Study On Microflora In Freshwater Grass Carp (Ctenopharyngodon Idella)

K.Sundaravalli¹,K.Revathi^{2*}, R.Devi², A.Anitha²

¹ Meenakshi Academy of Higher Education and Research, KK Nagar, Chennai 600078, India *Corresponding author Email: reva63@rediffmail.com

Abstract

Increasing threat of pollution in recent times may have a direct or indirect effect on the microbial flora of fish which in turn may cause a threat to human health due to frequent consumption of the fish. Grass carp (*Ctenopharyngodonidella*) were collected and analyzed for the presence of different microbial organisms from their gut content. Microbial counts were analyzed in the fish skin, gills and gut. The intestinal gut showed the highest biome of bacteria. *Pseudomonas spp., Aeromonas spp., Salmonella spp.,* and *Clostridium botulinum* were identified in 55%, 25%, 12% and 8% respectively from the gut biome. We found the occurrence of human pathogenic *Pseudomonas spp* the gut of grass carp were confirmed to be in higher count, which may affect the human health on consumption. The result analyzed were found to be statistically significant P<0.05.

1.Introduction

Human beings consume fish which acts as an extensive source of protein. Yet, it may cause serious effect in human on consumption of contaminated or spoilt fish due to existence of pathogenic and opportunistic microorganisms (1). There is an utmost health risk for consumers since food borne pathogens may result in serious disease outbreaks. Appearance of such causative food borne pathogens may be due to external sources like environmental pollution, water quality and damage caused by human, animal or plants (2). Determining the micro flora in the different parts of fish organ is vital to understand the contamination and quality of fish. Contamination is also associated with rigorous microbial growth which may detoriate the quality of fish which has natural habitat may harbor abundance of microorganisms especially bacteria (3). Settlement of bacteria in the gill and skin is due to continual subjection to infected water whereas the alimentary canal infection may happen due to contaminated feed and water. Infection in the muscle of fish may also occur if the immunological resistance is low in the fish (4). Antibiotic resistant bacteria associated with food chains have been found to cause untreatable, prolonged and complicated infections and sometimes death. Low economic, retarded sanitation, lack of hygiene and use of broad spectrum of antibiotic has been a bane with potential adverse effect in the population of fish found in the fresh waters (5).

Overcrowding of fish population and opportunistic contamination with pathogenic bacteria result with close interaction between pathogenic bacterial contaminations. It is therefore mandatory to analyze the ubiquity of pathogens present in fish which remains to be the major food source for a wide population (6). Study on the bacterial pathogens present in skin and gills are prevalent whereas infection in gut becomes a necessity to analyze and study since it may contaminate the muscles of the fish(4). Our study focuses on the analyzing the existence of varied pathogens in the gut of routinely consumed fish named Grass carp

Ctenopharyngodonidella. The Glass carp is highly cultivated freshwater fish species in the world (7). There are more than 34 different types of bacteria including 125 different bacterial species found to associate with the fish disease in the world (8). This study was conducted to enlighten the bacterial infection in the gut region of the glass carp.

2.Materials and Methods

2.1 Study sample collection

The Green carp fishes were collected from Poondi reservoir and were collected in a sterile polythene bag. The collected fish were of various body weight (50gms to 250gms) and length (10cm to 30 cm). The collected fishes were transferred to Central Laboratory of MAHER for further microbiological analysis.



Grass carp collected from Poondi Lake

2.2 Clinical Examination

The collected fishes were examined clinically to detect any abnormality in external changes of the fish. The fish was dissected aseptically using sterile scalpel and forceps. Different parts of fish were dissected and cut into pieces to obtain 10 grams and were homogenized. Homogenized samples were examined for the presence of Microbial contents.

2.3 Postmortem examination

Gill and Skin: For examination in Gill and skin, the fish was washed using double distilled water to remove the unwanted debris. A sterile Cotton swab was rubbed against the samples and inoculated in nutrient broth and MacConkey agar broth.

