

Histopathological Study Post *Helicobacter Pylori* Infection in Mice

*¹Aseel I. Ibrahim, ²Zainab I. Ibrahim, ²Zainab Jamal M. Jawad

Author Affiliations:

¹ Clinical laboratory Science department/ college of Pharmacy/ University of Baghdad

² department pathology and poultry disease/ college of veterinary medicine/ university of Baghdad

Abstract:

Helicobacter pylori is the cause of ulcers in the gastrointestinal tract such as gastric and peptic ulcers besides the malignancy of gastric carcinoma, lymphoma and non-gastric diseases.

The histopathological changes that occurred in different organs of mice post *H. pylori* infection were investigated. The bacterial isolate was taken from a patient with duodenal ulcer. The result tissue section revealed polymorphonuclear cells (PMNs) and lymphocytes infiltrated the layers of kidney, liver, and heart with degeneration.

H. pylori has a significant severe role in the incidence of the inflammatory reaction in the stomach.

Keyword: *Helicobacter pylori*, Rapid urease test, infection, diagnosis.

Introduction:

Most disorders of *Helicobacter pylori* infection are gastric or duodenal ulcers, which occur where their tissues lose the protection (Wang *et al.*, 2014), mechanism to dispose of acids & pepsin enzymes. Gastric ulcers occur when the mucosa of the stomach or duodenum is very weak for the protection.

Gastric ulcers mean inflammation of the stomach lining (gastritis) & perforation occurs when the ulcer is untreated or treated weak (Naito & Yoshikawa, 2002).

H. pylori produces urease enzyme which causes the release of free radicals that cause damage to the epithelium neither invading the cells of surrounding tissues & nor developing the immunity for repeated infection (Martin, 2005). *H. pylori* is represented as a major etiological factor of gastritis & duodenal ulcers (NIH Consensus conference, 1994; Franceschi, 2002). The association between *H. pylori* & causes of cancer was fixed as a carcinogen by the International Agency for Research on Cancer (IARC, 1994).

H. pylori colonizes epithelial cells of gastric mucosa and the infection was limited to the stomach, duodenum and esophagus (Graham, 2014).

Experimental procedure:

1- Bacterial isolation & identification:

A- The special media for isolation of *H. pylori* is Skirrow agar with the antibiotics (polymyxin B, Vancomycin, Trimethoprim B, & Amphotericin) (Dent *et al.* 1988).

B- Bacterial isolate is taken from biopsies of duodenal ulcer patient in the Educational Baghdad hospital. Two histological biopsies were taken, one used for bacterial culture with Skirrow media then transferred to microaerophilic conditions with gas-generating kit from Oxoid for 3-7 days at 37°C; the second biopsy used for rapid urease test by liquid urea medium prepared by Marshall (Marshall *et al.* 1987).

C- *H. pylori* diagnosed according to Bergey's classification (Holt *et al.* 1994); as follows:

- i. Microorganism examination: the smear of bacterial colonies stained with gram stain & examined by light microscope.
- ii. The motility of bacteria was examined by suspension drop with light microscope.
- iii. Biochemical tests: used catalase & oxidase test.
- iv. Sensitivity test for *cephalothin* & naldixic acid.

D- *H. pylori* Diagnosed in the biopsies was done by:

- i. Direct smear of biopsies according to (Montgomery *et al.* 1988).
- ii. Rapid urease test: the color of media was change from yellow to red through 5-10 min due to highly activity of urease enzyme.

2- Bacterial suspension for inoculation; prepared to contain 10^9 bacterial cells of *H. pylori* per 1ml of physiological phosphate buffer (Wang *et al.* 1997).

3-histopathological examination:

- i. Thirty of white male mice BALB/CMusMusculus, aged between 8-14 weeks & weight about 250-350 gm, healthy and good management from light to temperature available for them.
- ii. All animals were infected orally with 0.1ml of bacterial suspension which contains 10^9 bacterial cells/ml for 3 times, dose for each two days, and then the animals were monitored along periods of 3 & 4 weeks of experiment.
- iii. At 4th week post infection the histological samples were taken from the animals after killing, the samples are the liver, kidney & heart preserved in 10% formalin as fixative to prepare histopathological sections was done according to Guyer instructions (Guyer, 1953).

