

Effect of Cyclodextrins Inclusion Complex and Gelling Agents on the Release of Ciprofloxacin HCL from Topical Gel Formulations

Wisam Mohammed Naem^{1,2*}, Wedad k. Ali ², Suhad Faisal Hatem Al-Mugdadi ³

¹ Kerbala Health Directorate -Ministry of Health, Iraq.

² Department of Pharmaceutics, College of Pharmacy, University of Mustansiriyah, Baghdad, Iraq.

³ Department of Clinical Laboratory Sciences, College of Pharmacy, University of Mustansiriyah, Baghdad, Iraq.

*Corresponding Author

Email:khursaniwisam@gmail.com

Abstract

Background: Topically applied drugs have the advantage of supplying a higher concentration of the drug to the skin than systemically applied medications. The aim of this study was to develop and evaluate a gel formulation of ciprofloxacin hydrochloride (CP-HCL), also to provide a topical treatment for various bacterial skin infections.

Materials and methods: In order to improve the dissolution properties of CP-HCL, the physicochemical properties of CP-HCL:cyclodextrins (CDs) binary systems were studied in both solution and solid states after preparation of the complexes by two methods: simply by physical mixture and kneading methods. CP-HCL:CDs physical mixture and kneaded complexes were prepared and characterized using FTIR, DSC and SEM comparing with CP-HCL as received. Different gel bases and concentrations of hydroxypropylmethylcellulose (HPMC), sodium carboxymethylcellulose (Na CMC), and sodium alginate (Na Alginate) were used to formulate CP-HCL and its CDs complexes.

Results: The results showed that inclusion complexation occurred for both kneaded and physically mixed CP-HCL with CDs. Almost all of the 19 formulations produced had suitable physicochemical properties, with formula F9, which included a CP-HCL:BCD physical mixture complex and a base consisting of a 10% w/w HPMC gel formulation, providing a good viscosity and drug release properties. Color, consistency, pH, rheological and kinetic properties, skin permeability, and drug release pattern were among the parameters examined. Furthermore, the antibacterial activity of CP-HCL and CP-HCL:CDs complexes, as well as gel formula containing 10% w/w HPMC, was investigated.

Conclusion: Ciprofloxacin and its complexes were successfully incorporated into the various topical gel preparations which showed promising results for the treatment of skin infections.

Key words: Cyclodextrin, Ciprofloxacin, HPMC, Na CMC, Na Alginate, Inclusion complexation.

INTRODUCTION

Topical drug delivery is a successful way for treating of local conditions or penetrate through the skin's underlying layers, allowing for better absorption and hence the desired pharmacological systemic effect when other routes are ineffective (1).

Dealing with drugs that have limited water solubility, the formulation of suitable dosage forms for oral, parenteral, and topical routes is frequently an issue to be administered in large volume. Commonly, to improve the solubility of poorly-water soluble drugs different methods are used such as prodrug formation, use of non-aqueous co-solvents such as glycols or ethanol, drug solubilization in micelles using surfactants, pH adjustment and complex formation (2) (3).

Interaction of two or more molecules to form a non-bonded system with a well-defined stoichiometry is known as complexation (4). Complexation involves relatively weak forces such as hydrogen bonding, London forces, and hydrophobic interactions. Since there are no involved covalent bonds between them, these are also known as no-bond complexes (5). The formation of host-guest inclusion complexes by weak intermolecular interaction, known as cyclodextrin (CD) inclusion complexation, has been found to be a promising technique for improving the solubility, bioavailability as well as stability of drugs that are poorly water-soluble (6) (7).

Gel for topical application is the most favorable dosage form and could be formulated using gelling agents such as sodium carboxymethylcellulose, hydroxypropylmethylcellulose, and sodium alginate (8).

Ciprofloxacin hydrochloride (CP-HCL) is a broad spectrum antibiotic, it acts against most bacterial pathogens that cause respiratory, urinary tract, gastrointestinal and skin infections (9). As a salt, CP-HCL is only sparingly soluble in water, very slightly soluble in ethanol, slightly soluble in methanol, and practically insoluble in acetone, ethyl acetate, and methylene

chloride (10). Like other fluoroquinolone compounds, CP-HCL has a U-shaped pH-solubility profile, with high solubility at pH values below 5 and above 10, and low solubility near its isoelectric point (11) (12). Ciprofloxacin to be BCS Class IV.(13) It is a zwitter ion molecule containing two proton-binding sites values of pKa1 and pKa2 are 6.2 and 8.59, respectively (14).

This study aimed to develop and evaluate a gel formulation of ciprofloxacin hydrochloride (CP-HCL), to be used topically for treatment of various bacterial skin infections.

Materials: β -cyclodextrin and HP- β -cyclodextrin were purchased from Sigma Chemical Company (st. louis.mo), Ciprofloxacin HCl was obtained from PiONEER Co. for Pharmaceutical Industries, Hydroxyl propyl methyl cellulose K15M (HPMC) was purchased from Alpha Chemika, India, Sodium alginate was obtained from PiONEER Co. for Pharmaceutical Industries, and Sodium carboxy cellulose (SCMC) was purchased from Jiangsu yew al co., ltd (china).

Methods (Experimental Work)

Determination of Ciprofloxacin HCl Melting point

A small amount of the drug powder was placed in a capillary tube to calculate the melting point. The tube was flame-sealed on one side before being placed in Stuart's electric melting point (15).

Preparation of Solid Samples:

The following methods were used to make various CP-HCL/CDs formulations in a 1:1 molar ratio: **(a) Physical mixing method (Phy. Mix. Meth.)** (16): By which CP-HCL and CD were placed in a screw-cap glass vial and mixed for 30 minutes to ensure consistent mixing. **(b) Kneading method (Kn. Meth.):** In this method CDs and CP-HCL were mixed in a mortar and kneaded with the aid of about 1 ml deionized water for 15 minutes, then dried in an oven for 24 hours at 50°C. The resulting dry solid mass was finely powdered, passed through 60 mesh sieve, and placed in a sealed glass vial until used.

Phase Solubility Study:

In a series of 25ml conical flasks, excess amount of CP-HCL (100 mg) were introduced to either a buffer or β -CD aqueous solutions. The mixtures were shaken for 48 hours at (28°C) in a water bath shaker to reach equilibrium, then 2ml aliquots were taken every 12 hours and filtered via a 0.45 μ m syringe filter for dilution and CP-HCL concentration was determined by calculating absorbance at its own λ_{max} value (17, 18).