Gut and muscles: The dissected and cut samples were performed with the help of scalpel and further inoculated aseptically in the nutrient and MacConkey broth.

2.4 Isolation and Identification of bacteria:

Isolation and identification procedure was followed according to *Cheesbourgh 1984* .MPN technique was followed to enumerate the microbial counts.MPN technique was preferred since it is effective for even low population levels than Plate count method. This quantitative analysis was performed in order to project the count of bacteria. 10⁻⁷ dilutions were taken. The quantity of 10 microlitre of sample was poured into the Nutrient agar, MacConkey agar, Salmonella /Shigella agar. After pouring the samples spread plate method is followed. All the plates are marked and placed at 37°C for 24-48 hrs incubation. After incubation bacterial isolates were identified by their morphology and cultural characters.

2.5 Statistical analysis

Statistical analysis was performed using independent T test. The result were found to be statistically significant P < 0.05

3.Results

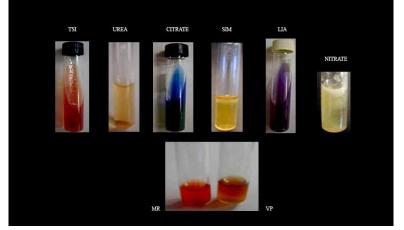
Bacterial infections and diseases are serious problem in aquaculture. Bacterial pathogens are common in fish associating the natural surroundings, during certain environmental factors fish mortalities are caused. *Pseudomonas* infection is implicated highly among the freshwater fish diseases. Pseudomonas sp., Aeromonas sp., Salmonella sp., and Clostridium sp., were isolated from the organs of *C.idella* collected from Poondi Lake. All these four organisms are said to affect in humans on consumption of bacterial pathogen infected fish due to ignorance or lack of awareness. The bacterial count values were obtained for three different dilution factors such as 10⁻³, 10⁻⁵, and 10⁻⁷. Table 1, Table 2 and Table 3 shows the counts of Pseudomonas sp., Aeromonas sp., Salmonella sp., and Clostridium sp., in 10⁻³, 10⁻⁵, 10⁻⁷ dilution factor respectively in three different organs namely Skin, Gill and Gut . Among these 10⁻⁷ dilution factor showed values feasible for counting which was further taken for replicate values. Three replicate counting were done for the dilution factor of 10^{-7} and named as R1. R2, R3 and the mean value was obtained. The *Pseudomonas* sp., overall showed higher counts than the other organisms. The salmonella sp., and clostridium sp., was comparatively high in the Skin of fish yet not more than the value of Pseudomonas sp., Aeromonas sp., expressed elevated counts in Gill than that of Skin and Gut however not more than the Pseudomonas sp., On the whole Pseudomonas sp., were found to be abundant in all the organs of the fish. The resultswere analyzed with independent T test and the results were found to be statistically significant (P>0.05)



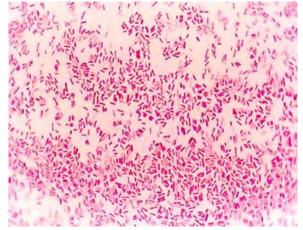
Clostridium species



Salmonella in Mac Conkey agar



Biochemical tests of pseudomonas sp.



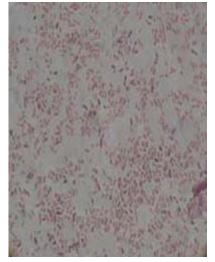
Gram stained Pseudomonas sp.