Results:

H. pylori isolated from laboratory Animals match the patient isolates.

Histopathological examined of the infected organs (kidney, liver, & heart) through the first three weeks demonstrated degenerative changes in the epithelial cells with progression of inflammatory response & infiltration of lymphocytes in all the examined tissues. In fourth week, viewed lesions characterized by intercellular swelling clearly in the renal tubular epithelium cells (Figure 1) & hepatocytes (figure 2, 3) to vacuolar degeneration of the cardiac muscles (Figure 4) & perivascular mononuclear cells cuffing, also few infiltration in the parenchyma, congestion & dilatation of blood vessels (Figure 5).

Discussion:

The *H. pylori* bacteria can cause histopathological changes in non-stomach tissues especially in the kidney, liver & heart, although their main role in infection of the stomach by causing gastric, peptic, duodenal ulcer & gastric cancer. These changes may attribute to virulence of two factors, Vacuating cytotoxin A (VacA) and Cytotoxin-associated gene (CagA) produced by *H. pylori*.

Vacuating cytotoxin A is pore-forming toxin, secreted by *H. pylori* caused increased in plasma membrane permeability, changes in the structure of endosome, mitochondrial membrane permeability and then cell death (Jones *et al.*, 2010), while CagA (Cytotoxin-associated gene A) is the second protein produced by *H. pylori* transfer from it into host cells by type IV secretion system (Terradot and Waksman, 2011). These two proteins are determined the pathogenesis of *H. pylori* (Figueiredo *et al.*, 2005).

In kidney tissue, show vacuolar degeneration (star shaped) with infiltration of lymphocytes within glomerulus, some studies showed correlation between chronic kidney disease and *H. pylori* prevalence (Ganji and Rafieian, 2017), there's one study has not supported the correlation between *H. pylori* presence and the kidney infection (Wijarnpreecha *et al.*, 2017), but our study agreement with Ganji and Rafieian study when showed agreement between *H. pylori* and kidney infection.

In the liver tissue, show vacuolar degeneration & leukocyte cuffing, vascular dilatation, enlargement of hepatocytes, depression of the sinusoids. *H. pylori* inoculated orally may reached the hepatocytes and cause inflammation as etiological factor (Huang *et al.*, 2009).

Rapid urease test is the most useful tool has used in our study for *H. pylori* diagnosis faster than other tools, rapid urease test had specificity and sensitivity reached 90%, expensive, and is the only test was performed within few minutes directly after collect biopsy specimen by endoscopy (Atkinson & Braden, 2016; Al-Rubai, A., 2006). One study reports the lowering levels of fasting blood ammonia in *H. pylori* infection than without *H. pylori* infection in cirrhosis case of the liver organ (Kini *et al.* 2001). Some studies depend on rapid urease test for evaluated the status of *H. pylori* (Al-Rubai, A., 2006; Vasconez *et al.* 1999).

In the heart tissue, show thrombosis, perivascular leukocyte cuffing & edema with vacuolar degeneration of cardiac muscle fibers.

Diseases of the heart like: ischemic heart disease, myocardial infarction, atherosclerosis, coronary artery disease, in addition to *H. pylori* has represents one of etiological factor of heart disease by releasing two main toxigenic nutrients, vacuolating associated gene (Vac A) and cytotoxic-associated gene A (Cag A), Cytotoxin-associated gene A (Cag A) is the most virulence factor participating in formation of cholesterol patches in the arteries causes autoimmune disorder, and started the immune response (Jamkhande *et al.* 2016). *H. pylori* adhesion to the host cell caused inflammation, cellular damage, and increases the release of most important virulence factors for *H. pylori*: Cag A and Vac A and found that *H. pylori* strains positive to Cag A are associated with heart disease more than strains negative to Cag A. *H. pylori* strains positive to Cag A increases the activities of cyclooxygenase-1 and 2 in the endothelial cells of the blood vessels. Inflammation caused by Cag A stimulate release many cytokines like IL-1 to IL-12, monocytes, macrophages (like tumor necrosis factor α (TNF- α)), T and B lymphocytes, then causes heart disease or shock, and autoimmune reaction among anti-Cag A antibodies and the vascular wall antigens, this suggested that antibodies participated in activation the inflammatory cells within atherosclerosis lesions (Lee *et al.*, 2018; Al-Quarashi & Hodhod, 2012).

