Performance of Germination, Development, Yield, and Pigment on Irrigation with Untreated Paint Industry Wastewateron*tagetes Erecta* L. Var Pusa Basanti

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Abstract

Tagetes erecta L. var Pusa basanti was used as the test plant to estimate the irrigation-potential of paint industry wastewater taken from Bergers Paint Private Limited situated at State Industrial Development Corporation, Samba, Jammu and Kashmir (SIDCO). The dilutions were prepared at T₀, T₂₀, T₄₀, T₆₀, T₈₀, and T₁₀₀ (concentrations ranging from 20% to 100%). The aim was to see the performance of germination, growth, yield, and pigment content at various concentrations. The parameters were found out to be maximum at the lowest concentration (T₂₀) with maximum dilution of 80%. Germination percentage was estimated to be maximum (41.6%). The positive parameters gave the highest values at 20% concentration (T_{20}), and values then declined giving the lowest value at 100% (T_{100}), and vice versa were obtained for the negative parameters. Growth parameters included length of root, shoot, rootshoot ratio, number of leaves, plant height, dry and fresh weight. These parameters indicated the peak values at lower concentrations and vice-versa was obtained at highest concentration. The economic output increased at initial levels upto 60% and then declined. Stover yield though increased at T₂₀ but further at higher concentrations, it declined, and the harvest index was highest at T₁₀₀. The study revealed that lower wastewater concentration caused a stimulatory effect in chlorophyll a and b, whereas total chlorophyll was significant at higher concentrations. Total chlorophyll increased up to 60% concentration but decreased at a higher concentration as the number of days after sowing (DAS) increased. Correlation and ANOVA were performed, which clearly described that the various parameters showed a declining trend with the rising wastewater concentration.

Key words: Carotenoids, Chlorophyll, Germination, Growth, Harvest index, Stover yield, Wastewater

Introduction

With nearly one percent of water resources available for human consumption, water still holds one of the highly plentiful natural resources on the planet (Grey et al., 2013;Kamboj and Choudhury 2013; Anjum et al., 2016; Tian et al., 2018;Choudhury et al., 2021). Throughout the world, water management systems are facing the huge challenge of ever-increasing insecurity of water, floods, and contamination thereof. As per the United Nations Organization estimates, about 80% of sewage is presently released without any prior treatment [UN WWAP (United Nations World Water Assessment Programme), 2017]. In India, industrial wastewater treatment, particularly in the urban and suburban zones, has been a huge challenge. In its report, the Central Pollution Control Board (CPCB) 2013, in

its report, declared that 19,827 MLD out of 53,998 MLD generated were treated in metropolitan, class one, and class two cities.

Further, it was stated that in 2017, out of 18.6% of total treatment capacity, merely 13.5% of sewage was treated (CPCB, 2013 and CPCB, 2017c). Industries are based on the use of water for many operations, which later on lead to the release of polluted water from the industries across the world (Owa et al., 2013; Kamali and Khodaparast, 2015), and the majority of them end up as industrial wastewater (Ranade and Bhandari, 2014; Banerjee et al., 2021). The release of wastewater from the industrial sector is a big concern these days which directly goes to the environment and causes toxicity problems. The paint industry is one among many such industries which is a source of environmental pollution. Paints with variety of chemical composition are used in the whole of the world daily. For example, emulsion paints are a combination of organic and inorganic pigments, extenders, latexes, cellulosic and non-cellulosic thickeners (Krithika and Ligy, 2016). Wastewater from the paint industry frequently comprises constituents that act as a precursor in the manufacturing of paints (Zak et al., 2014). These may include water from the clean-up procedures containing poisonous chemicals, remaining acids, coating metals (Chidozie and Nwakanma, 2017). These wastewaters also contain hazardous chemicals. If released into the environment without prior treatment, they may infiltrate and leach into the groundwater and pollute the groundwater (Olaoye and Oladeji, 2015; Paul et al., 2019). Ongetting discharged into the nearby water bodies, the heavy metals get mixed up in the water bodies. Due to their persistence, they become available for uptake by the aquatic life forms. Another issue is the health and related problems like cancer, deformities, the mutation in genes, and problems in the kidneys, resulting from heavy metals in the paints (Chidozie and Nwakanma, 2017). Irrigation with leftover water from industrial activity is a longstanding practice worldwide (Tripathi et al., 2011). It offers the farmer and agriculture sector water rich in nutrients for irrigation on the one hand and the social set up with a dependable and economical system where the treatment of wastewater and disposal is manageable (Feigin et al., 1991). As a nutrient source, it also acts as a cheap means of irrigation for the farmers, and hence they use industrial wastewater to irrigate their fields. It is challenging to manage industrial wastewater as it has to be used without treatment. As it is not free from toxic chemicals and therefore affects plants' growth when released directly onto the land without prior treatment.

To estimate the irrigation potential of industrial wastewater, paint industry's wastewater was used to irrigate the test plant, *Tagetes erecta* L. var. Pusa basanti (marigold). It was analyzed for germination, growth, and biochemical characteristics including chlorophyll and carotenoids under different untreated paint industry wastewater concentrations. This experiment was conducted to assess the potential of industrial wastewater to be used as a source of irrigation. To date, there is no study performed on the *Tagetes* using paint industry wastewater as irrigant. Therefore, an attempt has been done to estimate the probable effects of wastewater from the paint industry on germination parameters, growth pattern, and yield and pigment content of the *Tagetes erecta* at various concentrations.

Materials and Methods

Study area: It was an experimental work carried out in the department's laboratory at the University of Jammu, Jammu, and Kashmir. The study area lies at an altitude of 1030 feet above sea level with GPS coordinates of 740 24' and 50 18' east and 320 50' and 330 30' north. The annual precipitation of the zone is 1000m. The maximum temperature shoots up to 47° C in summer while the minimum temperature goes down to 4° C in winters. The

wastewater was collected from the Berger Paints industry located in the State Industrial Development Corporation (Samba), Jammu.

Materials: Materials used for the experiment were plastic trays of 450 mm \times 350 mm \times 75 mm dimensions, earthen pots having 220 mm diameter and 5 kg capacity, and chemicals and glassware as per the requirement. Soil samples were procured from the agricultural field at Akhnoor, Jammu, which was dried under the sun. The ratio of soil and farmyard manure was at 3:1. The fungicide bavistine at the rate of 2 gL-1 of water was sprayed on the soil and mixed thoroughly, and the hybrid seeds of *Tagetes erectaL*. var. Pusa basanti.

Methodology: The experiment of germination was conducted in the BOD incubator in November. Twenty healthy seeds were selected and sown in the petridishes in triplicates. The experiment was carried out for twelve days till the germination became constant in all the petridishes, and the counting of seeds was done every second day. Estimated parameters included percentage germination, germination value, peak value, germination period, germination speed, delay index, percent inhibition, and growth and development parameters, including length of root and shoot, weight in terms of dry and fresh as well as moisture content. Germination criteria were visible emergence of the seedlings from the soil, and it was expressed in percentage.

For parameters like growth, yield, and pigment analysis, a pot-level experiment was conducted. 100 ml of each treatment with tap water (in case of control) and different wastewater concentrations (in case of treatments) were sprinkled on the soil in respective sets on alternate days. The trays were also covered with muslin cloth until some germination was observed. The saplings were then transferred in the pots on the 30th day after sowing (DAS).

Germination parameters: Different germination parameters were calculated with the help of below mentioned formulas:

a) Percentage germination: Percentage germination= $\frac{Total \ seeds \ germinated}{Total \ seeds \ sown} \times 100$

b) Germination Index: It was estimated taking following Zucconi et al., 1981.

Germination Index = $\frac{No.of \ grown \ seeds \ in \ sample}{No.of \ grown \ seeds \ in \ control} \times \frac{Average \ sum \ of \ root \ lengt \ h \ in \ sample}{Average \ sum \ of \ root \ lengt \ h \ on \ control} \times 100$

c) Delay Index (DI): It was estimated for comparing performance of crop at various applications (Modified after Kaushik *et al.*, 2005).

DI=X/Y

here,

X=delay in germination time over control (upto 20%) Y=germination time for control

On similar lines, percent inhibition, germination value, peak value, speed of germination and germination time were also calculated with the help of formulae taken from Czabator (1962).

Growth parameters

a) Root length: Three plants were uprooted at specified intervals and their root lengths were measured and they were expressed in centimeters (cms).

b) Shoot length: The shoot lengths were also measured and were expressed in centimeters (cm).

- c) Plant length: Plant length was calculated and expressed in centimeters following Shakila and Usha, 2011.
- d) Biomass: Biomass of the plant samples was determined by the procedure given by APHA (2005

e) Moisture content: Percent moisture was estimated as:

Moisture content = $\frac{Fresh weight - Dry weight}{Fresh weight} \times 100$

Floral characteristics: Various flower characteristics like number of flowers per plant, average flower diameter, average flower weight (fresh), and weight of seeds per treatment were also measured.

Yield parameters: Crop yield was expressed both in terms of economic yield as weight of flowers per plant, as stover yield by subtracting economic yield from total biomass yield or straw yield and harvest index by dividing total economic yield by above ground biomass of plant.

Biochemical parameters: The amount of chlorophyll and carotenoid was calculated according to methods described by Arnon (1949) and Duxbury and Yentsch (1956), respectively.

Results

The various parameters estimated are explained below:

Germination parameters: The experiment of germination was conducted in the BOD incubator where germination was noted each day starting on 3^{rd} day after sowing upto 12^{th} day after sowing (DAS). The 12^{th} DAS was the day when seed germination became constant for all the treatments (**Plate 1 & 2**).



Plate 1 & 2: Uprooted plants after germination at T₀, T₂₀, T₄₀, T₆₀, T₈₀ and T₁₀₀ respectively

The maximum percentage of germination (51.6%) was revealed in the control set, T_0 . At various concentrations, the use of wastewater indicated the maximum percentage germination (41.6%) at T_{20} and minimum (8.3%) at T_{100} . The trend showed that there was decreased with the increase in wastewater concentration, i.e., 41.6%, 31.6%, 30%, 11.6%, and 8.3% at T_{20} , T_{40} , T_{60} , T_{80} , and T_{100} , respectively. The effect of wastewater at various concentrations on different germination parameters is presented graphically in **Fig. 1 and Fig. 2**, which revealed that as the concentration of wastewater increased, the values for positive germination parameters declined, whereas negative germination parameters values increased. Delay index and percent inhibition being the negative germination parameters were zero when rinsed with tap water.



Figure 1: Positive Germination parameters at T_0 to T_{100} Figure 2: Positive Vs negative germination parameters

The comparison between the positive and negative germination parameter at various concentration level is presented in **Fig. 2**. Relation between the positive and negative germination parameters is presented using correlation coefficient below in **table 1**.

Items	Percent germination	Speed of germination	Root length	Shoot length	Plant height	Percent inhibition
Percent germination	1.00000	0.97911	0.93314	0.94300	0.92640	-1.00000
Speed of germination	0.97911	1.00000	0.95852	0.94228	0.93917	-0.97911
Root length	0.93314	0.95852	1.00000	0.97770	0.99050	-0.93314
Shoot length	0.94300	0.94228	0.97770	1.00000	0.99465	-0.94300
Plant height	0.92640	0.93917	0.99050	0.99465	1.00000	-0.92640
Percent inhibition	-1.00000	-0.97911	-0.93314	-0.94300	-0.92640	1.00000

Table 1:	Table showing	the relation	between the	positive and	negative g	germination	parameters

Growth Parameters: The growth parameters estimated were length of root and shoot, leaf count, the height of the plant, weight in terms of fresh and dry for root and shoot, root-shoot ratio, and moisture content. The results for all these parameters have been described graphically in Figures 3a, 3b, and 3c. The highest length of root (1.23 cm), shoot (4.7 cm), and plant height (5.90 cm) were observed at T₂₀. Several leaves were observed to be maximum (2.0) in T₂₀ and T₄₀ and declined further. All the growth parameters (root length-1.57 cm, shoot length-5.5 cm, leaf count-2.0, and height of the plant-7.07 cm) were recorded to be maximum with tap water.



Figure 3a: Root and Shoot length at varied concentrations



6 4 2 0 T0 T20 T40 T60 T80 T100 R/S ratio

Figure 3b: Fresh weight of root, shoot and their ratios



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Moisture content and Biomass: Evident from results, the fresh and dry weight improved with high application of wastewater at the initial stages of sowing season up to 60th day after sowing (DAS) as the maximum fresh weight of plant (28.42 g) and dry weight (6.07 g) was observed at T_{80} respectively. While beyondcertain days after sowing, it i.e., on 90th DAS, the weight began to decline. Maximum fresh weight (14.66 g) and dry weight (4.10 g) were recorded at T_{20} , respectively. At the same time, maximum (3.47 g) and minimum (0.26 g) values for dry weight stood at T_{20} and T_{100} , respectively, on 120th DAS. Tap water irrigation revealed fresh weight values as 11.12 g, 23.23 g, 14.42 g, and dry weight as 4.46 g, 4.10 g, and 3.75 g on 60th, 90th, and 120th DAS, respectively. On 60th DAS, moisture content was observed to be maximum (82.04%) at T_{40} and minimum (62.48%) at T_{100} . On 90th DAS, moisture content was found to be maximum (87.08%) at T_{80} and minimum (75.35%) at T_{20} .

Effect on Flowering: The effect of untreated paint industry wastewater at various concentrations on flower count, flower diameter, and fresh weight for flowers was analyzed every week. The number of flowers varied with the wastewater concentration. A maximum number of flowers (2.0), diameter (7.900 cm), and fresh weight (3.206 g) were observed at T_{20} in the first week of flowering whereas, a minimum number of flowers (1.0), diameter (6.200 cm), and fresh weight (0.967 g) were observed at T_{100} . In the second week of the maximum flowering number of flowers (4.0), diameter (8.100 cm), and fresh weight (4.202 g) were found at T_{40} and T_{20} , respectively. Whereas the minimum number of flowers (3.0), diameter (2.800 cm), and fresh weight (2.208 g) were observed at T_{60} , T_{20} , and T_{80} , respectively. A maximum number of flowers (6.0), diameter (5.500 cm), and fresh weight (5.555 g) were noticed at T_{40} , a minimum number of flowers (3.0), diameter (3.800 cm), and fresh weight (1.287 g) were noticed at T_{100} in the third week of flowering. It was also recorded, the weight of seeds declined as the application of wastewater was raised. The maximum weight of seeds (6234.5 mg) was observed at T_{20} and the minimum (2459.8 mg) at T_{100} . Tap water irrigation (T_0) showed the maximum weight (6776.1 mg) of seeds. ANOVA was performed to determine the difference in length of root and shoot and leaf count on the 60th, 90th, and 120th DAS. The difference was found out to be significant at a 5% level of significance (Table 1).