Fourier Transform Infrared Spectroscopy (FTIR)

Samples of CP-HCl, β CD, HP- β CD, physical and kneading complex powders; each sample was pressed in a disk after being combined with potassium bromide. Shimatzu FTIR spectroscopy was used to study the disc (4000-400) cm^{-1} (19) (20).

Differential scanning calorimetry (DSC):

With a linseis DSC (STA PT-1000 linseis, Germany), the thermal habits of pure CP-HCL, BCD, HP-BCD, physical, and kneading complex powders were investigated, samples were loaded separately into aluminum crucibles pans and temperature was allowed to raise from 25 to 400 °C at a rate of 10 °C/min under an argon atmosphere (inert conditions) (21, 22).

Scanning electron microscopy (SEM)

The surface morphology of pure CP-HCL, β CD, HP- β CD as well as their physically mixed and kneaded inclusion complexes were examined by a INSPECT F50 scanning electron microscope (FEI, HOLLAND) (23) (24).

Preparation of Gel Formulas

Table (1) shows different gel formulations of Phy. Mix. Meth. and Kn.Meth. products using the following gelling agents:

Sodium alginate gel

The alginate was dissolved in distilled water and heated to 60°C while stirring. 0.5% w/w CP-HCL or (Phy. Mix. Meth. complex) and (Kn. Meth. complex) were dissolved in the resulting gel after cooling to below 40°C (25).

Hydroxypropylmethylcellulose (HPMC) gel

HPMC was dispersed in hot water (80°C) to make gels. After cooling the dispersions to 25°C, 0.5 % w/w CP-HCL or equal weight in (Phy. Mix. Meth.) or (Kn. Meth.) were added, stored at 4°C for the next 24 hours to allow maximum swelling of the polymer and produce homogenized systems (26).

Sodium carboxymethylcellulose (Na CMC) gel

In a glass mortar, Na CMC was blended with glycerin, and a clear gel was created by adding small amounts of the mixture to a solution of previously warmed deionized water while stirring. This method was used to make the Na CMC gel, and then 0.5 % w/w CP-HCL or equivalent weight of (Phy. Mix. Meth.) or (Kn. Meth.) were applied using a mortar and pestle (27).

Evaluation of the Ciprofloxacin gel

Physical Properties of the gel

The color of the prepared gel formulations was visually evaluated. Homogeneity, consistency, pH, and viscosity were also determined.(28)

Measurement of pH

A gel solution was made by dissolving 1 gram of the gel in 100 mL deionized water and allowing it to sit for 2 hours. After that, a digital pH meter was used to assess the pH of the prepared gel solution at 18 °C. (29)

Viscosity Measurements

A Brookfield viscometer was used to assess the viscosity of various gel formulations at 37°C (Brookfield DV-E Viscometer). The samples were rotated at 3, 5,10,20,30, 50, and 100 rpm using spindles S63 and S64, and the viscosities were measured with a 30 second interval between each rate. (29)

Table (1) The prepared gel formulas showing the main active and the complexation method.

Formula no.	CP-HCL g%	Sodium Alginate g%	HPMC g%	SCMC g%	Deionized water QS Up to 100 g	Type of drug added
F1	0.5	5			100	Kn. Meth. complex
F2	0.5	5			100	Phy. Mix. complex
F3	0.5	5			100	CP-HCL alone
F4	0.5	10			100	Kn. Meth. complex
F5	0.5	10			100	Phy. Mix. complex
F6	0.5	10			100	CP-HCL alone
F7	0.5		10		100	Kn. Meth. complex
F8	0.5		10		100	CP-HCL alone
F9	0.5		10		100	Phy. Mix. complex
F10	0.5		13		100	CP-HCL alone
F11	0.5		13		100	Phy. Mix. complex
F12	0.5		13		100	Kn. Meth. complex
F13	0.5			4	100	Phy. Mix. complex
F14	0.5			4	100	Kn. Meth. complex
F15	0.5			4	100	CP-HCL alone
F16	0.5			6	100	CP-HCL alone
F17	0.5			6	100	Phy. Mix. complex
F18	0.5			6	100	Kn. Meth. complex
F19	0.5		10		100	Phy. Mix. complex With HP-BCD

In Vitro Release Test

A small funnel with a diameter of 3 cm was adjusted to accommodate 2 gm of each formula containing an equivalent weight of 0.5 % w/w of CP-HCL.

A cellulose membrane was sealed in place with a rubber band over the funnel's mouth. In a 500 mL beaker of the dissolution apparatus, the modified dialysis cell was inverted in phosphate citrate buffer pH 5.5.

After 1, 2, 3, 4, 5, and 12 hours, the buffer solution was pipetted from the collecting medium and replaced with an equivalent amount of fresh buffer solution and the device maintained at 37°C(30). The samples were spectrophotometrically analyzed for CP-HCL content at its λ_{max} .

Effect of Different Polymers and β -cyclodextrin as Complexing Agent on the Release of CP-HCL in Phosphate citrate Buffer pH 5.5 at 37°C

In order to choose the appropriate polymer for gel formulation, different polymers (Sod. alginate, HPMC and Na.CMC) were used with CP-HCL or (Phy. Mix.) and (Kn.) complexes by investigating its effect on the release process.

Effect of Different Concentration of Polymers on the Release of CP-HCL in Phosphate Citrate Buffer pH 5.5 at 37°C

In order to select the best polymer concentration for formulating of the gel, different concentrations of sodium alginate, HPMC, and Na CMC were used with CP-HCL or (Phy. Mix. Meth.) and (Kn. Meth.) through studying its effect on the release process.

Determinations of the Drug release kinetics and mechanisms

Kinetics of Drug Release

To characterize drug release kinetics and propose a mechanism of drug release, the cumulative amount of (CP-HCL) released from the selected formulas at successive time intervals was fitted to zero order, first order kinetics, Higuchi, and Korsmeyer–Peppas models (31, 32).

Anti-bacterial activity test (33)

Some of the solid formulas and optimal gel formula (9 samples) were clinically tested against different strains of bacteria, then plated on agar media at 37°C for 18-24h. Clinical samples include gram positive bacteria *Streptococcus spp.* and gram negative bacteria *Klebsilla spp.* 100 μ l of inoculums (10^8 CFU/ml; 0.5 Mac Farland standard) of each type of bacteria was spread on Mueller Hinton Agar (MHA) plate using cotton swab. The diameter of three wells was punched (6mm). Into each well, 100 microliters of the prepared compounds or formulas in concentration (100 μ g of CP-HCL in each different formula) were used. CP-

HCL was used as a standard positive antimicrobial and the buffer solution was used as a negative control (34).