Burden of *H. pylori* is to contain protein similar to the heat shock protein-60 that present on the surface of endothelial cells in the artery. Immune cross reaction occurs between human and bacterial heat shock protein-60 due to the immune response to *H. pylori* which in turns causes autoimmune reaction and local inflammation in the artery (Sulewska, 2004). Another speculation about *H. pylori* role in development of atherosclerotic plaque, has found that bacterial deoxyribonucleic acid in the artery wall forms patches of infection, which results in heart disease (Sulewska, 2004). The role of *H. pylori* in many diseases depends on the basis of rapid urease test.

Infection of *H. pylori* play role in progression of vascular disease (Lee *et al.*, 2018; Elkind & Cole, 2006). Seroepidemiological and eradication studies demonstrate the relationship between atherosclerosis and the infection with *H. pylori* (Ando *et al.*, 2006).

Reached *H. pylori* and their biochemical contain from the mucosa of the stomach to the circulation (Guo *et al.*, 2007) are in agreement with the exposure of endothelium to virulence factors secreted by *H. pylori*. In atherosclerosis plaque sites, these factors reach to high levels in the microenvironment of the artery wall (systemic circulation) and cause endothelial dysfunction & lesion development (this may be attributed to Vac A, that altering intercellular vesicular causing formation of vacuole (Reyrate *et al.*, 1999).

Conclusion:

The significant pathologic infection of isolated *H. pylori* in mice. Anti-*H. pylori* antibodies cross react with antigens of erythrocyte membrane in the human, which probably a guide for the relationship between infection with *H. pylori* and vascular disorders. Successful *H. pylori* eradication led to a reduction in the platelet activation.

Acknowledgment:

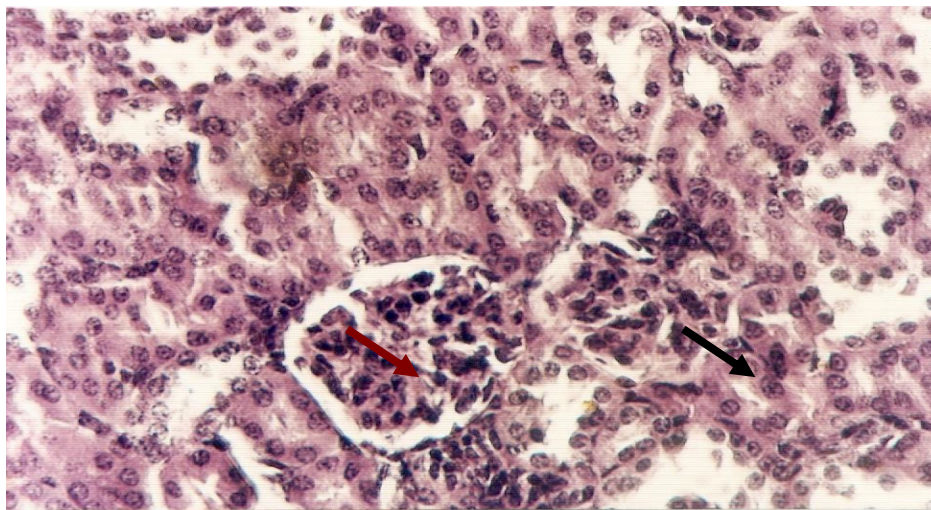
I would like to thank Dr. Yasser in the Endoscopy unit in the Al-Educational Baghdad Hospital for his facilitate the task of collecting biopsies samples, also I would like to thank Assistant prof. Dr. Zainab Ismail Ibrahim in the Diseases and Poultry department/ veterinary collage/university of Baghdad for her efforts in reading histological sections and reviewing the research, I also thank all employees in the Endoscopy unit of the hospital.

References:

1. **Al-Rubai, A.I., 2006.** The role of lipopolysaccharide in pathogenesis of *Helicobacter pylori* isolated from patients suffering from duodenal ulcer. College of Science, University of Baghdad. M.Sc. Thesis.
2. **Al-Quarashi A.M., Hodhod T.E (2013).** The association of Cag A-positive *Helicobacter pylori* and atherosclerosis in Najran area, Saudi Arabia. *J Am Sci.* 9:356–361.
3. **Ando, T., Minami, M., