

In Silico Study of Single Nucleotide Polymorphisms by Polg2 Gene

S.T. Gopukumar¹, R. Sonny Mon², Arun R Nair³, Sasikanth S.M⁴ and Lekshmi Gangadhar^{5*}

¹*Department of Medical Research, Sarada Krishna Homoeopathic Medical College, Kulasekharam, Kanniyakumari District, Tamil Nadu 629161*

²*Department of Organon of Medicine, Sarada Krishna Homoeopathic Medical College, Kulasekharam, Kanniyakumari District, Tamil Nadu 629161*

³*Department of Practice of Medicine, Sarada Krishna Homoeopathic Medical College, Kulasekharam, Kanniyakumari District, Tamil Nadu 629161*

⁴*Department of Physics, Noorul Islam Centre for Higher Education, Thuckalay, Kumarakoil 629180*

⁵*Department of Nanotechnology, Nanodot Research Pvt. Ltd, Trivandrum 695124*

ABSTRACT

In the knowledge of the genetic origin of several complicated human disorders, single-nucleotide polymorphisms (SNPs) play a prominent part. Also, understanding the roles of these SNPs may explain the biology of human phenotype heterogeneity. It would still be a big difficulty to describe the gene linked to disease, operational SNPs. We have studied the genetically variation in this study that can affect the expression and functioning of the POLG2 gene by utilizing the *in-silico* approaches. Among the total of 5828 SNPs, nonsynonymous (ns) SNPs were identified to be 341 and then 3 were recognized as pathogenic. Our analysis was able to classify the future prospects. nsSNPs which can be utilized for certain diseases that occur as just a genetic diagnostic tool. Perhaps a nsSNP is based on a correlation of the stabilization sequences of native and mutant proteins (rs104894632) mitochondrial disorders induced by the POLG2 gene may be a significant candidate.

KEYWORDS: Binding affinity, Genetic Disorder, POLG2, PolyPhen-2, Single Nucleotide Polymorphism.

*Corresponding Author Email: lekshmi.gangadhar@gmail.com

INTRODUCTION

Single Nucleotides Polymorphisms (SNPs) responsible for the maximum communal type of hereditary change in humans. Regarding throughout a coding areas of mammalian genomes, 500,000 SNPs fell into it ^[1]. Amongst, these the source of nonsynonymous SNPs (nsSNPs) variations in the metabolites of amino acids. It is probable those are a significant contributing factor to the structural complexity of the human population, encoded proteins ^[2]. About the nsSNPs by modifying DNA and transcription factors, gene regulation affects binding features ^[3] and then preservation of a stability of the system in skins and cells ^[4]. In addition, nsSNPs

influence the functional functions enzymes in the visual, hormonal, and signaling transduction as additional stimulants ^[5, 6].

Mutations in the POLG 2 gene leads to many disorders. Disorders which affect mitochondrial maintenance DNA (mtDNA) is characterized through degradation or numerous deletions in mtDNA in post-mitotic regions ^[7,8]. To in combination with these medical disorders, mutations within 12 nuclear-encoded genes were designated as date ^[9]. Mutations through one of those, the gene that encodes the POLG DNA polymerase gamma, the mitochondrial DNA-polymerase catalytic sub-unit, is by far the utmost normal nuclear gene that causes mitochondrial abnormalities. In fact, the DNA polymerase gamma mutation repository has documented more than 170 variants ^[10]. There are diverse medical phenotypes correlated by POLG mutations, varying within harshness between persistent exterior ophthalmoplegia ^[11] to mitochondrial heritable ataxia-syndrome ^[12] to progressive outside ophthalmoplegia, syndrome of Alpers-Huttenlocher ^[13]. Here we mention a syndrome of hereditary adult-onset ataxia, correlated within 3 new variants of POLG nucleotides. The numerical method POLG2 research has been carried out and shows that there is one of these is pathogenic.