Statistical analysis

The experimental data were presented as mean \pm of standard deviation (SD). Three-way ANOVA that include Bonferroni test (pairwise comparisons) was also used for statistical analysis at 95%: significance ($p < 0.05$) and ($p > 0.05$) for non-significance. Statistical calculation was done using Statistical Package for the Social Sciences (SPSS) 25.0 software.

Results and Discussion:

Ciprofloxacin HCL Characterization / Melting Point

The melting point of (CP-HCl) was determined to be 317 °C. This finding is consistent with that stated in references, indicating that the drug powder used in the analysis was pure.(15, 35)

Phase Solubility Study

The total solubility of CP-HCL versus total concentration of β CD is shown in table (2), while Fig. (1) Summarizes the findings of the phase solubility analysis at 28 °C, which were determined by plotting CP-HCL apparent equilibrium concentrations against β CD concentrations. The apparent solubility of CP-HCL at 28 °C increased linearly as a function of β CD concentration, and this linearity was confirmed by these results indicating the formation of water soluble complexes in the solution. The size of the complex stoichiometry was assumed to be 1:1 because the slope of the diagram was less than unity, (0.0029). The apparent stability constant was discovered to be 219 M⁻¹. (17) and the solubility of CP-HCL increased 1.32 fold.

The apparent stability constant (K1:1) for these complexes was determined using phase solubility diagrams and the equation below (36):

$K1:1 = \text{slope} / S_0$ (1- slope),

Where, S_0 is intercept, CP-HCL solubility at zero β -CD concentration.

Table (2) Total Solubility of CP-HCL against Total Concentration of β CD

Concentration* 10^{-3} M of β CD	Concentration * 10^{-5} M of CP-HCL
0	0.133
3.5	0.145
7	0.153
10.5	0.161
14	0.176

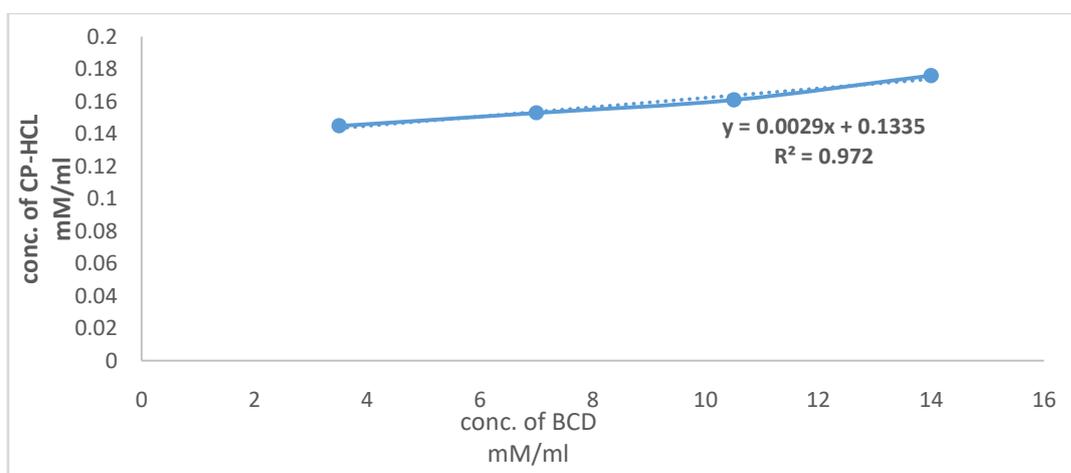


Figure (1) The diagram of Phase solubility for CP-HCL:CD solution.

Study of Fourier Transform Infrared Spectroscopy (FTIR) for solid samples:

The infrared spectra of CP-HCL, β CD, HP- β CD (physical mixture and kneaded samples of CP-HCL with β CD and HP- β CD) samples-CDS complexes are shown in (Fig 2). The absorption intensity of the CN group appearing in 1626 cm^{-1} was shifted (showed a red shift), and the absorption intensities of CN in all inclusion complexes were weaker than on CP-HCL alone, so it can be assumed that CN from CP-HCL was incorporated into the cavity of CDs (20).

Whereas the spectrum of β CD was characterized by bands at 3363 cm^{-1} of O-H stretching and a broad band due to H-bonding. the spectrum of the complex showed no shift to the band corresponding to OH stretching vibration. (15, 37).

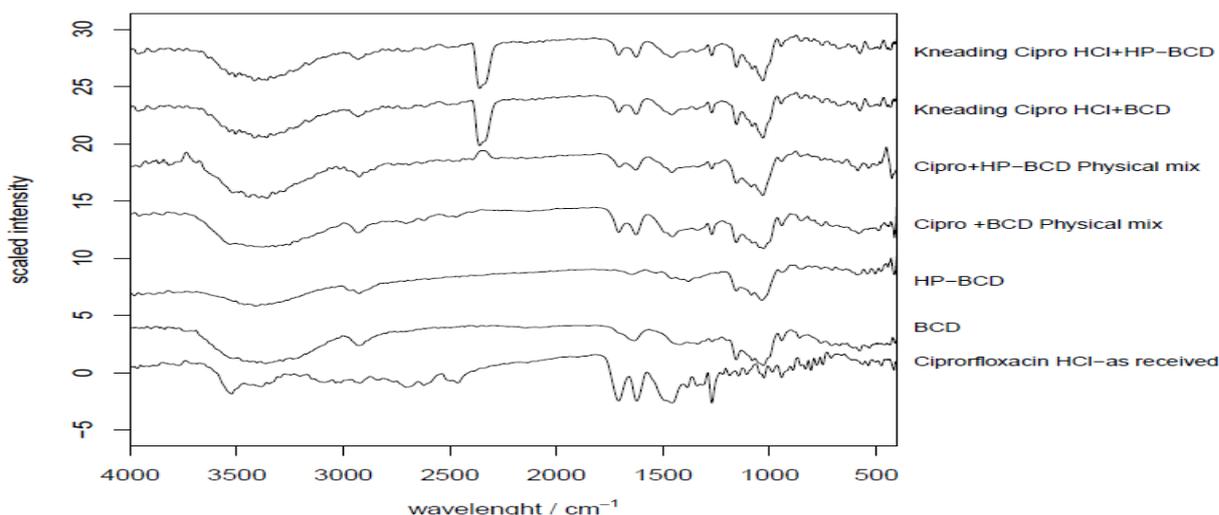


Figure (2) The IR spectra for all solid samples and the prepared solid complexes together by R studio drawing.

