pathogen to root canal treatment. (Estrela et al. 2008; Sassone et al. 2008) It possesses many survival mechanisms to live in unfavorable conditions, such as to grow in an environment with low oxygen, at high pH, at a wide range of temperatures between 10° and 60°, at high salinity or in a poorly nutrient environment. (Pinheiro et al. 2003; Rôças, Siqueira, and Santos 2004; Jhajharia et al. 2015; Narayanan and Vaishnavi 2010).

Our institution is passionate about high quality evidence based research and has excelled in various fields (Jayaseelan Vijayashree Priyadharsini 2019; Pc, Marimuthu, and Devadoss 2018; Ramesh et al. 2018; Ramadurai et al. 2019; Sridharan et al. 2019; Ezhilarasan, Apoorva, and Ashok Vardhan 2019; M. G. Mathew et al. 2020; Samuel 2021; R et al. 2020; Chandrasekar et al. 2020; J. Vijayashree Priyadharsini, Smiline Girija, and Paramasivam 2018)

In this current study *Solanum Xanthocarpum* induced with silver nanoparticles showed excellent antimicrobial activity. Highest antimicrobial activity was seen against *Enterococcus faecalis*. The activity was limited in *Candida albicans* spp (14 mm). The best activity was found in *E. faecalis* (35 mm), followed by *S. mutans* (23mm), and *S. aureus* sp. (20 mm) in 100 µg concentration.

CONCLUSION

Herbal medicine is gaining growing interest because of its cost effective and eco friendliness. The reduction of the metal ions through plant extracts leading to the formation of silver nanoparticles has been explained before but the capability of parts of the plant such as fruits as a capping and reducing agent has not been explained in previous studies. In the present study, we found that fruits were a good source for the synthesis of silver nanoparticles. It has many advantages such as, ease with which the process can be scaled up, economic viability and to obtain smaller particle size. This study demonstrated the antimicrobial activity

of using biologically synthesized silver nanoparticles against various oral pathogens. The preparation of silver nanoparticle by using *Solanum xanthocarpum* extracts has excellent antimicrobial activity against *Enterococcus faecalis*. Hence this technology can be applied in therapeutics.

Acknowledgement

With Sincere gratitude, we acknowledge the staff members of the department of Conservative Dentistry and Endodontics, Saveetha Dental College and study participants for their extended support towards the completion of research.

Financial Support and Sponsorship

Nil

Conflicts of Interest

There are no conflicts of interest.

REFERENCES

- [1] Alghamdi, Faisal, and Marwa Shakir. 2020. "The Influence of *Enterococcus Faecalis* as a Dental Root Canal Pathogen on Endodontic Treatment: A Systematic Review." *Cureus* 12 (3): e7257.

