

Western Blotting and Bicinchonic Acid Assessment for Improvement of Sperms Mitochondrial Functions by Fenugreek, Extract and Nanoparticles

Mohammed T. Jaafar^{1*}, Hayder A. N. Al-Zamely², Adnan Mansour Jasim³

^{1,3} Department of Physiology, Biochemistry and Pharmacology, College of Veterinary Medicine, Al-Qasim Green University, Al-Qasim, Babil, Iraq

² Department of Physiology, Biochemistry and Pharmacology, College of Veterinary Medicine, University of Al-Qadisiyah, Al-Diwaniyah, Iraq

Emails: dr.mohammed.t.g@gmail.com¹, hayder.alzamely@qu.edu.iq²,
adnan.mansouri81@vet.uoqasim.edu.iq³

** Corresponding author*

Abstract

This study was aimed for assessment the effect of fenugreek extract and fenugreek NPs on the concentration of total protein in testicular tissues through application of two assays, Western blotting and Bicinchonic Acid. Totally, 60 Wister rats were selected, prepared and equally divided into three groups; negative control that remains without any treatment and given only distilled water daily; T1 that orally treated with a daily dose (100) mg/kg of extract fenugreek, and T2 that orally treated with a daily dose (100) mg/kg of fenugreek NPs. After an experimental period continued for 60 days, all study animals were euthanized with chloroform to collect of testis samples that subjected for homogenization and measurement of proteins concentration targeting of SDHA and CS proteins. The findings revealed that there is a significant increase in concentration of total protein in both fenugreek extract and fenugreek NP groups when compared with the value of control group. However, no significant variation was found between values of both fenugreek extract and fenugreek NP groups. Expression of mitochondrial expression of SDHA protein was elevated in fenugreek extract group and fenugreek NP groups compared with control group. A comparison of values of treatment groups was revealed an elevation in values of fenugreek NP group more than fenugreek extract group. For CS protein, insignificant variation was reported between values of fenugreek extract and fenugreek NP groups when compared to value of control group. In conclusion, Western blotting demonstrated that is a powerful tool for detection and characterization of targeted proteins. Also, BCA revealed a high sensitivity and simple immunoassay for screening of increased total testicular protein. However, isolation of active components in fenugreek, and synthesizing of specific NPs based on one type of these components can provide new therapies with different characteristics. Also, the augmentation of fenugreek NPs

and proper regulation of steroidogenesis and mitochondrial biogenesis related genes is notably need to be furthermore studied.

Keywords: Succinate dehydrogenase, Citrate synthase, Chitosan, Immunoassay, Iraq

Introduction

Sperms are composed of important component of mitochondrial parts. Anifandis *et al.* (2017) was found a link between sperm mitochondrial defects and decreased sperm motility. The molecular level has revealed that deletions and other changes to mitochondrial DNA can result in decreased sperm functionality and male infertility (Bahrehmand Namaghi and Vaziri, 2017). Comparing sperm from asthenozoospermic samples to control samples revealed that sperm from asthenozoospermic samples contain abnormally high levels of specific mitochondrial RNAs, as well as, transcripts encoding mitochondrial proteins that are encoded by the nucleus (Ferramosca *et al.*, 2021). Another significant discovery has been the relationship between the activity of sperm mitochondrial enzymes including the mitochondrial electron transport chain complexes in addition to a variety of sperm parameters such as concentration, vitality and motility among other characteristics (Zhu *et al.*, 2019). Also, many researchers have been discovered the citrate synthase (CS) and succinate dehydrogenase (SDHA) as both of which are nuclear-encoded enzymes involved in the Krebs cycle, exhibited the strongest relationships with one another out of all of the enzymes studied (Beyramzadeh *et al.*, 2017; Fišar *et al.*, 2019). In relation to citrate synthase (which is commonly used to indicate the presence or absence of mitochondria), the researchers have discovered that, in the future, mitochondrial volume, rather than distinct enzymatic activities in samples of varying quality may serve as a primary explanation for the correlations in question (Soren *et al.*, 2018; Kang *et al.*, 2020).

One of the medicinal plants that has been used since antiquity in the traditional medicine and for which significant therapeutic properties have been mentioned is fenugreek (*Trigonella foenum-graecum*), (Srinivasa and Naidu, 2021). Fenugreek contains a number of chemical constituents including steroidal sapogenins such as diosgenin component that found in the oily embryo of fenugreek. There are two furastanol glycosides, F-ring opened precursors of diosgenin that have been reported in fenugreek as also hederagin glycosides (Chaudhary *et al.*, 2018). Alkaloid like trigocoumarin, nicotinic acid, trimethyl coumarin and trigonelline are present in stem. The mucilage is a standing out constituent of the seeds (Aher *et al.*, 2016).

Nanoparticles (NPs) represent an active area of research and a techno-economic sector with full expansion in many application domains. NPs have gained prominence in technological

advancements due to their tunable physicochemical characteristics such as melting point, wettability, electrical and thermal conductivity, catalytic activity, light absorption and scattering resulting in enhanced performance over their bulk counterparts (Hu, 2015). NPs have drawn increasing interest from every branch of medicine for their ability to deliver drugs in the optimum dosage range often resulting in increased therapeutic efficiency of the drugs, weakened side effects and improved patient compliance (Khan et al., 2018). The controlled release of pharmacologically active drugs to the precise action site at the therapeutically optimum degree and dose regimen has been a major goal in designing such devices (Simonazzi *et al.*, 2018). Hence, this study was aimed for assessment the effect of fenugreek extract and fenugreek NPs on the concentration of total protein in testicular tissues through application of two assays, Western blotting and Bicinchonic Acid.