We conducted this research primarily to carry out the POLG 2 gene, a computational study of the nsSNPs to pinpoint the mutations that are probable and suggest a model structure with the protein from the mutant. We're reporting that the glycine mutation to glutamic acid in the native protein at that residue location of 451 in the POLG 2 gene may be a prototype of significant concern for the POLG2-gene-induced disease.

MATERIALS AND METHODS

Sorting Intolerant from Tolerant, Polymorphism Phenotyping v2, SNAP2, PhD-SNP, SNPs & GO, PyMut, MUpPro, ConSurf, Ensemble Learning Approach for Stability Prediction of Interface and Core Mutations and mCSM-PPI2, respectively. The SNPs and its associated receptor series for POLG 2 gene was reclaimed from the dbSNP (<http://www.ncbi.nlm.nih.gov/SNP/>) in this *in-silico* study.

Extrapolation of functional effect of SNPs

POLG 2 human gene SNP [Accession number Q9UHN1] was reclaimed from dbSNPs of National Center for Biotechnology Information (NCBI) and UniProt Protein Database (<http://www.uniprot.org>). The assessment of the efficient impact on the POLG 2 gene due to the harmful nsSNPs were conceded out by employing Sorting Intolerant from Tolerant (SIFT), PolyPhen-2 and SNAP2. SIFT utilizes the PSI-BLAST database receptor thereby the consistent functional genetic variations were extracted. Extrapolation of the impact of amino acid replacement was used by employing SIFT input sequences, the impact was categorized as

either tolerated or deleterious. Polymorphism phenotyping v2 (PolyPhen-2) is a website related approach that estimates whether amino acid replacement profits in a conserved area and whether the replacement has a destructive impact on the receptor sequence manner. Variations were enlisted as possibly damaging, probably damaging as well as benign. SNAP2 processes SNP's functional impact utilizing neural machine-based learning platform. PROVEAN receptor variation impact analyzer evaluates the amino-acid replacement effects on receptor functions via alignment-based scoring method.

Extrapolation of disorder connotation of SNPs

PhD-SNP (<http://snps.biofold.org/phd-snp/phd-snp.html>) and SNPs&GO (<http://snps-andgo.biocomp.unibo.it/snps-and-go/>) were utilized for defining connotation of clarified SNPs with disorder. PhD-SNP was an online-based server utilized to examine the relationship of disorders by the SNPs to the precise rate of 78 percent. Categorizing the SNPs to disorder linked or neutral it positioned in a scale of 0 to 9. SNPs & GO is precise technique that evaluates disorder related amino acid variation within a one place within a particular receptor like functional arrangements by total precision of 82 % predictively. The queries provided to SNP database and gene ontology (SNPs & GO) was UniProt accession number of POLG 2 receptor and mutation location of both native and mutated amino-acid. Piezoelectric Micro machined Ultrasonic Transducers (PMut) was established utilizing PyMut repositories. It envisages if a SNP is either disease instigating or neutral.

Effect on receptor stability

The SNPs incline into impact receptor asset through moreover declining or enhancing the receptor steadiness. To evaluate this impact, the pairing of techniques was utilized to exploit the assurance of a modifications produced. I-Mutant (<http://folding.biofold.org/i-mutant/i-mutant2.0.html>), envisages an effect of SNPs within fluctuating constant condition of the receptor. The precision of the strategy goes nearly 77 percent. The query for I-Mutant was POLG 2 protein amino-acid series and mutations of residues alongside situations. MUpro, an assembly comprising numerous machine learning-based programs, ascertains the alterations in receptor condition and strength owing to its effect of amino-acid mutation. The queries for MUpro was I-Mutant but MUpro also begins the locations of replacements alongside unique and mutated residue.

Sequence-based conservation study

ConSurf, the web-based online technique was utilized for POLG-2 receptor conservation study (<http://ConSurf.tau.ac.il/>). ConSurf was the effective study for envisaging a higher-throughput features of a goal areas of the receptors. For each residues of receptor of attention, the conservation study was exposed on the scaling of 1-9. Within the scale, 1 to 3 score was

mentioned as variable, 4–6 was denoted to be average and 7–9 scores were displaying higher conserved areas. This method uses a query as FASTA protein sequence.