Differential Scanning Calorimetry

The DSC results showed endothermic peaks for CP-HCL, β CD, and HP- β CD at 154.1 °C, 113.9 °C, and 91.6 °C, respectively. The disappearance of CP-HCL, β CD, and HP- β CD endothermic peaks, as well as the appearance of another endothermic peak in the range started from 102.3°C to 149.8°C, could indicate the formation of an inclusion complex between CP-HCL and CDs. The melting points of CP-HCL and inclusion complexes with β CD and HP- β CD were determined to be 317 °C, and 259 °C, 252 °C, respectively (38). As showed in figure (3) below:

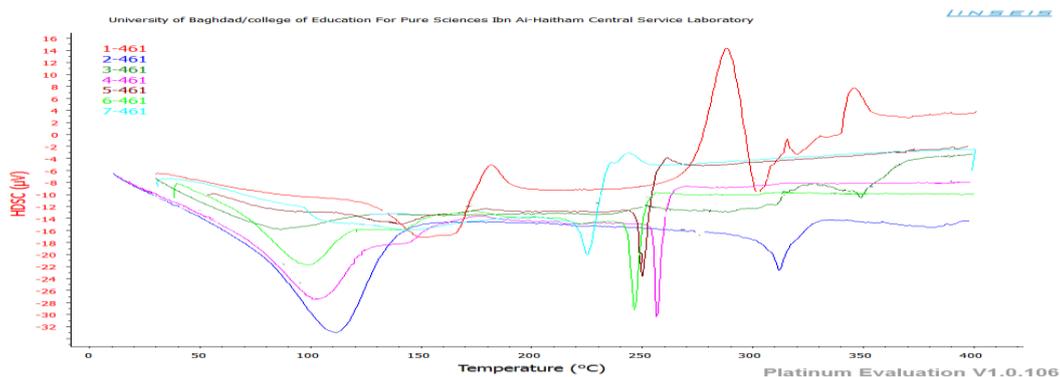


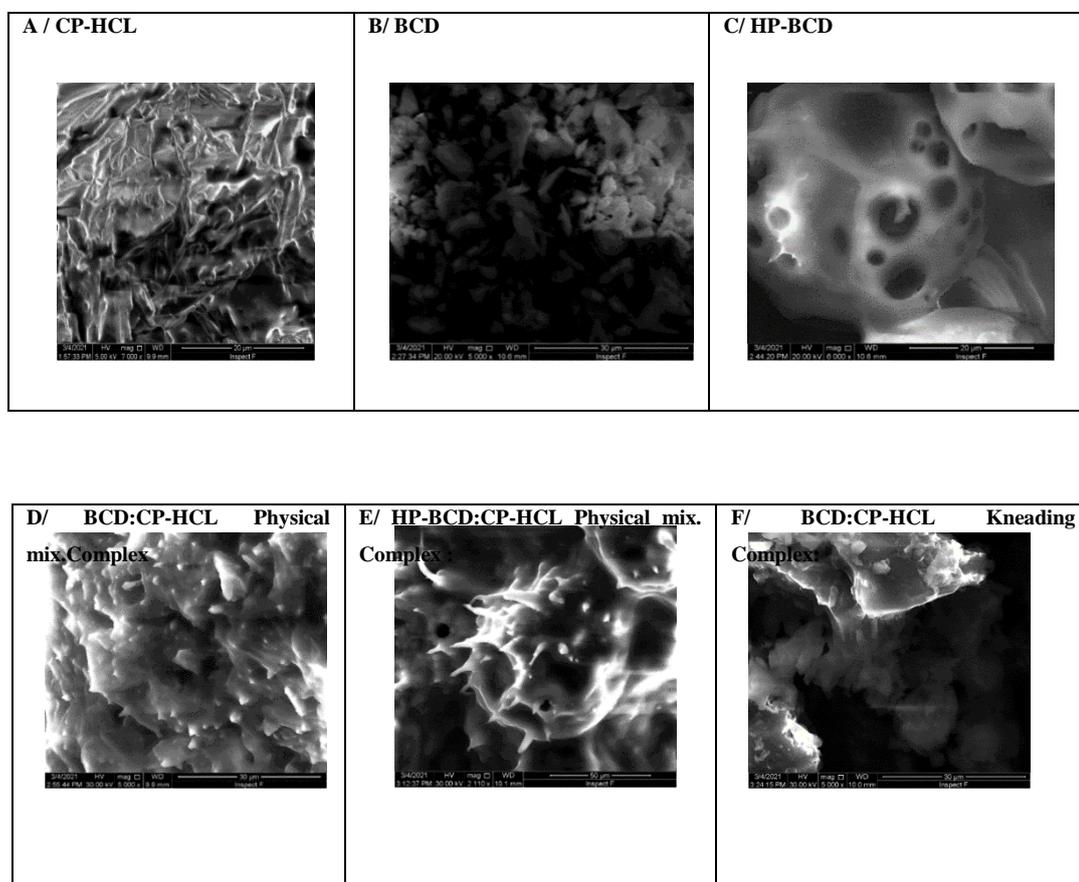
Figure (3) The DSC results for CP-HCL, β CD, and HP- β CD and their physical mixture and kneaded complexes showing the endothermic peaks plotted together.

Scanning electron microscopy (SEM) Results:

Figure (4) shows the existence of a crystalline particle of uniform size distinguishes pure CP-HCL. On the other hand, both BCD and HP-BCD appear as crystalline particles with no definable form.

The crystalline structure of both CP-HCL and CDs was visible in the physical inclusion complexes of CP-HCL:CDs. CP-HCL crystals mixed with CDs crystals were observed adhering to their surfaces or within their cavities. For kneaded samples showed the normal morphology of preparations made with this system in general. Where, small particles with a tendency for aggregation, indicating the presence of an amorphous substance with single component in the complex, implying maximum complex formation (17).

Although the SEM technique is incapable of concluding true complex formation, the micrographs obtained support the concept of creating a new specific component. So, in the case of CP-HCL CDs 1:1 M kneaded and physical mixture binary systems, the inclusion complexation can be confirmed using DSC and FTIR data together with this analysis (39).