Pseudomonas in Cetrimide agar



Clostridium sp., in Blood agar



Gram stained Aeromonas sp.,

Table: 1 Bacterial count of Pseudomonas sp., Aeromonas sp., Salmonella sp., andClostridium sp., in Dilution factor 10-3

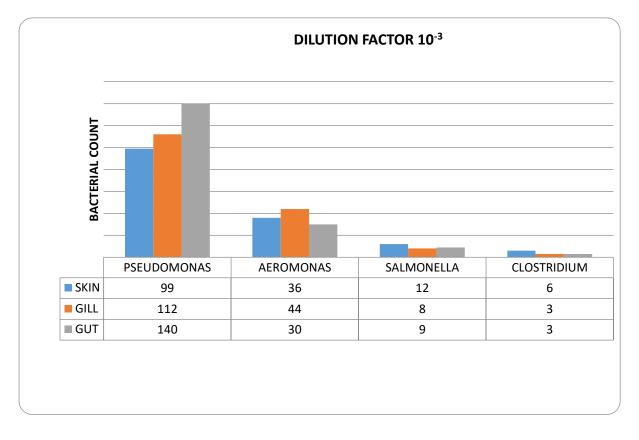


Table : 2 Bacterial counts of Pseudomonas sp., Aeromonas sp., Salmonella sp., andClostridium sp., in Dilution factor 10⁻⁵

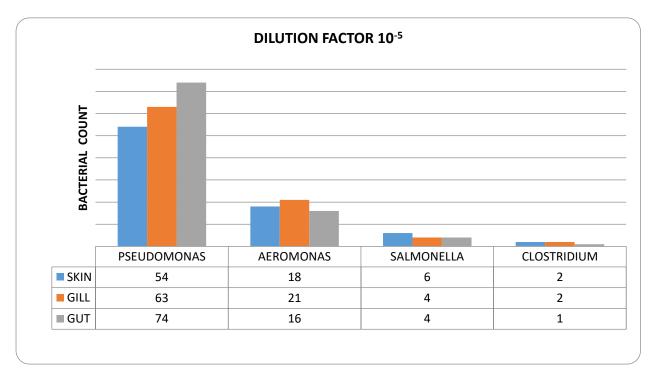
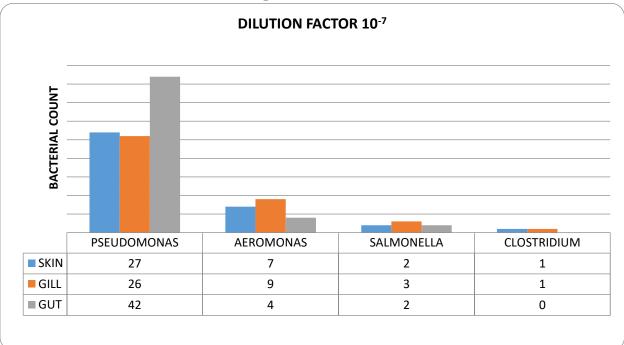


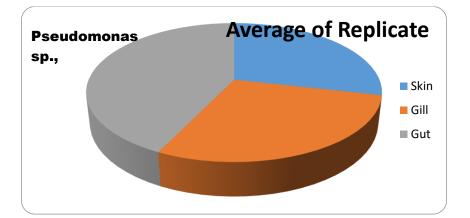
Table : 3 Bacterial counts of Pseudomonas sp., Aeromonas sp., Salmonella sp., andClostridium sp., in Dilution factor 10-7



	Pseudomonas sp.,			
Parts	R1	R2	R3	Average
of				
fish				
Skin	20	23	27	23.33333

Gill	29	24	26	26.33333
Gut	38	27	42	35.66667

Table 4 : Replicates of dilution factor 10⁻⁷ in pseudomonas sp.,



	Ae			
Parts of	R1	Average		
fish				
Skin	7	6	8	7
Gill	8	7	6	7
Gut	3	3	2	2.666667

Table 5 : Replicates of dilution factor 10⁻⁷ in Aeromonas sp.,

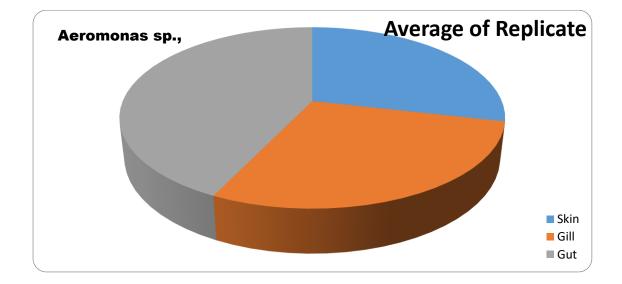
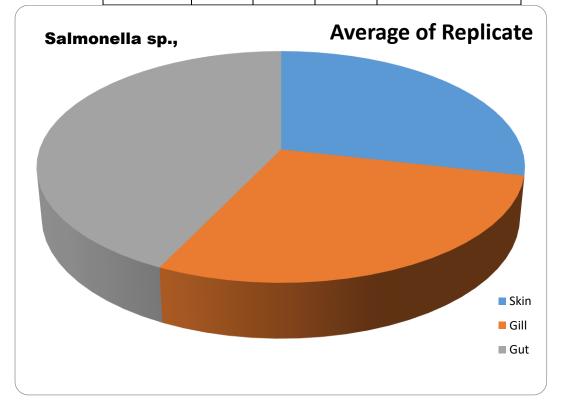


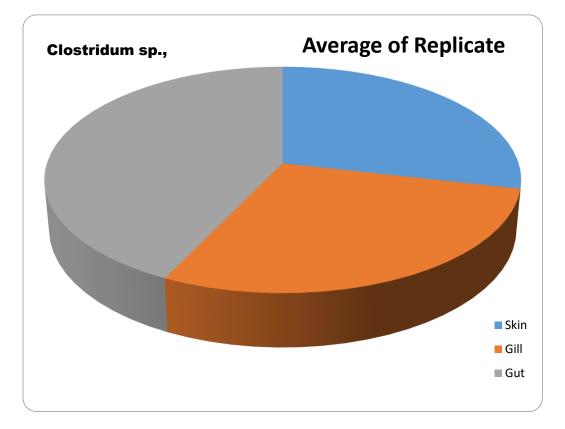
Table 6 : Replicates of dilution factor 10⁻⁷ in Salmonella sp.,

Parts of	R1	R2	R3	Average
fish				
Skin	2	3	3	2.666667
Gill	3	2	2	2.333333
Gut	2	1	0	1



,	Table 7 : Replicates of dilution factor 10 ⁻	⁷ in	Clostridium sp.,

	Clostridium sp	••			
Parts of fish	R1	R2	R3	Average	
Skin	1	1	0	0.666667	
Gill	0	1	1	0.666667	
Gut	1	1	1	1	



4.Discussion

Fish plays a vital role as a major dietary protein source yet they are prone to infection to microbes due to their polluted living environment especially in recent days. It also plays a role in the healthy living of humans who consume it regularly. Grass carp available commonly in local markets and freshwater pond was examined for the presence of microbe, in particular the bacterial flora which is a threat to human on consumption with ignorance(9).

Pseudomonas is a major organism affecting the fish of various species.*Pseudomonas* shows resilience surviving in brackish water and also in the temperatures below 50°F (10°C) and above 100°F (38°C) (10). *Pseudomonas* and *Aeromonas* are major pathogens affecting fish. dropsy, hemorrhagic septicemia, gill rot disease, fin and tail rot diseases(11). Ulcerative syndromes are caused by these microbes in fish (12).*Clostridium* species causes dangerous diseases namely necrotic enteritis to wound infection and gas gangrene which threatens life majorly(13).Gastro intestinal problems are to be caused by *clostridium* species and its high toxic occurrence in fresh fish is proved (14). Total of 1.3 billion cases of Human gastroenteritis annually were recorded which were due to the ingestion of Fish and Shell fish affected by *Salmonellas*p(15). Seepage of Sewage water into the fresh lakes or fish farm may contaminate the water and the fish gradually(16). Fish contamination could also happen through improper transportation or improper handling or washing with contaminated water.

5.Conclusion

The current study demonstrated the fundamental basis of bacterial infection in grass carp, this will help as an initial point for the diagnosis of bacterial infections, prevention and control of pathogenic bacterial infections in the poondi reservoir region. *Pseudomonas* and

*Aeromonas*were found to be highly prominent microbial agents affecting the grass crap. This could be further implicated with pathogenic infection of human after consumption of the fish leading to disease complication and mortalities.