Stability Prediction Impact in domain-domain Interfaces and domain cores as well as missense mutation

Ensemble Learning Approach for Stability Prediction of Interface and Core Mutations (ELASPIC) (<https://pubmed.ncbi.nlm.nih.gov/26801957/>) evaluates the impact of mutations on the receptor camping and protein-protein connections. For the meantime, the consequence results in the variation within a Gibbs free energy ($\Delta\Delta G$) of binding and camping for each area and edge diminished through SNP. The mCSM-PPI2 server is a user-based friendly tool which integrates the combined and inclusive computer prototypical to examine protein-protein-affinity impacts of missense mutations.

PDBsum for secondary structure evaluation and POLG 2 gene interactions

The PDBsum is an illustrative databank that comprises an overview of the natural surroundings of each three-dimensional framework deposited in PDB. It displays the substances which create a framework and their representations of interaction. For a better considerate of gene's function, it is significant to have an idea to its particular interacting partners. Consequently, the STRING database (<http://string-db.org>) was utilized to inspect the genes which communicate with POLG 2.

RESULTS AND DISCUSSION

Human POLG 2 comprises a total of 5828 SNPs. Out of that, only 341 were found to be non-synonymous, 4860 were existed in intron regions and 143 were lay out in synonymous region whereas 3 were found to be pathogenic.

Evaluating the nature of 3 nsSNPs

SIFT server was utilized to predict the given nsSNPs are deleterious (≤ 0.05) or not. A total of 3 pathogenic nsSNPs were given for SIFT analysis^[14-16], only one nsSNP like rs104894632 has come with an output as shown in Table 1. The identified nsSNP is exhibited a SIFT sore of 0.514 in a mutation on G451E gene^[17-19].

Prediction the disorder of SNPs

The filtered and identified polymorphism was carried forward to validate the disease related nsSNPs^[20-21]. The reclaimed rs104894632 nsSNP was subjected to PolyPhen-2 and SNAP2 prediction analysis is shown in Figure 1. The PolyPhen-2 indexing that was depends on the

operational idea which prophesied as probably damaging within a prediction value of 1.

PolyPhen-2 (Polymorphism Phenotyping v2) is a tool which predicts possible impact of an amino acid substitution on the structure and function of a human protein using straightforward physical and comparative considerations. Please, use the form below to submit your query.

Query Data																																											
Protein or SNP identifier	Q9UHN1																																										
Protein sequence in FASTA format																																											
Position	451																																										
Substitution	<table border="1"> <tr> <td>AA₁</td> <td>A</td><td>R</td><td>N</td><td>D</td><td>C</td><td>E</td><td>Q</td><td>G</td><td>H</td><td>I</td><td>L</td><td>K</td><td>M</td><td>F</td><td>P</td><td>S</td><td>T</td><td>W</td><td>Y</td><td>V</td> </tr> <tr> <td>AA₂</td> <td>A</td><td>R</td><td>N</td><td>D</td><td>C</td><td>E</td><td>Q</td><td>G</td><td>H</td><td>I</td><td>L</td><td>K</td><td>M</td><td>F</td><td>P</td><td>S</td><td>T</td><td>W</td><td>Y</td><td>V</td> </tr> </table>	AA ₁	A	R	N	D	C	E	Q	G	H	I	L	K	M	F	P	S	T	W	Y	V	AA ₂	A	R	N	D	C	E	Q	G	H	I	L	K	M	F	P	S	T	W	Y	V
AA ₁	A	R	N	D	C	E	Q	G	H	I	L	K	M	F	P	S	T	W	Y	V																							
AA ₂	A	R	N	D	C	E	Q	G	H	I	L	K	M	F	P	S	T	W	Y	V																							
Query description	mutation																																										
Submit Query Clear Check Status																																											
Display advanced query options																																											

Figure 1: Query input utilizing the PolyPhen-2 prediction.