F6	Na Alginate 10%	CP.HCL Alone	Excellent	Slightly Clear brawn	7.63±0.092
F7	HPMC 10%	BCD/Kneading	Excellent	Clear yellowish	7.83±0.269
F8	HPMC 10%	CP.HCL Alone	Excellent	Clear yellowish	5.37±0.318
F9	HPMC 10%	BCD/Physical	Excellent	Clear yellowish	5.82±0.088
F10	HPMC 13%	CP.HCL Alone	Excellent	Clear yellowish	5.15±0.136
F11	HPMC 13%	BCD/Physical	Excellent	Clear yellowish	5.12±0.235
F12	HPMC 13%	BCD/Kneading	Excellent	Clear yellowish	5.95±0.075
F13	Na CMC 4%	BCD/Physical	Excellent	Clear white	6.42±0.065
F14	Na CMC 4%	BCD/Kneading	Excellent	Clear white	6.16±0.102
F15	Na CMC 4%	CP.HCL Alone	Excellent	Clear white	6.5±0.081
F16	Na CMC 6%	CP.HCL Alone	Excellent	Clear white	6.62±0.065
F17	Na CMC 6%	BCD/Physical	Excellent	Clear white	6.71±0.237
F18	Na CMC 6%	BCD/Kneading	Excellent	Clear white	6.22±0.080
F19	HPMC 10%	HP-BCD/Kneading	Excellent	Clear yellowish	5.68±0.260

Viscosity Measurements

The viscosity measurement profiles revealed that as the share stress increased, normally arranged molecules aligned their long axes in the direction of flow orientation, lowering the material's internal resistance and decreasing viscosity (42). The findings shows that the viscosity for each kind of polymer increased as the polymer concentration increased. According to many literatures, they have a highly acceptable rheological profile range of 3999 -120000 cps⁰ (43).

Formulas with HPMC and Na alginate polymers at the highest concentration of the polymer (**group A**) rotated with spindle S63 and the resultant viscosities at different rates were the same for all these formulas; 23990, 12000, and 5999 cps at 5, 10, and 20 speed rotation respectively. Formulas with HPMC and Na alginate polymers at the lowest concentration of the polymer (**group B**) rotated with spindle S63 and the resultant viscosities at different rates were the same for all these formulas; 20000, 9998, and 3999 cps at 5, 10, and 20 speed rotation respectively. While formulas with Na CMC polymer at the highest concentration of the polymer (**group C**) rotated with spindle S64 and the resultant viscosities

at different rates were the same for all these formulas; 120000, 59990, and 29990 cps at 5, 10, and 20 speed rotation respectively. Formulas with Na CMC polymer at the lowest concentration of the polymer (**group D**) rotated with spindle S64 and the resultant viscosities at different rates were the same for all these formulas; 99980, 49990, and 20000 cps at 5, 10, and 20 speed rotation respectively. These findings are illustrated in the figure (5)

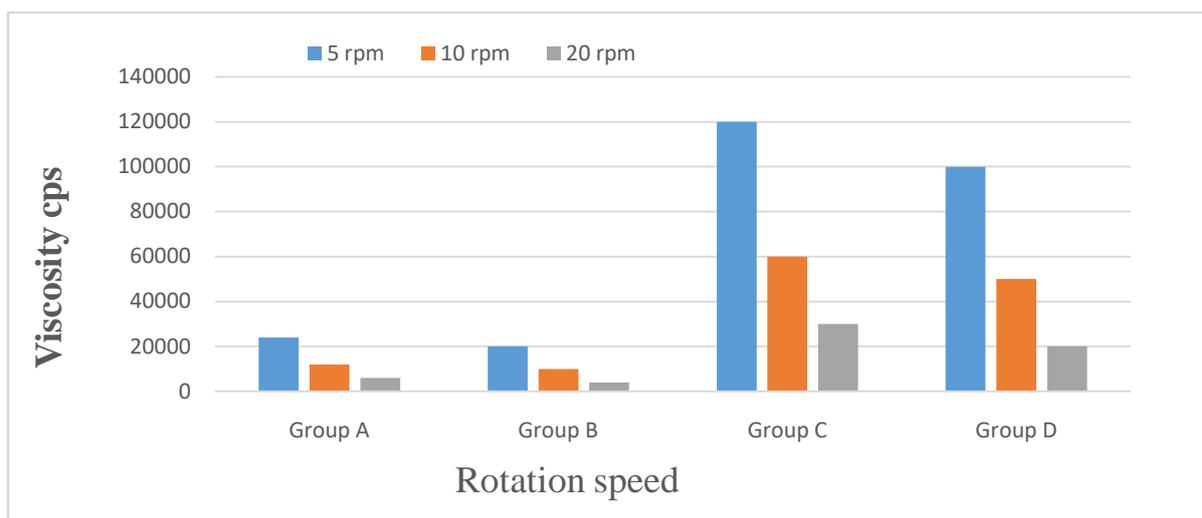


Figure (5) Gel formulas (F1-F19): four group division according to their polymer concentration; viscosity (cps) at 37 °C at different rotation per minute (rpm).

Effect of Different Polymers and Cyclodextrins as Complexing Agent on the Release of CP-HCL in Phosphate Citrate Buffer pH 5.5 at 37 °C

Figure (6) shows the release of CP-HCL as a function of square root of time from various bases containing 0.5 % w/w of the drug to obtain a straight line. According to the data, the release rate constant (k) reduced in the order shown in table (4): sod. Alginate < sod. CMC < HPMC. This could be explained by the fact that CP-HCL is slightly insoluble in water, so drug partitioning is reduced according to the nature of the base; the highly viscous Na CMC base showed a lower release rate than the HPMC gel (44).