- [2] Ankanna, Stnkv, Tnvkv Prasad, E. K. Elumalai, and N. Savithramma. 2010. "Production of Biogenic Silver Nanoparticles Using Boswellia Ovalifoliolata Stem Bark." *Digest Journal of Nanomaterials and Biostructures* 5 (2): 369–72.
- [3] Campeau, Philippe M., Dalia Kasperaviciute, James T. Lu, Lindsay C. Burrage, Choel Kim, Mutsuki Hori, Berkley R. Powell, et al. 2014. "The Genetic Basis of DOORS Syndrome: An Exome-Sequencing Study." *Lancet Neurology* 13 (1): 44–58.
- [4] Chandrasekar, Raghavan, Shyamala Chandrasekhar, K. K. Shantha Sundari, and Poornima Ravi. 2020. "Development and Validation of a Formula for Objective Assessment of Cervical Vertebral Bone Age." *Progress in Orthodontics* 21 (1): 38.
- [5] Chen, J. C., Z. H. Lin, and X. X. Ma. 2003. "Evidence of the Production of Silver Nanoparticles via Pretreatment of Phoma sp.3.2883 with Silver Nitrate." *Letters in Applied Microbiology* 37 (2): 105–8.
- [6] Christabel, A., P. Anantanarayanan, P. Subash, C. L. Soh, M. Ramanathan, M. R. Muthusekhar, and V. Narayanan. 2016. "Comparison of Pterygomaxillary Dysjunction with Tuberosity Separation in Isolated Le Fort I Osteotomies: A Prospective, Multi-Centre, Triple-Blind, Randomized Controlled Trial." *International Journal of Oral and Maxillofacial Surgery* 45 (2): 180–85.
- [7] Christabel, S. Linda. 2015. "Prevalence of Type of Frenal Attachment and Morphology of Frenum in Children, Chennai, Tamil Nadu." *World Journal of Dentistry* 6 (4): 203–7.
- [8] Elumalai, E. K., Tnvkv Prasad, J. Hemachandran, S. Viviyana Therasa, T. Thirumalai, and E. David. 2010. "Extracellular Synthesis of Silver Nanoparticles Using Leaves of Euphorbia Hirta and Their Antibacterial Activities." *J Pharm Sci Res* 2 (9): 549–54.
- [9] Estrela, Carlos, Julio Almeida Silva, Ana Helena Gonçalves de Alencar, Claudio Rodrigues Leles, and Daniel Almeida Decurcio. 2008. "Efficacy of Sodium Hypochlorite and Chlorhexidine against Enterococcus Faecalis--a Systematic Review." *Journal of Applied Oral Science: Revista FOB* 16 (6): 364–68.
- [10] Ezhilarasan, Devaraj, Velluru S. Apoorva, and Nandhigam Ashok Vardhan. 2019. "Syzygium Cumini Extract Induced Reactive Oxygen Species-Mediated Apoptosis in Human Oral Squamous Carcinoma Cells." *Journal of Oral Pathology & Medicine: Official Publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology* 48 (2): 115–21.
- [11] "Fluoride, Fluoridated Toothpaste Efficacy and Its Safety in Children - Review." 2018. *International Journal of Pharmaceutical Research* 10 (04). <https://doi.org/10.31838/ijpr/2018.10.04.017>.
- [12] Geethalakshmi, R., and D. V. L. Sarada. 2010. "Synthesis of Plant-Mediated Silver Nanoparticles Using Trianthema Decandra Extract and Evaluation of Their Anti Microbial Activities." *International Journal of Engineering Science and Technology* 2 (5): 970–75.
- [13] Govindaraju, K., S. Tamilselvan, V. Kiruthiga, and G. Singaravelu. 2010. "Biogenic Silver Nanoparticles by Solanum Torvum and Their Promising Antimicrobial Activity." *Journal of Biopesticides* 3 (Special Issue): 394.
- [14] Govindaraju, Lavanya, and Deepa Gurunathan. 2017. "Effectiveness of Chewable Tooth Brush in Children-A Prospective Clinical Study." *Journal of Clinical and Diagnostic Research: JCDR* 11 (3): ZC31–34.
- [15] Ingle, Avinash, Aniket Gade, Sebastien Pierrat, Carsten Sonnichsen, and Mahendra Rai. 2008. "Mycosynthesis of Silver Nanoparticles Using the Fungus Fusarium Acuminatum and Its Activity Against Some Human Pathogenic Bacteria." *Current Nanoscience* 4 (2): 141–44.