The SNAP2 server further confirms that these nsSNP produced the damaged effect to the body. The calculated results for the study of SIFT, PolyPhen-2 and SNAP2 were enlisted in Table 1.

Table 1: Result of SIFT, PolyPhen2, and SNAP2 with their corresponding score.

dsSNP ID	Variants	Alleles	SIFT		PolyPhen-2		SNAP2
			Effect	Score	prediction	score	prediction
rs1048	G451	C/T	Deleterious	0.514	Probably damaging	1	Effect

Furthermore, these nsSNP was taken forward for PhD-SNP (Figure 2), SNPs&GO, PMut and PROVEAN prediction to assess the functionality of deleterious nsSNP and envisage of disorder associated mutations nsSNP (Table 2) respectively.

Table 2: Result of the disorder linked SNPs forecast from PhD-SNP, SNPs & GO, PMut and PROVEAN servers.

S. No	Variants	Alleles	PhD-SNP		SNPs&GO		PMut	PROVEAN
			Prediction	R	Prediction	R	Prediction	Prediction
1	G451E	C/T	Disease	3	Disease	6	Disease, 0.81	Deleterious

Figure 2: Prediction of deleterious SNP using PhD-SNP algorithm.

Stability evaluation impact POLG 2 Interface or Core and silico solvent accessibility

Forecasting the structural framework of proteins, ELASPIC consider into account of homology-based study and plotting nsSNP to a multifaceted domain structure [22]. Consequently, it envisages when the mutation cascades to the surface area or core region of the area. The mutant nsSNP G451E was exist in the core arena which can impact the constancy of a receptor structure; exemplified in Table 3. The Table 4 unveils the ΔG_{wt} , ΔG_{mut} and $\Delta\Delta G$ with a value of 155.774, 158.589 and -0.308, respectively.

Table 3: Mutation effect predictions on target protein using ELASPIC

S. No.	Variants	Alleles	ELASPIC	ΔG_{wt}	ΔG_{mut}	$\Delta\Delta G$
--------	----------	---------	---------	-----------------	------------------	------------------

rs104894	G451	C/T	Core	155.7	158.58	-0.308
632	E			74	9	

Prediction of mutant impact on POLG 2 affinity

The catalytic site residues for the POLG 2 protein was prophesied through mCSM-PPI2 which exposed the decrease affinity requisite with $\Delta\Delta G$ of protein-protein interface as accessible in Figure 3 and Table 4. G451E have exhibited a $\Delta\Delta G$ affinity value of -0.128 kcal/mol, whereas it also identifies the distance to interface value as 34.27 Å, respectively.

Table 4: Effect of mutation on binding affinity of POLG2 predicted by mCSM-PPI2.

S. No.	Variants	Distance to Interface (Å)	Predicted $\Delta\Delta G$ Affinity (kcal/mol)	Affinity
1.	G451E	34.27	-0.128	Decrease

Overall, from all the computational analysis ^[23], it was identified that the screened nsSNP that is rs104894632 were projected to be disease related in the POLG 2 protein because of the presence of mutation in the glycine to glutamic acid in the position of 451.

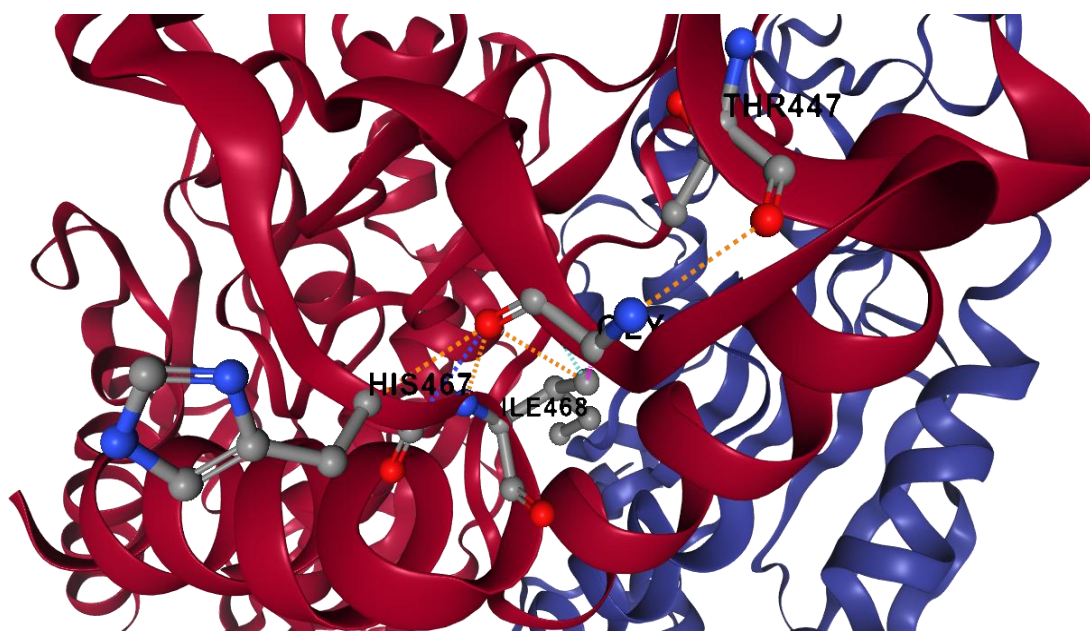


Figure 3: Binding site interaction analysis for POLG 2 prophesied by mCSM-PPI2.

Examination of the interactions of POLG 2 gene and its presence in the STRING database networks

In order to understand the POLG 2 gene's function, it is essential to study the deeper understanding of their interactions with other particular binding partners. Consequently, the STRING database was utilized to scrutinize the genes which interact with the POLG 2 protein. The database of STRING does indispensable assessment and incorporation of both straight (physical) and subsidiary (functional) relations to that of various protein-protein interactions. POLG 2 has direct interfaces with POLG, C10orf2, GABPA, IFIT2, EPRS gene, etc. respectively as demonstrated in Figure 4.