Source of Variation	SS	Df	MS	F	P-value	F crit
	~~	21	1120	-	1 14100	
Root Length	44.467777	2	22.2338888	4.275565147	0.0339309	3.6823203
	78		9		7	44
Shoot length	445.35444	2	222.677222	11.43955134	0.000961	3.68232
0	44		2			
Number of leaves	21547.11	2	10773.56	17.75503	0.000111	3.68232

Table 1: ANOVA table for length of root and shoot as well as leaf count at different DAS
ANOVA

However, to see the impact of wastewater on length of root and shoot as well as leaf count, ANOVA was again performed and the results have been recorded below in Table 2. The above table revealed that difference amongst the treatments (T_0 , T_{20} , T_{40} , T_{60} , T_{80} , T_{100}) measured across 60th DAS, 90th DAS and 120th DAS were not significant in terms of length of root and shoot as well as leaf count at 5% level of significance.

Table 2: ANOVA table for length of root and shoot as well as leaf count at different concentra	tion level
ANOVA	

Source of Variation	SS	Df	MS	F	P-value	F crit
Root Length	51.684444	5	10.3368888	1.752345074	0.1973817	3.10587523
	44		9		16	9
Shoot Length	90.577777	5	18.1155555	0.336116437	0.8814125	3.10587523

	78		6		18	9
Number of leaves	2383.6111	5	476.722222	0.202391622	0.9552564	3.10587523
	11		2		91	9

To check the difference if fresh weight, dry weight and moisture measured on 60th DAS, 90th DAS and 120th DAS were significantly different, ANOVA has been used.

 Table 3: ANOVA table for Fresh Weight, Dry Weight and Moisture at different DAS

 ANOVA

Source of Variation	SS	Df	MS	F	P-value	F crit
Fresh Weight	70.99574	2	35.49787	0.767628	0.481507	3.68232
Dry Weight	12.68268	2	6.341339	2.754568	0.095737	3.68232
Moisture	404.55	2	202.275	3.675572	0.050227	3.68232

The above table showed that fresh weight, dry weight and moisture content measured on 60th, 90th and 120th DAS were significantly different. However, to observe the effect of wastewater on dry and fresh weight as well as moisture, ANOVA was again performed and the results are shown in below table.

	Table 4: ANOVA	table Fresh Weight	, Dry Weight and N	Aoisture at different co	acentration level
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ANOVA						
Source of Variation	SS	Df	MS	F	P-value	F crit
Fresh Weight	265.8928	5	53.17856	1.279468	0.33463	3.105875
Dry Weight	17.55184	5	3.510369	1.420119	0.285593	3.105875
Moisture	197.109	5	39.4218	0.457983	0.799995	3.105875

The above table showed that difference amongst the treatments (T_0 , T_{20} , T_{40} , T_{60} , T_{80} , T_{100}) measured across 60th DAS, 90th DAS and 120th DAS were not significant in terms of fresh weight, dry weight and moisture at 5% level of significance.

Effect on Yield: To observe the effect on the yield of the test plant, the economic yield, stover index, and harvest index were calculated with different wastewater concentrations. The test plant's economic yield was maximum (10.325 g plant-1) at T_{60} and minimum (4.491 g plant-1) at T_{100} , which implied an increase in the economic yield using wastewater up to 60% but further higher up the yield declined. The stover yield was found to be highest (32.497) at T_{20} and lowest (6.075) at T_{100} , indicating the intolerance of paint industrial wastewater even at minimum concentration. The harvest index was found to be maximum (0.425) at T_{100} and minimum (0.170) at T_{20} , giving the inference that harvest index increased with the help of nutrients available in the paint industry wastewater. The economic yield decreased with the increase in wastewater application. The economic yield remained highest with tap water whereas, stover yield increased at 20 percent and further declined with the increase in wastewater concentration. Harvest index, on the other hand, was observed to be highest at 100% effluent concentration(**Fig. 4**).



Figure 4: Various yield parameters with tap water and waste water at different concentrations

Pigment analysis: The pigments estimated using wastewater were chlorophyll a,b and total chlorophyll, as well as carotenoid content at various time intervals (**Fig. 5**).

Chlorophyll: Chlorophyll a was found to be highest (0.211 mg/g, 0.251 mg/g and 0.140 mg/g) at T_{20} whereas, least values (0.156 mg/g, 0.122 mg/g and 0.114 mg/g) were observed at T_{100} on 60th, 90th and 120th DAS respectively. Chlorophyll b was found to be maximum (0.055 mg/g, 0.096 mg/g and 0.048 mg/g) at T_{20} on 60th, 90th and 120th DAS and minimum values of chlorophyll b (0.045 mg/g, 0.044 mg/g and 0.034 mg/g) at T_{100} on 60th, 90th and 120th DAS. Tap water (T_0) revealed the values of chlorophyll b as 0.061 mg g-1, 0.059 mg g-1, and 0.033 mg g-1 on 60th, 90th, and 120th DAS, respectively. With a unique difference, it was found out that the highest value of total chlorophyll (0.301 mg/g) was observed at T_{100} on 60th DAS. Whereas on 90th and 120th DAS, the maximum was (0.347 mg/g and 0.180 mg/g) values were observed at T_{20} . T_0 revealed the total chlorophyll as 0.272 mg/g, 0.222 mg/g, and 0.131 mg/g on 60th, 90th, and 120th DAS. Thus, it was found that chlorophyll a and b increased at lower concentrations but decreased at higher concentrations. It was also found that the chlorophyll a and b declined with time and raised the application of wastewater whereas, total chlorophyll showed a varied trend over the period.

Carotenoids: The maximum value of carotenoids was observed as 0.104 mg/g and 0.115 mg/g at T_{20} on 60th and 90th DAS, respectively, and 0.056 mg/g at T60 on 120th DAS. Whereas minimum values (0.093 mg/g, 0.050 mg/g, and 0.043 mg/g) were observed at T_{100} on 60th, 90th, and 120th DAS, respectively. Tap water irrigation revealed the values to be 0.103 mg/g, 0.070 mg/g, and 0.038 mg/g on 60th, 90th, and 120th DAS. Thus, it was recorded that there was a substantial rise in the pigment content at lower wastewater applications whereas, it declined at raised levels of applications.



Figure 5: Pigment determination (Chl a, b and total chlorophyll and carotenoids) at varied concentrations

Discussion

Several studies have been conducted so far in this field, where various industrial wastewaters are used to check the irrigation potential of various crops. The experiments have been conducted with treated as well as untreated wastewater. Paint industrial wastewater studies conducted by various workers are Aniyikaiye et al., 2019; Damaraju et al., 2019; Bhat et al., 2018; Chidozie and Nwakanma, 2017; Guven et al., 2017; Krihika and Ligy, 2016; Olaoye and Oladeji, 2015; Jolly et al., 2008. Various studies by a variety of researchers carried out using a range of industrial wastewater on different crops are by Lallawmsanga et al., 2012; Vijayaragvan et al., 2011; Malaviya and Sharma 2011; Kumar, 2011; Yousaf et al., 2010; wins and Murugan (2010); Ogunwenmo et al., 2010; Singh, 2010; Nagajyothi et al., 2009; Rehman et al., 2009. Our findings were quite close to the studies conducted by Patro et al., 2007; Malaviya et al. (2007); Garg and Kaushik, (2007); Nath et al. (2007); Singh et al.(2006); Pandey (2006); Pragasam et al. (2005), Singh et al. (2004). Pieces of evidence were also presented by Gouia et al.(2003); Singh et al. (2002); Ramana et al. (2002); Augusthy and Mani, (2001); Reddy and Borse, (2001); Javaid et al.(2000); Prasanna et al. (1997), Leubner-Mergzer et al.(1995), Krupa et al.(1993), Kelly et al.(1992), Groot et al.(1992) Gautam and Bishnoi (1990), Choudhury and Choudhury(2014) Purkayastha, P et al. (2015). Our study was different because the untreated paint industry wastewater did not support the plant growth and development even at the lowest percentage of 20% concentration and 80% water.

Conclusion

The paint industry wastewater proved to be toxic for *Tagetes erecta*, as revealed through results and statistical analysis performed on them. The germination parameters showed that with increasing concentration, the positive germination parameters showed a decline while the negative parameters increased. Germination is the positively affected parameter of *Tagetes erecta* with untreated wastewater from the paint industry at lower concentrations but not at higher dose. The case of other parameters like growth, yield, and pigment contents revealed a rising trend at initial applications upto 60th DAS but declined further with higher dose and number of DAS. Economic output enhanced with the rise in the wastewater up to a certain level, whereas it declined further. The harvest index showed to be highest at the highest concentration of 100%. Therefore, paint industry wastewater requires pre-treatment before it is discharged into the environment or released into drain drains from where it is further carried to the agricultural fields. The correlation table showed the relationship between the parameters. The closely related parameters were expected to have a high correlation and vice-versa.

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References

- 1. Aniyikaiye, T.E., Oluseyi, T., Odiyo, J.O. & Edokpayi, J.N. (2019). Physico-chemical analysis of wastewater discharge from selected paint industries in Lagos, Nigeria. *International Journal of Environmental Research and Public Health*, 1-17.
- 2. Anjum, M., Miandad, R., Waqas, M., Gehany, F. & Barakat, M.A. (2016). Remediation of wastewater using various nano-materials. *Arab. J. Chem.*, <u>https://doi.org/10.1016/j</u>. arabjc.
- 3. APHA (American Public Health Association). (2005). Standard methods for the examination of water and waste water. 21st edition, American water works association; Water Environment Federation, Washington DC.
- 4. Arnon, D.I. (1949). Copper enzymes in isolated chloroplast, polyphenol oxidase in *Beta vulgaris*. *Plant Physiology*, 24, 1-15.

- 5. Augusthy, P.O. & Mani, A.S. (2001). Effect of rubber factory effluent on seed germination and seedling growth of *Vigna radiata L. Journal of Environment Biology*, 22, 137-139.
- Banerjee, A., Choudhury, M., Chakravorty, A., Raghavan, V., Biswas, B., Sana, SS., Rayan, R.A., Abhishek, N., Lala, N.L., Ramakrishna, S., (2021) Microbiological Degradation of Organic Pollutants from Industrial Wastewater in A Kumar & C Rao (Eds) *Nanobiotechnology for Green Environment* (pp 83-116) London, UK by CRC Press <u>10.1201/9780367461362-4</u>
- 7. Bhat, N.B., Parveen, S. & Hassan, T. (2018). Seasonal Assessment of Physicochemical Parameters and Evaluation of Water Quality of River Yamuna, India. *Adv. Environ. Technol.*, 1, 41–49.
- 8. Chidozie, K. & Nwakanma, C. (2017). Assessment of Saclux Paint Industrial Effluents on Nkoho River in Abia State, Nigeria. J. Ecosyst. Ecography, 7, 1–8.
- 9. Choudhury, M. & Choudhury, M. (2014). Trends of Urban Solid Waste Management in Agartala City, Tripura, India. *Universal Journal of Environmental Research & Technology*, 4(4).
- Choudhury, M., Jyethi, D.S., Dutta, J. (2021). Investigation of groundwater and soil quality near to a municipal waste disposal site in Silchar, Assam, India. Int J Energ Water Reshttps://doi.org/10.1007/s42108-021-00117-5
- 11. Choudhury. M, Paul C. &Kamboj, N. (2014). Potable water is a serious Environmental issue: A special study on Umiam area, of RI-Bhoi District, Meghalaya, India. *Int Res J Environ Sci*, 3, 37-42.
- CPCB (2013). Performance Evaluation of Treatment Plants in India under funding of NRDC. Available online at: <u>http://www.indiaenvironmentportal.org.in/files/</u> file/STP__REPORT.pdf (accessed December 12, 2019).
- 13. CPCB (2017c). Status of STPs. Available online at: <u>https://cpcb.nic.in/status-ofstps/(accessed July 01, 2019)</u>.
- 14. Czabator, F.J. (1962). Germination value: An index combining speed and completeness of pine seed germination. *Forensic Science*, 8, 386-396.
- 15. Damaraju, M., Bhattacharya, D., Panda, T.K. & Kurilla, K.K. (2019). Marigold wastewater treatment in a lab-scale continuous bipolar-mode electrocoagulation system. *Journal of Cleaner Production*, 1-40.
- 16. Debeujan, I., Karen, M. & Koorneef, L.M. (2000). Influence of the testa on seed dormancy, germination and longevity in *Arabidopsis*. *Plant physiology*, 122, 403-413.
- 17. Duxbury, A.C. & Yentsch, C.S. (1956). Plankton pigment monographs. *Journal of Marine Research*, 15, 91-101.
- 18. Feigin, A., Ravina, I. & Shalhevet. J. (1991). Irrigation with treated sewage effluent. Management for Environmental Protection. *Advanced Series in Agricultural Sciences*, 17, 224. Springer-Verla.
- Feng, X., Liu, Q., Chen, Y., Yin, L. & Fu, B. (2018). Linking water research with the sustainability of the human - natural system. *Curr. Opin. Environ. Sustain*, 33, 99–103. <u>https://doi.org/10.1016/j.cosust.2018.05.012</u>.
- 20. Garg, V.K. & Kaushik, P. (2007). Influence of textile mill waste water irrigation on growth of Sorghum cultivars. *Applied Ecology and Environmental Research*, 6, 1-12.
- 21. Gautam, D.D. & Bishnoi, S. (1990). Effect of dairy effluent on wheat (*Triticum aestivum*). Journal of Ecobiology, 4, 111-115.
- 22. Gouia, H., Suzuki, A., Brulfert, & Ghorbal, M.H. (2003). Effects of cadmium on the co-ordination of nitrogen and carbon metabolism in seedlings. *Plant Physiology*, 160, 367-376.

- 23. Grey, D., Garrick, D., Blackmore, D., Kelman, J., Muller, M. & Sadoff, C. (2013). Water security in one blue planet: twenty-first century policy challenges for science. *Philos. Trans. R. Soc. A Math. Phys. Eng. Sci*, 371, 20120406.
- 24. Groot, S.P.C. & Karsen, C.M. (1992). Dormancy and germination of abscisic acid and deficient tomato seeds. *Plant Physiology*, 99: 952-958.
- 25. Guven, D., Hanhan, O., Aksoy, E.C., Insel, G. & Cokgor, E. (2017). Impact of paint shop decanter effluents on biological treatability of automotive industry wastewater. *Journal of Hazardous Materials*, 330, 61-67.
- Javaid, A., Ashraf, S. & Bajwa, R. (2000). Effect of tannery industrial effluents on crop growth and VAM colonization in *Vigna radiata* (L) Wilczek and *Zea mays* L. *Pakistan journal of Biological Sciences*, 3, 1292-1295.
- 27. Jolly, Y.N., Islam, A. & Quraishi, S.B. (2008). Effects of paint industry effluent on soil productivity. Pp-41-53.
- 28. Kamali, M. & Khodaparast, Z. (2015). Review on recent developments on pulp and paper mill wastewater treatment. *Ecotoxicol. Environ. Saf*, 114, 326–342.
- 29. Kaushik, P., Garg, V.K. & Singh, B. (2005). Effect of textile effluents on growth performance of wheat cultivars. *Bioresource Technology*, 96, 1189-1193.
- 30. Kelly, K.M., Van Staden, J. & Bell, W.E. (1992). Seed coat structure and dormancy. *Plant growth Regulation*, 11, 201-209.
- 31. Krithika, D. & Ligy, P. (2016). Treatment of Wastewater from Water Based Paint Industries Using Submerged Attached Growth Reactor. *Int. Biodeter. Biodegr. J*, 107, 31–41.
- 32. Krupa, Z., Oquist, G. & Huner, N. (1993). The effect of cadmium on photosynthesis of *Phaseolus vulgaris*-A fluorescence analysis. *Physiologia Planatarum*, 88, 626-630.
- 33. Kumar, A. (2011). Effect of dairy effluents on seed germination and early seedling growth of *Pennisetum typhoides*. *International Journal of Applied Engineering Reasearch*, 1.
- Lallawmsanga, M.K., Balakumaran, D.J., Ravikumar, M.D., Jeyarathi, M.J. & Kalaichelvan, P.T. (2012). Ameliorating effect of vermicompost and cow dung compost on growth and biochemical characteristics of *Solanum melongena* L. treated with paint industrial effluent. *Annals of Biological Research*, 3, 2268-2274.
- 35. Leubner- Metgzer, G., Fruindt, C., Vogeli-Lange, R. & Meins, J. (1995). 1,2- Glucanase in the endosperm of tobacco during germination. *Plant physiology*, 109, 751-759.
- 36. Malaviya, P. & Sharma, A. (2011). Effect of distillery effluent on yield attributes of *Brassicanapus* L. *Journal of Environmental Biology*, 32, 385-389.
- 37. Malaviya, P., Kour, R. & Sharma, N. (2007). Growth and yield response of *Capsicum annum* L. to distillery effluent irrigation. *Indian Journal of Environment and Eco planning*, 14, 643-646.
- 38. Mishra, V. & Pandey, S.D. (2002). Effect of distillery effluent and leachates of industrial sludge in the germination of black gram (*Cicer arietinum*). *Pollution Research*, 21, 461-467.
- Nagajyothi, P.C., Dinakar, N., Suresh, S., Udaykiran, Y., Suresh, C. & Damodharan, T. (2009). Effect of industrial effluent on the morphological parameters and chlorophyll content of green gram (*Phaseolus aureus* Roxb). *Journal of Environmental Biology*, 30, 385-388.
- 40. Nath, K., Singh, D. & Sharma, Y.K. (2007). Combinatorial effects of distillery and sugar effluents in crop plants. *Journal of Environmental Biology*, 28, 577-582.

- 41. Nitin, K. & Choudhury, M. (2013). "Impact of solid waste disposal on ground water quality near Gazipur dumping site, Delhi, India." *Journal of Applied and Natural Science*, 2, 306-312.
- 42. Ogunwenmo, K.O., Oyelana, O.A., Ibidunmoye, O., Anyaso, G. & Ogunnowo, A.A. (2010). Effects of brewery, textile and paint effluent on seed germination of leafy vegetables- *Amaranthus hybridus* and *Celosia argentea* (Amaranthaceae). *Journal of Biological Sciences*, 10, 151-156.
- 43. Olaoye, R.A. & Oladeji, O.S. (2015). Preliminary Assessment of Effects of Paint Industry Effluents on Local Groundwater Regime in Ibadan, Nigeria. *Int. J. Eng. Res.*, 4, 518–522.
- Olvera, R.C., Silva, S.L., Robles-Belmont. E. & Lau, E.Z. (2017). Review of nanotechnology value chain for water treatment applications in Mexico. *Resour. Technol.*, 3, 1–11. <u>https://doi.org/10.1016/j.reffit.2017.01.008</u>.
- 45. Owa, F.D. (2013). Water Pollution: Sources, Effects, Control and Management. *Mediterr. J. Soc. Sci.*, 4, 65–68.
- 46. Pandey, S.N. (2006). Effect of brewery effluent on seedlings of *Cajanus cajan* and *Vigna mungo*. Journal of Applied Biosciences, 32, 199-122.
- Patro, B., Ghosh, S. & Panigrahi, S. (2007). Effect of industrial effluent on photosynthetic pigment degradation in paddy (*Oryza sativa* L.) leaves during senescence. *Environmental Conservation Journal*, 6, 381-384.
- Paul, S., Choudhury, M., Deb, U., Pegu, R., Das, S., & Bhattacharya, S. S. (2019). Assessing the ecological impacts of ageing on hazard potential of solid waste landfills: A green approach through vermitechnology. *Journal of Cleaner Production*, 236, 117643.
- 49. Pragasam, A., Praveena, R. & Prasena, J. (2005). Effects of brewery effluent on the photosynthetic pigments, starch, nitrate reductase activity, and protein content of *Vigna mungo*. *Indian Journal of Environmental Ecoplanning*, 10, 13-18.
- 50. Prasanna, K.P.G., Pandit, B.R. & Kumar, M.R.K. (1997). Effect of dairy effluent on seed germination, seedling growth and pigment content of green gram (*Phaseolus aureus*) and black gram (*Phaseolus mungo*). Advances in Plant Sciences, 10, 129-136.
- 51. Purkayastha, PS, Choudhury, M., Deb, D., &Paul, C.(2015) "Arsenic contamination in ground water is a serious threat in the North Karimganj block of Karimganj district, Southern part of Assam, India." *Journal of Chemical and Pharmaceutical Research* 7.8, 371-378.
- 52. Ramana, S., Biswas, A.K., Kundu, S., Saha, J.K. & Yadava, R.B.R. (2002). Effect of distillery effluent on seed germination in some vegetable crops. *Bioresource Technology*, 82, 73-275.
- 53. Ranade, V.V. & Bhandari, V.M. (2014). Chapter 1 Chemical Engineering and Process Development Division. In Industrial Wastewater Treatment, Recycling, and Reuse: An Overview; CSIR-National Chemical Laboratory: Pune, India. 1–80.
- 54. Reddy, P.G. & Borse, R.D. (2001). Effect of pulp and paper mill effluent on seed germination and seedling growth of *Trigonella foenum-graecum* L. (Methi). *Journal of Industrial Pollution Control*, 17, 165-169.
- 55. Rehman, A., Bhatti, H.N. & Athar, H.R. (2009). Textile effluents affected seed germination and early growth of some water vegetable crops: A case study. *Water Air Soil Pollut.*, 198, 155-163.
- 56. Shakila, P.B. & Usha, K. (2011). Effect of tannery effluent on water and soil profile, plant growth and human health. *Advanced Studies in Biology*, 8, 391-398.
- 57. Shuval, H.I., Adin, A., Fattal, B., Rawitz. E. & Yekutiel, P. (1986). Wastewater irrigation in developing countries, Health effects and technical solutions. 325, World Bank Technical Report 51.

- Singh, A., Aggarwal, S.B. Rai, J.P.N. & Singh, P.L. (2002). Assessment of pulp and paper mill effluent on growth, yield and nutrient quality of wheat (*Triticum aestivum* L.). *Journal of Environmental Biology*, 23, 283-288.
- 59. Singh, C.S. (2010). Effect of nickel plating industry effluent on seed germination of *Cicerarietinum* C.V. G-30 and *Cicer arietinum* C.V. H-208. *Indian Journal of Science Research*, 1, 63-65.
- 60. Singh, P.P. Mall, M. & Singh, J. (2006). Impact of fertilizer factory effluent on seed germination, seedling growth and chlorophyll content of gram (*Cicer arietinum*). *Journal of Environmental Biology*, 27, 153-156.
- Singh, S., Srivastava, P., Garade, S. & Bhatnagar, M.K. (2006). Effect of chlor-alkali plant effluent on seed germination and early growth performance of some common plant species. *Pollution Research*, 25, 451-452.
- 62. Tian, X., Sarkis, J., Geng, Y., Qian, Y., Gao, C., Bleischwitz, R. & Xu, Y. (2018). Evolution of China's water footprint and virtual water trade: a global trade assessment. *Environ. Int.*, 121, 178–188. https://doi.org/10.1016/j.envint.2018.09.011.
- Tortajada, C. & Biswas, A.K. (2018). Achieving universal access to clean water and sanitation in an era of water scarcity: strengthening contributions from academia. *Curr. Opin. Environ. Sustain.*, 34, 21–25. <u>https://doi.org/10.1016/j.cosust.2018.08.001</u>.
- 64. Tripathi, D.M., Tripathi, S. & Tripathi, B.D. (2011). Implications of secondary treated distillery effluent irrigation on soil cellulase and urease activities. Journal of Environment Protection. 2, 655-661.
- 65. UN WWAP (United Nations World Water Assessment Programme) (2017). The United Nations World Water Development Report (2017). Wastewater: TheUntapped Resource. Paris: UNESCO.
- Vijayaragavan, M., Prabhakar, C., Sureshkumar, J., Natarajan, A., Vijayarengan, P. & Sharavanan, S. (2011). Soil irrigation effect of sugar mill effluent on changes of growth and biochemical contents of *Raphanus sativus* L. *Botany Current*, 2, 9-13.
- 67. Wins, J.A. & Murugan, M. (2010). Effect of textile mill effluent on growth and germination of black gram *Vigna mungo* (L.) Hepper. *International Journal of Pharmaceuticals and Biosciences*, 1, 1-7.
- 68. Yousaf, I., Ali, S.M. & Yasmin, A. (2010). Germination and early growth response of *Glycine max* varieties in textile and paper industry effluents. *Pakistan Journal of Botany*, 42, 3857-3863.
- 69. Zak, S., Rauckyte-Z' ak, T., Laurinavic'ius, A. & Siudzin' ski, P. (2014). Research on Physico-chemical Pretreatment of Wastewater from the Production of Wood Coating Materials. *Ecol. Chem. Eng. S.*, 21, 101–112.
- Zhang, L., Cheng, L., Chiew, F. & Fu, B. (2018a). Understanding the impacts of climate and landuse change on water yield. *Curr. Opin. Environ. Sustain.*, 33, 167–174. https:// doi.org/10.1016/j.cosust.2018.04.017.
- 71. Zucconi, F.M., Forte. A., Monaco, & Bertoldo, M. (1981). Biological evaluation of compost maturity. *Biocycle*, 22, 27-29.