- [16] Jhajharia, Kapil, Abhishek Parolia, K. Vikram Shetty, and Lata Kiran Mehta. 2015. "Biofilm in Endodontics: A Review." *Journal of International Society of Preventive & Community Dentistry* 5 (1): 1–12.
- [17] Kumar, Santhosh, and Reshma Rahman. 2017. "Knowledge, Awareness, and Practices Regarding Biomedical Waste Management among Undergraduate Dental Students." *Asian Journal of Pharmaceutical and Clinical Research* 10 (8): 341.
- [18] Kumar, Santhosh, and Sneha S. 2016. "Knowledge and Awareness Regarding Antibiotic Prophylaxis for Infective Endocarditis among Undergraduate Dental Students." *Asian Journal of Pharmaceutical and Clinical Research*, September, 154.
- [19] Mathew, K. M. 1983. "The Flora of the Tamil Nadu Carnatic, The Rapinet Herbarium, St." *Joseph's College, Tiruchirapalli, India*.
- [20] Mathew, Mebin George, S. R. Samuel, Ashu Jagdish Soni, and Korishettar Basavaraj Roopa. 2020. "Evaluation of Adhesion of Streptococcus Mutans, Plaque Accumulation on Zirconia and Stainless Steel Crowns, and Surrounding Gingival Inflammation in Primary Molars: Randomized Controlled Trial." *Clinical Oral Investigations*, 1–6.
- [21] Mehta, Meenu, Deeksha, Devesh Tewari, Gaurav Gupta, Rajendra Awasthi, Harjeet Singh, Parijat Pandey, et al. 2019. "Oligonucleotide Therapy: An Emerging Focus Area for Drug Delivery in Chronic Inflammatory Respiratory Diseases." *Chemico-Biological Interactions* 308 (August): 206–15.
- [22] Narayanan, L. Lakshmi, and C. Vaishnavi. 2010. "Endodontic Microbiology." *Journal of Conservative Dentistry: JCD* 13 (4): 233–39.
- [23] Pandey, Hari Prakash. 2004. "Seed Fume of Solanum Surattense: A Traditional Panacea for Teeth and Gums." <http://nopr.niscair.res.in/handle/123456789/9344>.
- [24] Pc, J., T. Marimuthu, and P. Devadoss. 2018. "Prevalence and Measurement of Anterior Loop of the Mandibular Canal Using CBCT: A Cross Sectional Study." *Clinical Implant Dentistry and Related Research*. <https://europepmc.org/article/med/29624863>.
- [25] Pinheiro, E. T., B. P. F. A. Gomes, C. C. R. Ferraz, E. L. R. Sousa, F. B. Teixeira, and F. J. Souza-Filho. 2003. "Microorganisms from Canals of Root-Filled Teeth with Periapical Lesions." *International Endodontic Journal* 36 (1): 1–11.
- [26] Ponnulakshmi, R., B. Shyamaladevi, P. Vijayalakshmi, and J. Selvaraj. 2019. "In Silico and in Vivo Analysis to Identify the Antidiabetic Activity of Beta Sitosterol in Adipose Tissue of High Fat Diet and Sucrose Induced Type-2 Diabetic Experimental Rats." *Toxicology Mechanisms and Methods* 29 (4): 276–90.
- [27] Prempeh, Aba, and J. Mensah-Attipoe. 2008. "Analgesic Activity of Crude Aqueous Extract of the Root Bark of Zanthoxylum Xanthoxyloides." *Ghana Medical Journal* 42 (2): 79–84.
- [28] Ramadurai, Neeraja, Deepa Gurunathan, A. Victor Samuel, Emg Subramanian, and Steven J. L. Rodrigues. 2019. "Effectiveness of 2% Articaine as an Anesthetic Agent in Children: Randomized Controlled Trial." *Clinical Oral Investigations* 23 (9): 3543–50.
- [29] Ramanathan, T., K. Satyavani, and S. Gurudeeban. 2011. "Plant Mediated Synthesis of Biomedical Silver Nanoparticles by Using Leaf Extract of Citrullus Colocynthis." *Research Journal of Nanoscience and Nanotechnology*. <https://doi.org/10.3923/rjnn.2011.95.101>.
- [30] Ramesh, Asha, Sheeja Varghese, Nadathur D. Jayakumar, and Sankari Malaiappan. 2018. "Comparative Estimation of Sulfiredoxin Levels between Chronic Periodontitis and Healthy Patients - A Case-Control

Study.” *Journal of Periodontology* 89 (10): 1241–48.