Materials and methods

Ethical approval

The current study was performed under the license of the Department of Physiology, Biochemistry and Pharmacology, College of Veterinary Medicine, Al-Qasim Green University (Al-Qasim, Babil, Iraq).

Preparation of fenugreek extract and NPs

Initially, a total 300 g of fenugreek seeds were purchased from the local market of herbal medicine in AL-Qasim city, grinded using a blender max. The powder of fenugreek seeds was dissolved then into a totally 1000 ml of alcohol solution, hydroalcoholic acid (70%), with mechanical shaking by magnetic stirrer at 55°C for 6 hours. The mixture was filtered, dried under the vacuum for 20 hours, incubated at 37°C for 36 hours, and stored in deep freezes at -20°C (Banso and Adeyemo, 2006). Chitosan was used to prepare of the NPs by the ionic gelation method with some modifications (Agarwal et al., 2018; Shafiei et al., 2019).

Study design and sample collection

Sixty Wister rats were randomly selected and subjected for preparatory period; during which, they kept at $23 \pm 2^\circ\text{C}$, fed on basal laboratory diet and tap water, and exposed to 12/12 light and dark conditions. After one week, the study animals were equally divided into three groups as following:

1. First group (Negative control group): Rats of this group still without any treatment and given only distilled water daily.
2. Second group (T1): Rats of this group treated with a daily dose (100) mg/kg of extract fenugreek that given orally by stomach tube.

3. Third group (T2): Rats of this group treated with a daily dose (100) mg/kg of fenugreek NPs that given orally by stomach tube (Al-Zamely and Kshash, 2021). After an experimental period continued for 60 days, all study animals were euthanized with chloroform and subjected for sampling of testis.

Bicinchonic Acid Assay

The concentrations of total proteins in homogenized testicular tissues were determined following the manufacturer's instructions of the Bicinchoninic Acid Assay Kit (Elabscience, China). Briefly, all samples were diluted, pipetted into the wells of the plate, and the BCA reagents were added. The plate was covered, incubated at 37°C for 30 min, and read at an optical density (OD) of 562 nm using a Microplate reader (BioTek, USA). The linear standard curve of mean absorbance was used to measurement of the concentration of protein in all samples.

Western blotting assay (SDS-PAGE)

All buffers and reagent used in this assay were imported from the Elabscience Company (China). The samples were taken, and the tissues were washed thoroughly with pre-cooled PBS Buffer to remove the surface blood and internal debris. Then, the tissues were weighted and smashed with adding of an appropriate ratio of RIPA Lysis Buffer to homogenizely lyse the tissue. After homogenization, the sample was shacked and lysed on the ice for 30 min, and then, sonicated for 1 min to make cells fully lysis and reduce the viscosity of sample. The lysate was centrifuged at 12,000 rpm for 10 min at 4°C, and the supernatant was taken. The protein concentration was measure as mentioned in Bicinchonic Acid Assay method. According to molecular weight of targeted proteins, separation gel was prepared, and the test samples were added to each well with adding 5µl of Precision Plus Protein™ Dual Color Standards Marker to a reserved well in order to verify the target molecular weight and the extent of membrane transfer. Electrophoresis Buffer was added and electrophoresis was carried out at 80V for 2-3 hours till bromophenol blue reaches the bottom of the gel. The PVDF Membrane was soaked in methanol for 1 min to activate it, and then soaked in Transmembrane Buffer. The transmembrane conditions were adjusted, and the transmembrane process was carried out at low temperatures. After the transmembrane, the PVDF Membrane was taken out carefully and washed with TBST Buffer for 1 min. The PVDF Membrane was soaked with TBST Buffer and incubated at room temperature for 1.5 hour. According to dilution ratio, the TBST Buffer was used to dilute the primary antibody according to the antibody manual. The PVDF Membrane was soaked in the primary antibody working solution, incubated overnight at 4°C, and then, the PVDF Membrane was washed with TBST Buffer for 3 times. According to the recommended dilution ratio, secondary antibody was diluted with TBST Buffer,

incubated at room temperature for 1 hour, and then, the PVDF Membrane was washed with TBST Buffer for 3 times. The A and B Buffers were mixed in the Excellent Chemiluminescent Substrate Detection Kit at the ratio of 1:1 as working solution, and the PVDF Membrane was taken out from the TBST Buffer. The PVDF Membrane was placed inside the X-ray cassette with adding of ECL working solution continuously on the PVDF Membrane. Protein bands intensity was measured using Image Studio Lite software version 5.2, and the unit used for bands measurement was Arbitrary units (AU).

Statistical analysis

All collected data were analyzed by ANOVA at $P < 0.05$ in GraphPad Prism Software (version 6.0.1), (Al-Gharban and Al-Taee, 2016; Gharban et al., 2019).

Results and discussion

Bicinchonic Acid Assay

The results of total protein concentration seen that there was a significant increase ($p < 0.05$) in values of both fenugreek extract (19.87 ± 1.245) and fenugreek NP (20.93 ± 2.128) groups when compared with the value of control (15.33 ± 0.895) group. However, no significant variation was found between values of both fenugreek extract and fenugreek NP groups (Table 1; Figure 1). Based on our data, BCA revealed a high sensitivity and simple immunoassay to screening of increased total testicular protein as a result of administration of fenugreek extract and fenugreek NPs which in agreement with that reported recently (Li *et al.*, 2018; Zhou *et al.*, 2020; Singh *et al.*, 2022). Sedha *et al.* (2015) delineated that apoptosis of germ cells and generation of oxidative stress can further compromise capacity of leydig cells to be steroidogenic and ability to differentiate of germinal epithelium into normal spermatozoa. Hamza *et al.* (2016) advocated that the aqueous extract of fenugreek seeds appreciatively attenuate the testicular damage induced by cisplatin. Fenugreek extract have shown various curative qualities suchlike hypocholesterolaemic, antibacterial, antioxidant, antidiabetic agent, hepatoprotective effect and anticancer (Wani and Kumar, 2018). Neha *et al.* (2019) reported that fenugreek extract showed a protective effect against lipid peroxidation in testicular tissue. Kaur and Sadwal (2020) demonstrated that the administration of fenugreek was improved the deleterious effects on male reproductive system as indicated by improved sperm parameters.