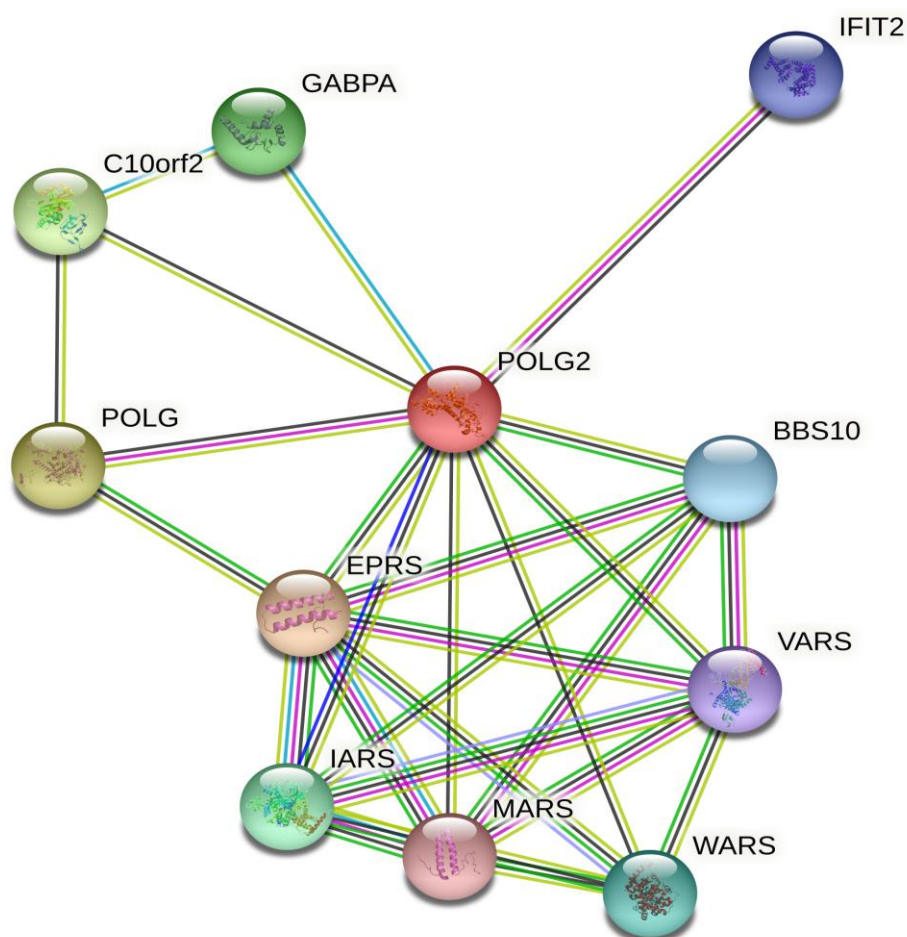


Figure 4: POLG 2 functionally interrelates with other related genes by STRING database.

CONCLUSION

The disease caused POLG 2 gene was explored in our study by assessing the impact of functional SNPs by various computational tools. Amongst, all the 5828 SNPs within a POLG

2 gene, 341 was originated as non-synonymous and 3 were found to possess pathogenic. Initially, we employed 3 nsSNPs namely, rs104894632, rs397514659 and rs886037843 to arrange the damaging nsSNPs and enhance the precision of the study. Out of 3, only one nsSNP that is rs104894632 where found to exhibits a more deleterious amongst other two nsSNPs ^[24]. These predicted nsSNP was both diseases related and uncontrolled showing substantial roles in instigating diverse neurodegenerative disorders ^[25]. These nsSNP was also convoluted in disturbing the stable and functional nature of the POLG 2 protein.

REFERENCES

1. Shameem, M. M., Sasikanth, S. M., Annamalai, R., & Raman, R. G. (2021). A brief review on polymer nanocomposites and its applications. *Materials Today: Proceedings*.
2. Sasikanth, S. M., & .R, G. R. (2019). A Brief Review on Synthesis of Metal Oxide Based Nano-composites and their Photocatalytic Applications. *International Journal of Advanced Science and Technology*, 28(7), 118 - 123.
3. J. Barroso, M. Gurnell, V.E. Croeley, M. Agostini, J.W. Schwabe, M.A. Soos, et al. Dominant negative mutations in human PPAR gamma associated with severe insulin resistance diabetes mellitus and hypertension, *Nature* 402 (1999) 880–883.
4. R. Thomas, R. McConnell, J. Whittacker, P. Kirkpatrick, J. Bradley, R. Stanford, Identification of mutations in the repeated part of the autosomal dominant polycystic kidney disease type 1 gene PKD1 by long range PCR, *Am. J. Hum. Genet.* 65 (1999) 39–49.
5. T.P. Dryja, T.L. McGee, L.B. Halu, G.S. Conley, J.E. Olsson, E. Reichel, et al., Mutations within the rhodopsin gene in patients with autosomal dominant retinitis pigmentosa, *N. Engl. J. Med.* 323 (1990) 1302–1307.
6. E.P. Smith, J. Boyd, G.R. Frank, M. Takahashi, R.M. Cohen, B. Specker, et al., Estrogen resistance caused by a mutation in the estrogen-receptor gene in a man, *N. Engl. J. Med.* 331 (1994) 1056–1061.
7. Y. Miki, J. Swensen, D. Shattuck-Eidens, et al., A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1, *Science* 266 (1994) 66–71.
8. Spinazzola A, Zeviani M: Disorders from perturbations of nuclear mitochondrial inter-genomic cross-talk. *J Intern Med* 2009, 265:174–192.
9. Copeland WC: Defects in mitochondrial DNA replication and human disease. *Crit Rev Biochem Mol Biol* 2012, 47:64–74.
10. Suomalainen A, Isohanni P: Mitochondrial DNA depletion syndromes—many genes, common mechanisms. *Neuromuscul Disord* 2010, 20:429–437.
11. Sasikanth, S. M. (2021). Synthesis And Characterization Of Eu₂WO₆ By Hydrothermal Method. *Turkish Journal of Computer and Mathematics Education (TURCOMAT)*, 12(6), 188-192.