- [31] Ramesh, Asha, Sheeja Saji Varghese, Jayakumar Nadathur Doraiswamy, and Sankari Malaiappan. 2016. “Herbs as an Antioxidant Arsenal for Periodontal Diseases.” *Journal of Intercultural Ethnopharmacology* 5 (1): 92–96.
- [32] Rao, M. L., and N. Savithramma. 2011. “Biological Synthesis of Silver Nanoparticles Using *Svensonia Hyderabadensis* Leaf Extract and Evaluation of Their Antimicrobial Efficacy.” *Journal of Pharmaceutical Sciences*. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.447.6711&rep=rep1&type=pdf>.
- [33] R, Hannah, R. Hannah, Pratibha Ramani, Arvind Ramanathan, Jancy Merlin R, S. Gheena, Abilasha Ramasubramanian, and K. Monika. 2020. “CYP2 C9 Polymorphism among Patients with Oral Squamous Cell Carcinoma and Its Role in Altering the Metabolism of Benzo[a]pyrene.” *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. <https://doi.org/10.1016/j.oooo.2020.06.021>.
- [34] Rôças, Isabela N., José F. Siqueira Jr, and Kátia R. N. Santos. 2004. “Association of *Enterococcus Faecalis* with Different Forms of Periradicular Diseases.” *Journal of Endodontia* 30 (5): 315–20.
- [35] Samiei, Mohammad, Mohammad Aghazadeh, Mehrdad Lotfi, Sahar Shakoei, Zahra Aghazadeh, and Seyyed Mahdi Vahid Pakdel. 2013. “Antimicrobial Efficacy of Mineral Trioxide Aggregate with and without Silver Nanoparticles.” *Iranian Endodontic Journal* 8 (4): 166–70.
- [36] Samuel, Srinivasan Raj. 2021. “Can 5-Year-Olds Sensibly Self-Report the Impact of Developmental Enamel Defects on Their Quality of Life?” *International Journal of Paediatric Dentistry / the British Paedodontic Society [and] the International Association of Dentistry for Children* 31 (2): 285–86.
- [37] Sassone, Luciana M., Rivail Antonio Sergio Fidel, Cristiana Francescutti Murad, Sandra Rivera Fidel, and Rafael Hirata Jr. 2008. “Antimicrobial Activity of Sodium Hypochlorite and Chlorhexidine by Two Different Tests.” *Australian Endodontic Journal: The Journal of the Australian Society of Endodontology Inc* 34 (1): 19–24.
- [38] Sheeba, E. 1970. “Antibacterial Activity of *Solanum Surattense* Burm. F.” *Kathmandu University Journal of Science, Engineering and Technology*. <https://doi.org/10.3126/kuset.v6i1.3278>.
- [39] Soh, C. L., and V. Narayanan. 2013. “Quality of Life Assessment in Patients with Dentofacial Deformity Undergoing Orthognathic Surgery--a Systematic Review.” *International Journal of Oral and Maxillofacial Surgery* 42 (8): 974–80.
- [40] Sridharan, Gokul, Pratibha Ramani, and Sangeeta Patankar. 2017. “Serum Metabolomics in Oral Leukoplakia and Oral Squamous Cell Carcinoma.” *Journal of Cancer Research and Therapeutics* 13 (3): 556–61.
- [41] Sridharan, Gokul, Pratibha Ramani, Sangeeta Patankar, and Rajagopalan Vijayaraghavan. 2019. “Evaluation of Salivary Metabolomics in Oral Leukoplakia and Oral Squamous Cell Carcinoma.” *Journal of Oral Pathology & Medicine: Official Publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology* 48 (4): 299–306.
- [42] Thamaraiselvan, Murugan, Sugumari Elavarasu, Suthanthiran Thangakumaran, Jayaprakash Sharanabasappa Gadagi, and Thangavelu Arthie. 2015. “Comparative Clinical Evaluation of Coronally Advanced Flap with or without Platelet Rich Fibrin Membrane in the Treatment of Isolated Gingival Recession.” *Journal of Indian Society of Periodontology* 19 (1): 66–71.
- [43] Thangaraj, Soundara Viveka, Vidyarani Shyamsundar, Arvind Krishnamurthy, Pratibha Ramani, Kumaresan Ganesan, Muthulakshmi Muthuswami, and Vijayalakshmi Ramshankar. 2016. “Molecular Portrait of Oral Tongue Squamous Cell Carcinoma Shown by Integrative Meta-Analysis of Expression

Profiles with Validations.” *PloS One* 11 (6): e0156582.

- [44] Udayakumar, R., K. Velmurugan, D. Srinivasan, and Raghu Ram Krishna. 2003. “Phytochemical and Antimicrobial Studies of Extracts of *Solanum Xanthocarpum*.” *Ancient Science of Life* 23 (2): 90–94.
- [45] Venkateswarlu, P., S. Ankanna, Elumalai E. K. Prasad TNVKV, P. C. Nagajyothi, and N. Savithramma. 2010. “Green Synthesis of Silver Nanoparticles Using *Shorea Tumbuggaia* Stem Bark.” *American Journal of Respiratory Medicine: Drugs, Devices, and Other Interventions* 2 (4): 720–23.
- [46] Vijayashree Priyadharsini, Jayaseelan. 2019. “In Silico Validation of the Non-Antibiotic Drugs Acetaminophen and Ibuprofen as Antibacterial Agents against Red Complex Pathogens.” *Journal of Periodontology* 90 (12): 1441–48.
- [47] Vijayashree Priyadharsini, J., A. S. Smiline Girija, and A. Paramasivam. 2018. “In Silico Analysis of Virulence Genes in an Emerging Dental Pathogen *A. Baumannii* and Related Species.” *Archives of Oral Biology* 94 (October): 93–98.