Table (1): Concentration of total protein among study groups

Standard concentrations of BSA ¹ (mg/ml)	² Reading data Absorbance unite (nm)		Samples Codes	² Reading data Absorbance unite (nm)		Final Concentration of total protein (µg/µl)
3	2.694	2.613	W1	1.594	1.669	16.9
2.4	2.22	2.238	W2	1.464	1.523	15.3
1.8	1.749	1.758	W3	2.303	1.391	13.8
1.2	1.272	1.275	R1	1.914	1.993	20.8
0.6	0.756	0.822	R2	2.058	1.944	21.4
0.3	0.435	0.429	R3	1.65	1.685	17.4
0	0.177	0.171	B1	1.35	2.311	25.1
			B2	1.865	1.845	19.6
			B3	1.771	1.679	18.1

¹ Bovine serum albumin standard concentrations

² Absorbance reading using Microplate reader at 562 nm wavelength

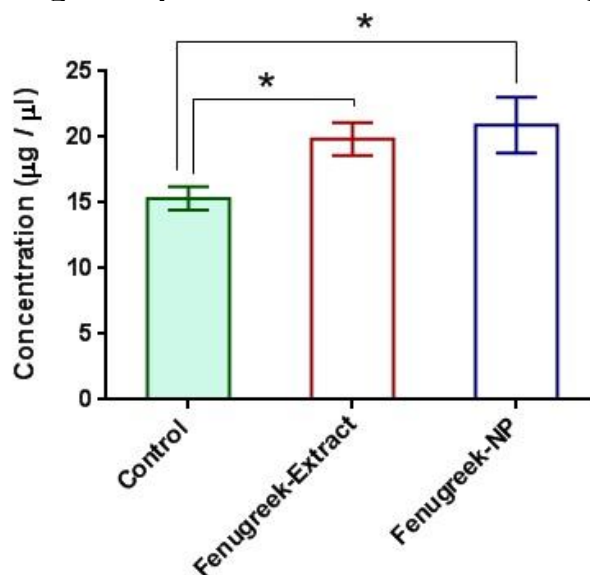


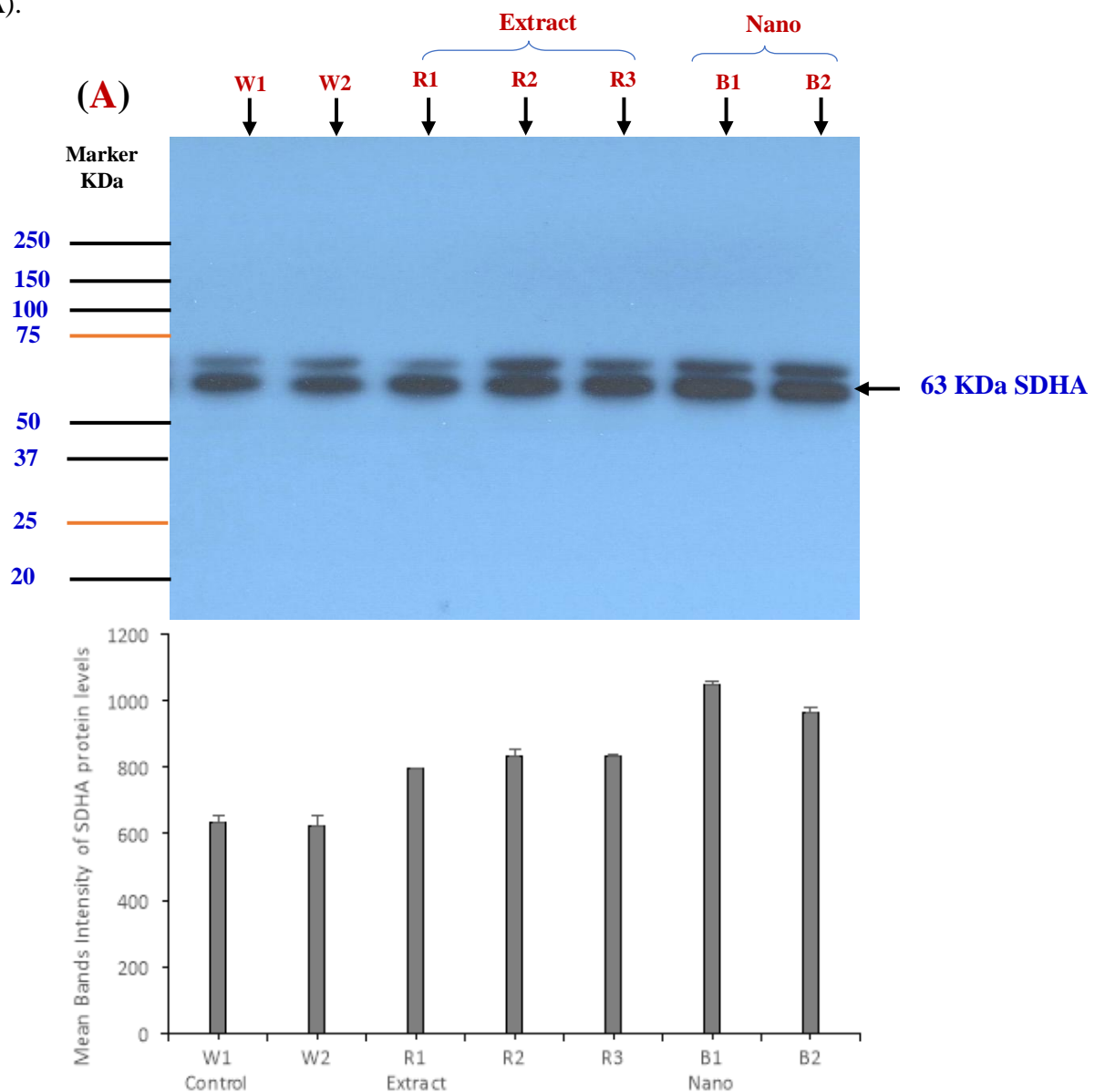
Figure (1): Values (M±SE) of total protein concentration among study groups