Competing interest

The author declares no conflict of interest.

6.References

1. Bintsis T. Foodborne pathogens. AIMS Microbiol. 2017;3(3):529-63.

2. Alegbeleye OO, Singleton I, Sant'Ana AS. Sources and contamination routes of microbial pathogens to fresh produce during field cultivation: A review. Food Microbiol. 2018;73:177-208.

3. Novoslavskij A, Terentjeva M, Eizenberga I, Valciņa O, Bartkevičs V, Bērziņš A. Major foodborne pathogens in fish and fish products: a review. Annals of Microbiology. 2016;66(1):1-15.

4. Overstreet R.M. HWE. Diseases and Mortalities of Fishes and Other Animals in The Gulf of Mexico. In: Ward C. (eds) Habitats and Biota of the Gulf of Mexico: Before the Deepwater Horizon Oil Spill. Springer, New York, NY. 2017;978(1):4939-3456.

5. Manyi-Loh C, Mamphweli S, Meyer E, Okoh A. Antibiotic Use in Agriculture and Its Consequential Resistance in Environmental Sources: Potential Public Health Implications. Molecules. 2018;23(4):795.

6. Kousar R, Shafi N, Andleeb S, Ali NM, Akhtar T, Khalid S. Assessment and incidence of fish associated bacterial pathogens at hatcheries of Azad Kashmir, Pakistan. Brazilian Journal of Biology. 2020;80:607-14.

7. Awatef H. Hamouda EMM. Insight on Prevailing Bacterial Diseases Affecting Grass carp (Ctenopharyngodon idella) at Aswan Fish Hatchery, Egypt. AJVS. 2020;64(1):5-16.

8. Sudheesh PS, Al-Ghabshi A, Al-Mazrooei N, Al-Habsi S. Comparative Pathogenomics of Bacteria Causing Infectious Diseases in Fish. International Journal of Evolutionary Biology. 2012;2012:457264.

9. Sheng L, Wang L. The microbial safety of fish and fish products: Recent advances in understanding its significance, contamination sources, and control strategies. Comprehensive Reviews in Food Science and Food Safety. 2021;20(1):738-86.

10. Moradali MF, Ghods S, Rehm BHA. Pseudomonas aeruginosa Lifestyle: A Paradigm for Adaptation, Survival, and Persistence. Front Cell Infect Microbiol. 2017;7:39-.

11. Mandal KsR. Fresh water fish diseases in west bengal, India. international journal of fisheries and aquatic studies. 2018;6(5):356-62.

12. Li Z, Wang G, Zhang K, Gong W, Yu E, Tian J, et al. Epizootic ulcerative syndrome causes cutaneous dysbacteriosis in hybrid snakehead (Channa maculata $\Im \times$ Channa argus \Im). PeerJ. 2019;7:e6674-e.

13. Uzal FA, Freedman JC, Shrestha A, Theoret JR, Garcia J, Awad MM, et al. Towards an understanding of the role of Clostridium perfringens toxins in human and animal disease. Future Microbiol. 2014;9(3):361-77.

14. Mariel A. Marlow PhD M, Ruth E. Luna-Gierke MPH, Patricia M. Griffin MD, and Antonio R. Vieira DVM, PhD, MPH. Foodborne Disease Outbreaks in Correctional Institutions—United States, 1998–2014. American journal of public health.

15. Heinitz ML, Ruble RD, Wagner DE, Tatini SR. Incidence of Salmonella in fish and seafood. Journal of food protection. 2000;63(5):579-92.

16. Sasakova N, Gregova G, Takacova D, Mojzisova J, Papajova I, Venglovsky J, et al. Pollution of Surface and Ground Water by Sources Related to Agricultural Activities. Frontiers in Sustainable Food Systems. 2018;2(42).