When some types of polymers and cyclodextrins were both present, a significant reduction in the cyclodextrin solubilizing efficiency towards the drug was observed, which was attributed to a possible competition effect between the Na alginate and the drug for the interaction with the macrocycle.(45)

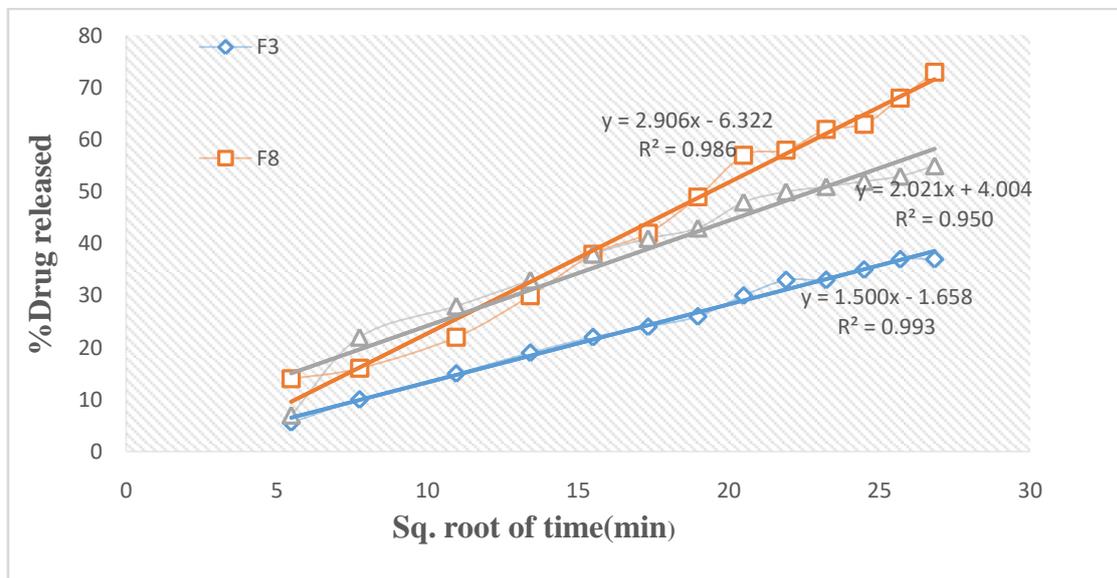


Figure (6) Effect of Different Polymers on the Release of CP-HCL in Phosphate Citrate Buffer pH 5.5 at 37 °C as a function of square root of time in (min)

Table (4) Effect of Different Polymers on the Release Rate Constant (K) of CP-HCL in Phosphate Citrate Buffer pH 5.5 at 37 °C

Type of polymer (base)	K mg.(min.)-1/2	Correlation coefficient (r)
F3 (Na alginate 5%)	1.50	0.9931
F8 (HPMC 10 %)	2.90	0.9868
F15 (Na CMC 4%)	2.02	0.9508

Effect of Different Concentration of Polymers on the Release of CP-HCL in Phosphate Citrate Buffer pH 5.5 at 37 °C

The effect of various HPMC and sodium CMC concentrations on the CP-HCL release rate as a function of square root of time. The release rate constants are listed in table (5). The result showed that the concentration of the polymer had

an effect on the release rate constant (k). Increasing in the polymer concentration a decrease the value of (k).

This result's view is in agreement with previous findings, which indicated that a drug's release rate is determined by the physical structure of the polymer network, when the gel is extremely hydrated (low polymer concentration), the drug diffuses through the pores, while when the gel is dehydrated (high polymer concentration), the drug dissolves or remains in the polymer and is transported between its chains, so the hydrophobicity of the gel is increased by cross-linking, which reduces the drug's release rate (3). However, as the impact of increased viscosity on drug release retardation and formula stability becomes more widely recognized, viscosity becomes more significant (46)

Table (5) Effect of sodium CMC and HPMC concentrations on the release rate constant (k) of CP-HCL in phosphate citrate buffer pH 5.5 at 37 °C).

Type of polymer (base)	K mg.(min.)-1/2	Correlation coefficient (r)
F7 (HPMC 10%) / Kn.meth. complex	3.4405	0.9910
F12 (HPMC 13%) / Kn.meth. complex	2.9556	0.9865
F14 (Na CMC 4%) / Kn.meth. complex	1.6914	0.9401
F18 (Na CMC 6%) / Kn.meth. complex	1.6626	0.9481

Effect of Different types of Cyclodextrins (BCD and HP-BCD) as Complexing Agent on the Release of CP-HCL in Phosphate Citrate Buffer pH 5.5 at 37 °C

According to the the release rate constant (k) of CP-HCL in phosphate citrate buffer pH 5.5 at 37 °C) as shown in table (6), the effect of Different types of CDs as Complexing Agent F9 (with **BCD**) and F19 (with **HP-BCD**); it was found that the F9 was more faster than F19 this may due to **HP-BCD** form

aggregation that restrict drug release. The statistical analysis showed no significant difference was found if different CDs used.

Table (6) Effect of Different types of Cyclodextrins as Complexing Agent F9 (with **BCD**) and F19 (with **HP-BCD**) on the release rate constant (k) of CP-HCL in phosphate citrate buffer pH 5.5 at 37 °C)

Type of Cyclodextrin	K mg.(min.) ^{-1/2}	Correlation coefficient (r)
F9 (HPMC 10%) / Kn.meth. complex with BCD	3.8488	0.9985
F19 (HPMC 10%) / Kn.meth. complex With HP-BCD	2.3916	0.8691

Drug Release Mechanisms and Kinetics (Mathematical Analysis)

In order to understand the mechanism of drug release and the release rate from dosage forms, in-vitro release data was fitted to various mathematical models such as Zero order, First order, Higuchi, and Korsmeyer- Peppas models.

The majority of the formulations employ Higuchi kinetics, while a few others had first order, with the following formulas fitting best to a favorable zero order model: (F4,F5,F8,F9,F10,F13,F17,F18,F19), indicated by high regression value (R²).The highest one is **F9**.

The value of the release exponent (n) in the Korsmeyer-Pappas model determines the release mechanism; all formulas with n values between 0.45 and 0.89 indicate anomalous (non-Fickian) transport, which corresponds to a combination of drug diffusion and matrix erosion drug release mechanism. But F4, n values greater than 0.89 indicate super case II transport, in which polymer relaxation and chain disentanglement are involved in drug release.

Anti-bacterial activity test

The antibacterial activity of all compounds was assessed in this study by measuring the inhibition zone (mm), and the effect of CP-HCL alone as a positive control, as well as formula F9 and various prepared complexes and

polymer composing this formula. It was tested on *Klebsilla* spp., and was found that **F9** the selected formula zone of inhibition is 34 compared with 30 for CP- HCL alone. Also tested on *streptococcus* spp., and was found that **F9** the selected formula zone of inhibition is 35 mm compared with 30mm for CP- HCL alone.