Further, sperm motility and sperm concentration were elevated in rat administered the extract demonstrating the antioxidant potential of fenugreek that helps to decrease the oxidative stress. Sadogh *et al.* (2022) mentioned that taking fenugreek seed oil drops improves the sperm count in

men with a low concentration of sperm.

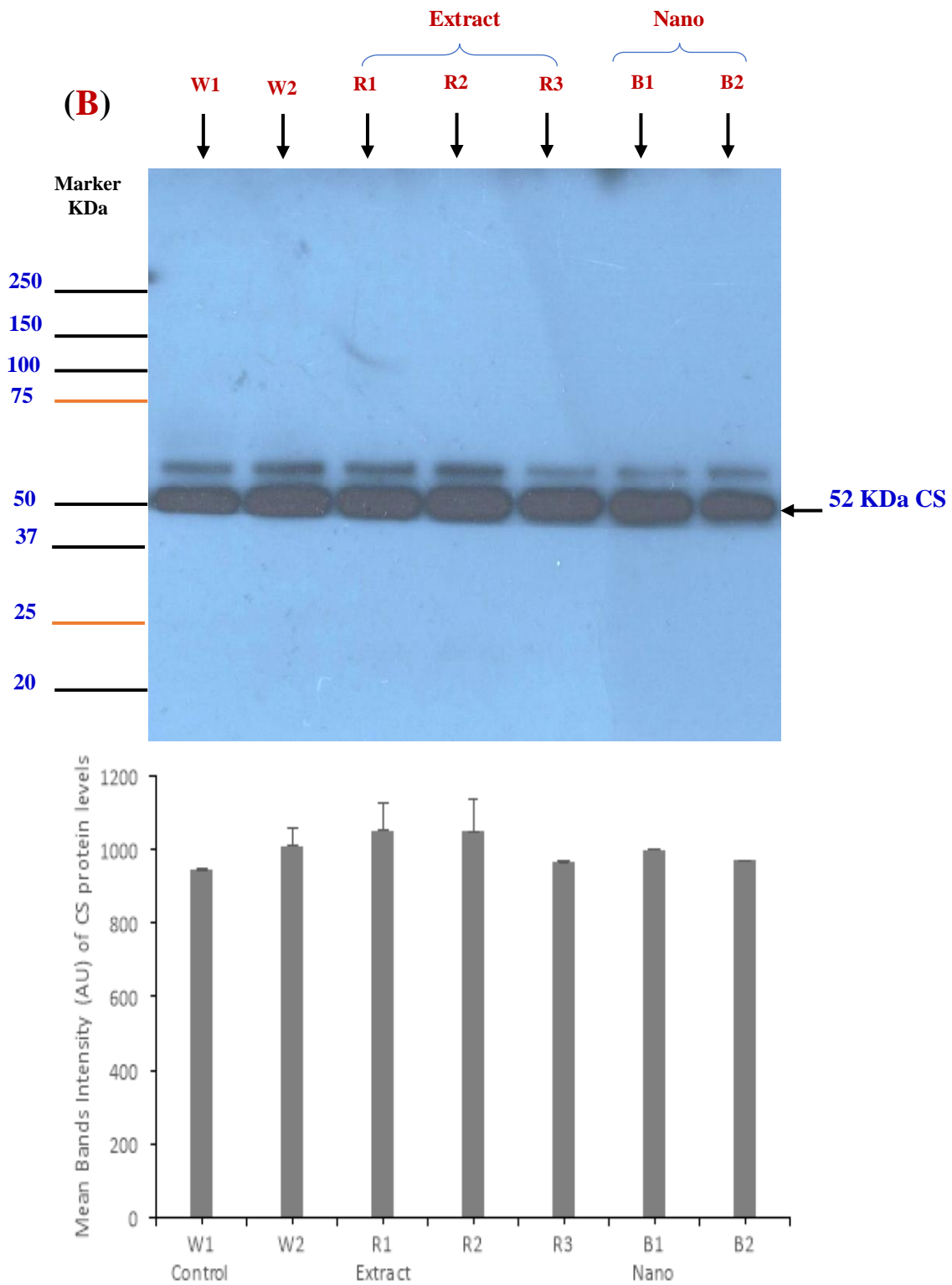
Western Blotting Assay

The results of the current study showed that there was a significant increase ($P < 0.002$) in expression of mitochondrial expression of SDHA protein in fenugreek extract group (819.67 ± 11.465) and fenugreek NP (1006 ± 13.537) groups compared with control (629.17 ± 26.308) group. A comparison between the result of the treatment groups revealed that there were significant elevation ($P < 0.0001$) in values of fenugreek NP group more than fenugreek extract group (Figure 2A).



For CS protein, insignificant variation was reported between values of fenugreek extract (1023 ± 52.009) and fenugreek NP (984.83 ± 20.726) groups when compared to value of control

group (977.67 ± 27.818) at $P < 0.485$ and $P < 0.809$, respectively. In comparison between the treatment groups, no significant differences ($P > 0.05$) were detected between values of fenugreek extract and fenugreek NP groups (Figure 2B).



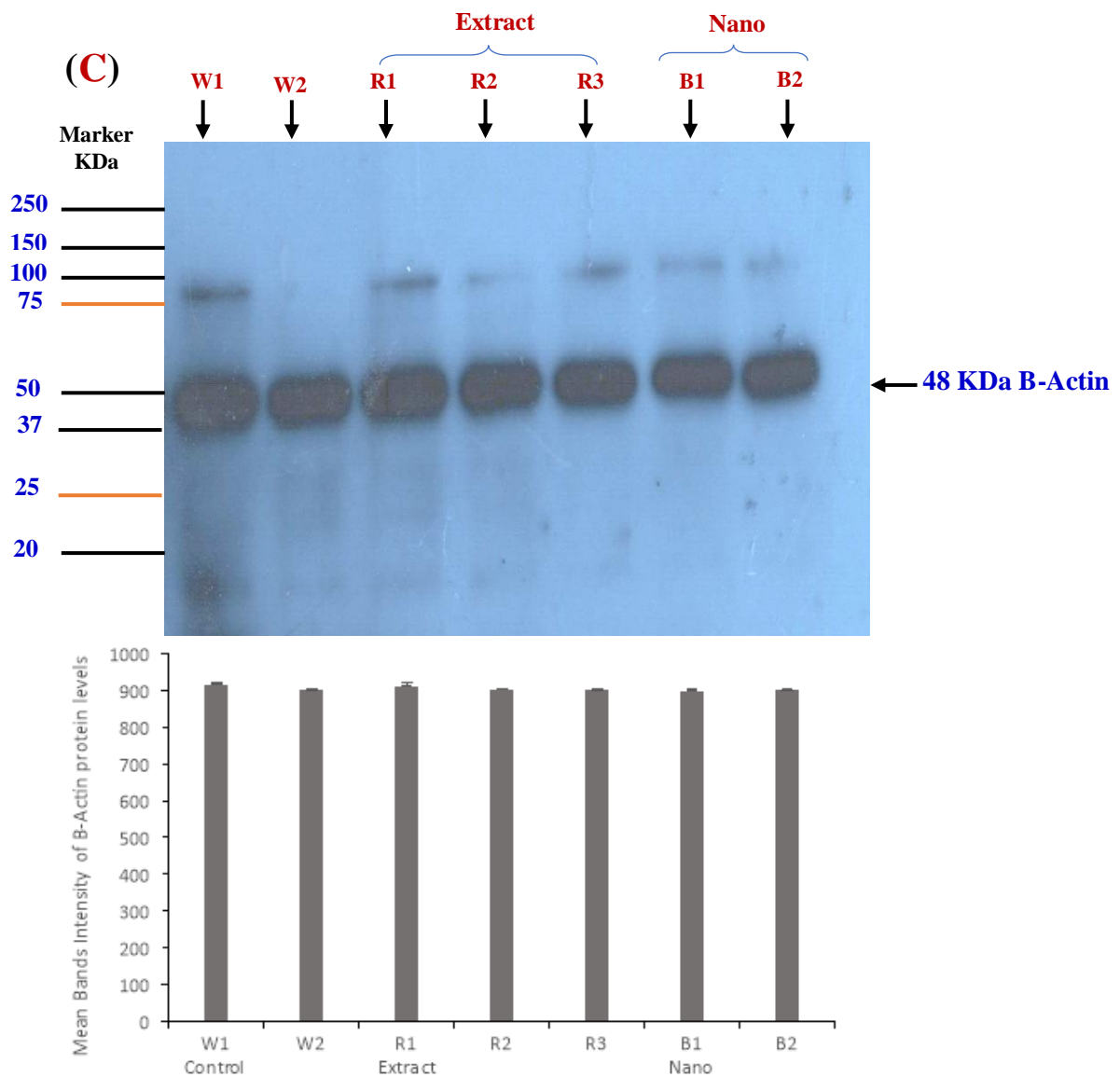


Figure (2): Mitochondrial expression of SDHA and CS proteins in treated group with extract (R1, R2, R3) or NPs (B1, B2) compared to control group (W1, W2) for epididymus of mature wister male rat during spermatogenesis.

(A): The image showing level of SDHA protein in fenugreek extract and fenugreek NPs groups compared to non treated control group. SDHA protein increased significantly in both treated group in compare with the control group. Moreover, the fenugreek NPs group showed a significant high increasing in level of SDHA compared with the fenugreek extract group

(B): The image showing the level of CS protein in the fenugreek extract and fenugreek NPs groups compared to non treated control group, in which, no significant changes were seen between

fenugreek extract, fenugreek NPs and control groups.

(C): The image showing the level of B-Actin as a housekeeping gene that presents the equalizing of pipetting error and non effecting of cell homeostasis during treated process.