12. Winterthun S, Ferrari G, He L, Taylor RW, Zeviani M, Turnbull DM, Engelsen BA, Moen G, Bindoff LA: Autosomal recessive mitochondrial ataxic syndrome due to mitochondrial polymerase gamma mutations. *Neurology* 2005, 64:1204–1208.
13. Saneto RP, Cohen BH, Copeland WC, Naviaux RK: Alpers-Huttenlocher syndrome. *Pediatr Neurol* 2013, 48:167–178.
14. Wong LJ, Naviaux RK, Brunetti-Pierri N, Zhang Q, Schmitt ES, Truong C, Milone M, Cohen BH, Wical B, Ganesh J, Basinger AA. Molecular and clinical genetics of mitochondrial diseases due to POLG mutations. *Human mutation*. 2008 Sep;29(9):E150-72.
15. Stumpf JD, Copeland WC. Mitochondrial DNA replication and disease: insights from DNA polymerase γ mutations. *Cellular and Molecular Life Sciences*. 2011 Jan 1;68(2):219-33.
16. Hakonen AH, Davidzon G, Salemi R, Bindoff LA, Van Goethem G, DiMauro S, Thorburn DR, Suomalainen A. Abundance of the POLG disease mutations in Europe, Australia, New Zealand, and the United States explained by single ancient European founders. *European journal of human genetics*. 2007 Jul;15(7):779-83.
17. Ait El Cadi C, Krami AM, Charoute H, Elkarhat Z, Sifeddine N, Lakhiari H, Rouba H, Barakat A, Nahili H. Prediction of the Impact of Deleterious Nonsynonymous Single Nucleotide Polymorphisms on the Human RRM2B Gene: A Molecular Modeling Study. *BioMed research international*. 2020 Jul 26;2020.
18. Subbiah HV, Babu PR, Subbiah U. In silico analysis of non-synonymous single nucleotide polymorphisms of human DEFB1 gene. *Egyptian Journal of Medical Human Genetics*. 2020 Dec;21(1):1-9.
19. Vaser R, Adusumalli S, Leng SN, et al (2016) SIFT missense predictions for genomes. *Nat Protoc* 11:1–9.
20. Adzhubei IA, Schmidt S, Peshkin L, et al (2010) A method and server for predicting damaging missense mutations. *Nat. Methods* 7:248–249.
21. Hecht M, Bromberg Y, Rost B (2015) Better prediction of functional effects for sequence variants. *BMC Genomics* 16.
22. Choi Y, Sims GE, Murphy S, et al (2012) Predicting the Functional Effect of Amino Acid Substitutions and Indels. *PLoS One* 7:e46688.
23. Ferrer-Costa, C., Gelpí, J. L., Zamakola, L., Parraga, I., de la Cruz, X. & Orozco, M. PMUT: a web-based tool for the annotation of pathological mutations on proteins. *Bioinformatics* 21, 3176-3178.
24. Cheng J, Randall A, Baldi P (2006) Prediction of protein stability changes for single site mutations using support vector machines. *Proteins Struct Funct Genet* 62:1125– 1132.
25. Rodrigues CHM, Myung Y, Pires DEV, Ascher DB (2019) MCSM-PPI2: predicting the effects of mutations on protein-protein interactions. *Nucleic Acids Res* 47:W338– W344.