CDs improved antibacterial activity against both studied bacteria for solid complexes and selected formula, according to the findings.(47) The increase in antibacterial activity for all forms of CDs complexes alone or in the gel formula may be due to an increase in the nature of water solubility of the drug molecule, which resulted in greater drug diffusivity into the agar medium, resulting in a greater zone of inhibition diameter for CDs complexes .(48)

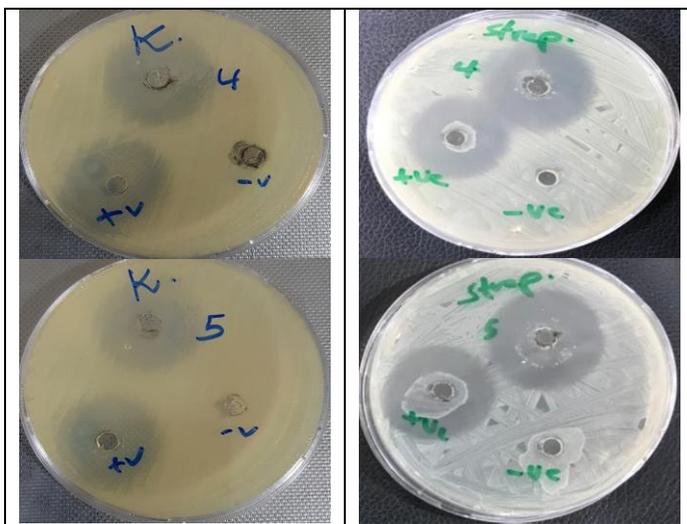


Figure (7) The Zone of Inhibition for the selected formula (F9 containing phy.mix complex) compared with (F8 containing CP-HCL only).

Conclusion:

Nearly all of the 19 formulations produced had suitable physicochemical properties, with formula F9 containing CP-HCL: BCD physical mixture complex and the base consist of a 10% w/w HPMC gel formulation having the best viscosity and drug release properties. Color, consistency, pH, rheological and kinetic properties, skin permeability, antibacterial activity study and drug release pattern are some of the parameters that were evaluated.

Refrence

1. Kaur P, Kaur L, Khan M. Topical formulations and Hydro-gel: An overview. *International Journal of Current Research*. 2013;2(1):201-06.
2. Rasool AA, Hussain AA, Dittert LW. Solubility enhancement of some water-insoluble drugs in the presence of nicotinamide and related compounds. *Journal of pharmaceutical sciences*. 1991;80(4):387-93.
3. Loftsson T. Drug solubilization by complexation. *International journal of pharmaceutics*. 2017;531(1):276-80.
4. Kumar SK, Sushma M, Raju PY. Dissolution enhancement of poorly soluble drugs by using complexation technique-A review. *Journal of Pharmaceutical Sciences and Research*. 2013;5(5):120.
5. Ding J, Li J, Mao S. Development and evaluation of vinpocetine inclusion complex for brain targeting. *asian journal of pharmaceutical sciences*. 2015;10(2):114-20.
6. Yang L-J, Chen W, Ma S-X, Gao Y-T, Huang R, Yan S-J, et al. Host-guest system of taxifolin and native cyclodextrin or its derivative: Preparation, characterization, inclusion mode, and solubilization. *Carbohydrate polymers*. 2011;85(3):629-37.
7. Uekama K. Recent aspects of pharmaceutical application of cyclodextrins. *Journal of inclusion phenomena and macrocyclic chemistry*. 2002;44(1-4):3-7.
8. Allen L, Ansel HC. *Ansel's pharmaceutical dosage forms and drug delivery systems*: Lippincott Williams & Wilkins; 2013.
9. Ball P. Quinolone generations: natural history or natural selection? *Journal of Antimicrobial Chemotherapy*. 2000;46(suppl_3):17-24.
10. Escribano E, Calpena A, Garrigues T, Freixas JLC, Domenech J, Moreno J. Structure-absorption relationships of a series of 6-fluoroquinolones. *Antimicrobial agents and chemotherapy*. 1997;41:1996-2000.
11. Olivera M, Manzo R, Junginger H, Midha K, Shah V, Stavchansky S, et al. Biowaiver monographs for immediate release solid oral dosage forms: Ciprofloxacin hydrochloride. *Journal of pharmaceutical sciences*. 2011;100(1):22-33.
12. Yu X, Zipp GL, Davidson III GR. The effect of temperature and pH on the solubility of quinolone compounds: estimation of heat of fusion. *Pharmaceutical research*. 1994;11(4):522-7.
13. Lindenberg M, Kopp S, Dressman JB. Classification of orally administered drugs on the World Health Organization Model list of Essential Medicines according to the biopharmaceutics classification system. *Eur J Pharm Biopharm*. 2004;58(2):265-78.
14. Sun J, Sakai S, Tauchi Y, Deguchi Y, Chen J, Zhang R, et al. Determination of lipophilicity of two quinolone antibacterials, ciprofloxacin and grepafloxacin, in the protonation equilibrium. *European journal of pharmaceutics and biopharmaceutics*. 2002;54(1):51-8.
15. Moffat AC, Osselton MD, Widdop B, Watts J. *Clarke's analysis of drugs and poisons*: Pharmaceutical press London; 2011.
16. Sathigari S, Chadha G, Lee YP, Wright N, Parsons DL, Rangari VK, et al. Physicochemical characterization of efavirenz-cyclodextrin inclusion complexes. *Aaps Pharmscitech*. 2009;10(1):81-7.
17. Baboota S, Dhaliwal M, Kohli K. Physicochemical characterization, in vitro dissolution behavior, and pharmacodynamic studies of rofecoxib-cyclodextrin inclusion compounds. Preparation and properties of rofecoxib hydroxypropyl β -cyclodextrin inclusion complex: A technical note. *AAPS PharmSciTech*. 2005;6(1):E83-E90.
18. Higuchi T, Connors KA. *Advances in analytical chemistry and instrumentation*. Phase Solubility Studies. 1965:117-212.
19. Sambasevam KP, Mohamad S, Sarih NM, Ismail NA. Synthesis and characterization of the inclusion complex of β -cyclodextrin and azomethine. *International journal of molecular sciences*. 2013;14(2):3671-82.