Western blotting assay is a powerful tool to detect and characterize a multitude of proteins, especially those proteins that are of low abundance. It offers various specific advantages such that the wet membranes are pliable and are easy to handle compared to gels, easy accessibility of the proteins immobilized on the membrane to different ligands, only small amount of reagents are required for transfer analysis, multiple replicas of a gel are possible, prolonged storage of transferred patterns, prior to use, becomes possible, and the same protein transfer can be used for multiple successive analyses (Kurien and Scofield, 2015; Neris *et al.*, 2021).

The activity of spermatozoa mitochondria has been correlated with spermatozoa motility and thus with fertilization potential. Moreover, many studies demonstrated that the mitochondrial capsule localizes a variety of signaling proteins to the mid-piece and, thus, could play a critical role in spermatozoa function (Lu *et al.*, 2017; Wang *et al.*, 2020; Gallo *et al.*, 2021). Further analysis of the identified proteins with reference to their biological function showed that the vast majority of the identified proteins were engaged in cell metabolism and energy production; while the other identified proteins were involved in sperm tail structure/motility, protein metabolism, mitochondrial maintenance, spermatogenesis, sperm maturation, intracellular trafficking, and immunity (Nowicka-Bauer *et al.*, 2018). In this study, the findings of western blotting assay were supporting and confirmed results obtained by the BCA, in which, fenugreek extract and fenugreek NPs were participated significantly in increasing of SDHA protein but not CS protein that showed insignificant changes in their concentration among different study groups. Several experiments have been conducted in order to address these issues (Ibrahim *et al.*, 2019; Kaur and Sadwal, 2020; Mansour *et al.*, 2021). Jagtap *et al.* (2022) mentioned that fenugreek helps with reproductive problems and a variety of other disorders, and that the liquid extract of fenugreek increases sperm mortality and cation sperms channel proteins. Other study concluded that fenugreek seeds improved male sexual functioning, improved sexual life, and regulated orchid hormones and lactogen (Khanna *et al.*, 2021). Rao (2020) mentioned that the pharmacological actions of fenugreek are attributed to a diverse array of phytonutrients, with the main groups being steroidal saponins that thought to be responsible for androgenic effects and have been used previously as precursors for the synthetic production of steroid hormones. Worldwide, there was a number of animal and human studies introduced fenugreek extract as a food supplement boosting testosterone (Heufelder *et al.*, 2009; Aswar *et al.*, 2010; Hamden *et al.*, 2010; Nguyen *et al.*, 2017), but the results of these studies

are not consistent.

Torra-Massana *et al.* (2021) concluded that the sperm samples from patients with fertilization failure present altered abundance of different proteins, including mainly mitochondrial proteins. By using the western blotting assay, Italiano *et al.* (2012) identified the loss of SDHA protein expression in tumor cases whereas expression was retained in the non-tumor cases referring that these results were expected because mutations that resulted by tumor which lead to a truncated SDHA protein. The *SDHA* gene encodes the major catalytic subunit of the succinate dehydrogenase complex II; and the germline mutations in *SDHA* are associated with neurodegenerative diseases such as an early-onset encephalopathy, optic atrophy, ataxia and myopathy (Di Donato, 2009; Ghaoui and Sue, 2018).

Conclusion

This study provided new insights into the mitochondrial expression of SDHA and CS proteins using the Western Blotting and Bicinchonnic Acid assays. The administration of both fenugreek extract and fenugreek NPs were revealed a significant protein expression with in particular with using of fenugreek NPs that did not cause any side effects along the periods of the study. However, isolation of active components in fenugreek, and synthesizing of specific NPs based on one type of these components can provide new therapies with different characteristics. Also the augmentation of fenugreek NPs and proper regulation of steroidogenesis and mitochondrial biogenesis related genes is notably need to be furthermore studied.

Acknowledgments

The author is grateful to The Head and all staffs of the Department of Physiology, Biochemistry and Pharmacology (College of Veterinary Medicine, Al-Qasim Green University) for all facilities and helping in this work.

References

1. Agarwal, K., Trivedi, M., and Nirmalkar, N. (2022). Does salting-out effect nucleate nanobubbles in water: Spontaneous nucleation?. *Ultrasonics sonochemistry*, 82, 105860.
2. Aher, R. R., Belge, S. A., Kadam, S. R., Kharade, S. S., Misal, A. V., and Yeole, P. T. (2016). Therapeutic importance of fenugreek (*Trigonellafoenum-graecum L.*). A review. *J Plant Sci Res*, 3(1), 149.
3. Al-Gharban, H. A., and Al-Tae, H. S. (2016). Seroclinical diagnosis of *Anaplasma marginale* bacteria in carrier arabian one-humped camels. *Basrah J Vet Res*, 15, 346-359.
4. Al-Zamely A. N. and kshash M. J. 2021. Evaluation Effect of *Trigonella FoenumGraecum* (fenugreek) Seeds Nanoparticles On Male Reproductive Rats Efficiency. Al-Qasim Green Uni..

Vet. Med. College. 33-34.

5. Anifandis, G., Amiridis, G., Dafopoulos, K., Daponte, A., Dovolou, E., Gavriil, E., and Psarra, A. M. G. (2017). The in vitro impact of the herbicide roundup on human sperm motility and sperm mitochondria. *Toxics*, 6(1), 2.
6. Aswar, U., Bodhankar, S. L., Mohan, V., and Thakurdesai, P. A. (2010). Effect of furostanol glycosides from *Trigonella foenum-graecum* on the reproductive system of male albino rats. *Phytotherapy research*, 24(10), 1482-1488.
7. Bahrehmand Namaghi, I., and Vaziri, H. (2017). Sperm mitochondrial DNA deletion in Iranian infertiles with asthenozoospermia. *Andrologia*, 49(3), e12627.
8. Bansa, A., and Adeyemo, S. (2006). Phytochemical screening and antimicrobial assessment of *Abutilon mauritianum*, *Bacopa monnifera* and *Datura stramonium*. *Biokemistri*, 18(1).
9. Beyramzadeh, M., Dikmen, Z. G., Erturk, N. K., Tuncer, Z. S., and Akbiyik, F. (2017). Placental respiratory chain complex activities in high risk pregnancies. *The Journal of Maternal-Fetal and Neonatal Medicine*, 30(24), 2911-2917.
10. Chaudhary, S., Chaudhary, P. S., Chikara, S. K., Sharma, M. C., and Iriti, M. (2018). Review on fenugreek (*Trigonella foenum-graecum* L.) and its important secondary metabolite diosgenin. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 46(1), 22-31.
11. Di Donato, S. (2009). Multisystem manifestations of mitochondrial disorders. *Journal of neurology*, 256(5), 693-710.
12. Ferramosca, A., Lorenzetti, S., Di Giacomo, M., Lunetti, P., Murrieri, F., Capobianco, L., and Zara, V. (2021). Modulation of human sperm mitochondrial respiration efficiency by plant polyphenols. *Antioxidants*, 10(2), 217.
13. Fišar, Z., Hansíková, H., Křížová, J., Jiráček, R., Kitzlerová, E., Zvěřová, M., and Raboch, J. (2019). Activities of mitochondrial respiratory chain complexes in platelets of patients with Alzheimer's disease and depressive disorder. *Mitochondrion*, 48, 67-77.
14. Gallo, A., Esposito, M. C., Tosti, E., and Boni, R. (2021). Sperm motility, oxidative status, and mitochondrial activity: exploring correlation in different species. *Antioxidants*, 10(7), 1131.
15. Ghaoui, R., and Sue, C. M. (2018). Movement disorders in mitochondrial disease. *Journal of Neurology*, 265(5), 1230-1240.
16. Gharban, H. A., Al-Shaeli, S. J., Al-Fattli, H. H., and Altaee, M. N. (2019). Molecular and histopathological confirmation of clinically diagnosed lumpy skin disease in cattle, Baghdad Province of Iraq. *Veterinary world*, 12(11), 1827-1832.
17. Hamden, K., Jaouadi, B., Carreau, S., Aouidet, A., El-Fazaa, S., Gharbi, N., and Elfeki, A. (2010). Potential protective effect on key steroidogenesis and metabolic enzymes and sperm

- abnormalities by fenugreek steroids in testis and epididymis of surviving diabetic rats. *Archives of physiology and biochemistry*, 116(3), 146-155.
18. Hamza, A. A., Elwy, H. M., and Badawi, A. M. (2016). Fenugreek seed extract attenuates cisplatin-induced testicular damage in Wistar rats. *Andrologia*, 48(2), 211-221.
 19. Heufelder, A. E., Saad, F., Bunck, M. C., and Gooren, L. (2009). Fifty-two-week treatment with diet and exercise plus transdermal testosterone reverses the metabolic syndrome and improves glycemic control in men with newly diagnosed type 2 diabetes and subnormal plasma testosterone. *Journal of andrology*, 30(6), 726-733.
 20. Hu, M. (2015). The significance of nanotechnology in architectural design. *of Architectural Research*, 90.
 21. Ibrahim, N. A., Eid, B. M., Abd El-Aziz, E., and Abou Elmaaty, T. M. (2013). Functionalization of linen/cotton pigment prints using inorganic nano structure materials. *Carbohydrate polymers*, 97(2), 537-545.
 22. Italiano, A., Chen, C. L., Sung, Y. S., Singer, S., DeMatteo, R. P., LaQuaglia, M. P., and Antonescu, C. R. (2012). SDHA loss of function mutations in a subset of young adult wild-type gastrointestinal stromal tumors. *BMC cancer*, 12(1), 1-7.
 23. Jagtap, S., D Shejul, D., and Gawade, M. B. (2022). Trigonella foenum graecum (Fenugreek): An Herb with Impressive Health Benefits and Pharmacological Therapeutic Effects. *Asian Food Science Journal*, 19-28.
 24. Kang, W., Harada, Y., Yamatoya, K., Kawano, N., Kanai, S., Miyamoto, Y., and Miyado, K. (2020). Extra-mitochondrial citrate synthase initiates calcium oscillation and suppresses age-dependent sperm dysfunction. *Laboratory investigation*, 100(4), 583-595.
 25. Kaur, S., and Sadwal, S. (2020). Studies on the phyto-modulatory potential of fenugreek (Trigonella foenum-graecum) on bisphenol-A induced testicular damage in mice. *Andrologia*, 52(2), e13492.
 26. Khan, F., Negi, K., and Kumar, T. (2018). Effect of sprouted fenugreek seeds on various diseases: A review. *J. Diabetes Metab. Disord. Control*, 5, 119-125.
 27. Khan, I., Saeed, K., and Khan, I. (2019). Nanoparticles: Properties, applications and toxicities. *Arabian journal of chemistry*, 12(7), 908-931.
 28. Khanna, A., Thomas, J., John, F., Maliakel, B., and Krishnakumar, I. M. (2021). Safety and influence of a novel extract of fenugreek on healthy young women: a randomized, double-blinded, placebo-controlled study. *Clinical Phytoscience*, 7(1), 1-12.
 29. Kurien, B. T., and Scofield, R. H. (2015). Western blotting: an introduction. *Western Blotting*, 17-30.

30. Li, G., Luan, G., He, Y., Tie, F., Wang, Z., Suo, Y., and Wang, H. (2018). Polyphenol stilbenes from fenugreek (*Trigonella foenum-graecum* L.) seeds improve insulin sensitivity and mitochondrial function in 3T3-L1 adipocytes. *Oxidative Medicine and Cellular Longevity*, 2018.
31. Lu, A. X., Oh, H., Terrell, J. L., Bentley, W. E., and Raghavan, S. R. (2017). A new design for an artificial cell: polymer microcapsules with addressable inner compartments that can harbor biomolecules, colloids or microbial species. *Chemical science*, 8(10), 6893-6903.
32. Mansour, A. B., Abou Elghait, A., Abo-youssef, A., S Abdelwahab, N., and Helaly, H. (2021). A COMPARATIVE STUDY ON THE EFFECTS OF THE FENUGREEK SEEDS'POWDER AND ITS AQUEOUS AND OIL EXTRACTS ON THE MALE REPRODUCTIVE SYSTEM IN ALBINO RATS. *Bulletin of Pharmaceutical Sciences. Assiut*, 44(2), 593-605.
33. Neha, S., Anand, K., and Sunanda, P. (2019). Administration of fenugreek seed extract produces better effects in glibenclamide-induced inhibition in hepatic lipid peroxidation: an in vitro Study. *Chinese journal of integrative medicine*, 25(4), 278-284.
34. Neris, R. L. S., Dobles, A. M. C., and Gomes, A. V. (2021). Western blotting using in-gel protein labeling as a normalization control: advantages of stain-free technology. In *Proteomic Profiling Humana*, New York, NY. Pp: 443-456.
35. Nguyen, S. M., Ko, N. K., Sattar, A. S., Ipek, E. G., and Ali, S. (2017). Pulmonary embolism secondary to testosterone-enhancing herbal supplement use. *Cureus*, 9(8), 22-37.
36. Nowicka-Bauer, K., Lepczynski, A., Ozgo, M., Kamieniczna, M., Fraczek, M., Stanski, L., and Kurpisz, M. K. (2018). Sperm mitochondrial dysfunction and oxidative stress as possible reasons for isolated asthenozoospermia. *Journal of Physiology and Pharmacology*, 69(3).
37. Rao, A. (2020). An investigation of the effects of *Trigonella foenum-graecum* L.(Fabaceae)(fenugreek) extract on testosterone concentrations in men and women (Doctoral dissertation, University of Sydney).
38. Sadogh, A., Gorji, N., and Moeini, R. (2022). Herbal foodstuffs in Avicenna's recommended diet to improve sperm quality and increase male fertility; an evidence-based approach. *Journal of Complementary and Integrative Medicine*, 19(1), 47-70.
39. Sedha, S., Kumar, S., and Shukla, S. (2015). Role of oxidative stress in male reproductive dysfunctions with reference to phthalate compounds. *Urology journal*, 12(5), 2304-2316.
40. Shafiei, M., Jafarizadeh-Malmiri, H., and Rezaei, M. (2019). Biological activities of chitosan and prepared chitosan-tripolyphosphate nanoparticles using ionic gelation method against various pathogenic bacteria and fungi strains. *Biologia*, 74(11), 1561-1568.
41. Simonazzi, A., Cid, A. G., Villegas, M., Romero, A. I., Palma, S. D., and Bermúdez, J. M.

- (2018). Nanotechnology applications in drug controlled release. In *Drug targeting and stimuli sensitive drug delivery systems* (pp. 81-116). William Andrew Publishing.
42. Singh, A., Sarkar, D., and Singh, S. K. (2022). Effect of *Trigonella foenum-graecum* L. seed extract on the reproductive system of male mice and possible mechanism of its action on spermatogenesis. *Andrologia*, e14429.
43. Soren, S., Vir Singh, S., and Singh, P. (2018). Seasonal variation of mitochondria activity related and heat shock protein genes in spermatozoa of Karan Fries bulls in tropical climate. *Biological Rhythm Research*, 49(3), 366-381.
44. Srinivasa, U. M., and Naidu, M. M. (2021). Fenugreek (*Trigonella foenum-graecum* L.) seed: promising source of nutraceutical. *Studies in Natural Products Chemistry*, 71, 141-184.
45. Torra-Massana, M., Jodar, M., Barragán, M., Soler-Ventura, A., Delgado-Dueñas, D., Rodríguez, A., and Vassena, R. (2021). Altered mitochondrial function in spermatozoa from patients with repetitive fertilization failure after ICSI revealed by proteomics. *Andrology*, 9(4), 1192-1204.
46. Wang, X., Lv, C., Guo, Y., and Yuan, S. (2020). Mitochondria associated germinal structures in spermatogenesis: piRNA pathway regulation and beyond. *Cells*, 9(2), 399.
47. Wani, S. A., and Kumar, P. (2018). Fenugreek: A review on its nutraceutical properties and utilization in various food products. *Journal of the Saudi Society of Agricultural Sciences*, 17(2), 97-106.
48. Zhou, C., Qin, Y., Chen, R., Gao, F., Zhang, J., and Lu, F. (2020). Fenugreek attenuates obesity-induced inflammation and improves insulin resistance through downregulation of iRhom2/TACE. *Life Sciences*, 258, 118222.
49. Zhu, Z., Kawai, T., Umehara, T., Hoque, S. M., Zeng, W., and Shimada, M. (2019). Negative effects of ROS generated during linear sperm motility on gene expression and ATP generation in boar sperm mitochondria. *Free Radical Biology and Medicine*, 141, 159-171.