20. Chao J, Meng D, Li J, Xu H, Huang S. Preparation and study on the novel solid inclusion complex of ciprofloxacin with HP- β -cyclodextrin. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2004;60(3):729-34.
21. Khudhur AQ, Maraie NK, Rauf AM. Identification and Quantitation of Phospholipid Binding Sites in Lipid Drug Conjugate Pharmacosomes using Model Drug. *Systematic Reviews in Pharmacy*. 2020;11(5):458-63.
22. Guo B, Liu H, Li Y, Zhao J, Yang D, Wang X, et al. Application of phospholipid complex technique to improve the dissolution and pharmacokinetic of probucol by solvent-evaporation and co-grinding methods. *International journal of pharmaceutics*. 2014;474(1-2):50-6.
23. Syukri Y, Ulfa F, Lestari A, Saputri LA, Istikharah R, Kusuma AP. Characterization, formulation and evaluation of glibenclamide with β -cyclodextrin inclusion complexes tablets. *Jurnal Kedokteran dan Kesehatan Indonesia*. 2018;9(3):139-48.
24. Li B, Zhang W, Ma H. Physicochemical characterization of inclusion complex of catechin and glucosyl- β -cyclodextrin. *Tropical Journal of Pharmaceutical Research*. 2016;15(1):167-72.
25. Singh M, Nagori B, Shaw N, Tiwari M, Jhanwar B. Formulation development & evaluation of topical gel formulations using different gelling agents and its comparison with marketed gel formulation. *International Journal of Pharmaceutical Erudition*. 2013;3(3):1-10.
26. Mario J, Mira B-L, Biserka C. Influence of cyclodextrin complexation on piroxicam gel formulaion. *Acta Pharma* 2005; 55: 223. 2005;236.
27. Mohammed FA. Topical permeation characteristics of diclofenac sodium from NaCMC gels in comparison with conventional gel formulations. *Drug Dev Ind Pharm*. 2001;27(10):1083-97.
28. Prasanthi D, Lakshmi P. Optimisation of transdermal gel formulations of tolterodine tartrate by experimental design. *Turk J Pharm Sci*. 2013;10(2):273-86.
29. Parhi R, Terapalli BR, Teja B. Formulation and in vitro evaluation of minoxidil topical gel. *Turk J Pharm Sci*. 2014;11(2):153-62.
30. Klein RR, Heckart JL, Thakker KD. In vitro release testing methodology and variability with the vertical diffusion cell (VDC). *Dissolution Technol*. 2018;25:52-61.
31. Dash S, Murthy PN, Nath L, Chowdhury P. Kinetic modeling on drug release from controlled drug delivery systems. *Acta Pol Pharm*. 2010;67(3):217-23.
32. Costa P, Lobo JMS. Modeling and comparison of dissolution profiles. *European journal of pharmaceutical sciences*. 2001;13(2):123-33.
33. Pfoze NL, Kumar Y, Myrboh B, Bhagobaty RK, Joshi SR. In vitro antibacterial activity of alkaloid extract from stem bark of Mahonia manipurensis Takeda. *Journal of Medicinal Plants Research*. 2011;5(5):859-61.
34. Al-Majidi SM, Rasheed HAM, Al-Mugdadi SFH. Synthesis, Identification and Evaluation of Antimicrobial Activities of some New N-substituted 2-azetidinone, Imidazolidinone and tetrazole derivatives of 2-(methylthio) benzimidazole. *International Journal of Science and Research (IJSR)*. 2017;6(6):1009-16.
35. Reynolds JE. *Martindale: the extra pharmacopoeia*: London, UK; The Pharmaceutical Press; 1982.
36. Nalluri BN, Chowdary K, Murthy K, Hayman A, Becket G. Physicochemical characterization and dissolution properties of nimesulide-cyclodextrin binary systems. *AAPs PharmSciTech*. 2003;4(1):6-17.
37. Wang S, Jia X-D, Liu M-L, Lu Y, Guo H-Y. Synthesis, antimycobacterial and antibacterial activity of ciprofloxacin derivatives containing a N-substituted benzyl moiety. *Bioorganic & medicinal chemistry letters*. 2012;22(18):5971-5.
38. Başaran B, Bozkir A. Thermosensitive and pH induced in situ ophthalmic gelling system for ciprofloxacin hydrochloride: hydroxypropyl- β -cyclodextrin complex. *Acta Pol Pharm*. 2012;69(6):1137-47.

39. Naidu NB, Chowdary K, Murthy K, Satyanarayana V, Hayman A, Becket G. Physicochemical characterization and dissolution properties of meloxicam–cyclodextrin binary systems. *Journal of pharmaceutical and biomedical analysis*. 2004;35(1):75-86.
40. Mohammed WH, Ali WK, Al-Awady MJ. Evaluation of in vitro drug release kinetics and antibacterial activity of vancomycin HCl-loaded nanogel for topical application. *Journal of Pharmaceutical Sciences and Research*. 2018;10(11):2747-56.
41. Lambers H, Piessens S, Bloem A, Pronk H, Finkel P. Natural skin surface pH is on average below 5, which is beneficial for its resident flora. *International journal of cosmetic science*. 2006;28(5):359-70.
42. Tan YT, Peh KK, Al-Hanbali O. Effect of Carbopol and polyvinylpyrrolidone on the mechanical, rheological, and release properties of bioadhesive polyethylene glycol gels. *Aaps Pharmscitech*. 2000;1(3):69-78.
43. Chandira R, Pradeep PA, Bhowmik D, Chiranjib JB, Tripathi K, Sampath Kumar K. Design, development and formulation of antiacne dermatological gel. *Journal of Chemical and Pharmaceutical Research*. 2010;2(1):401-4.
44. Masar B. Formulation and evaluation of meloxicam as a topical preparation. MSc Collage of pharmacy, University of Baghdad. 2004.
45. Zerrouk N, Corti G, Ancillotti S, Maestrelli F, Cirri M, Mura P. Influence of cyclodextrins and chitosan, separately or in combination, on glyburide solubility and permeability. *European journal of pharmaceutics and biopharmaceutics*. 2006;62(3):241-6.
46. Zimmer Ł, Kasperek R, Poleszak E. Modern polymers in matrix tablets technology. *Polimery w medycynie*. 2014;44(3):189-96.
47. Patilaya P, Husori DI, Marhafanny L. Susceptibility of Klebsiella Pneumoniae Isolated from Pus Specimens of Post-Surgery Patients in Medan, Indonesia to Selected Antibiotics. *Open access Macedonian journal of medical sciences*. 2019;7(22):3861.
48. ISMAEL QA, HAMEED GS, AZIZ FM. Effect of Introduction of Polymers on the Antibacterial Activity of Crystalline Antibiotics. *International Journal of Pharmaceutical Research*. 2020;12(3).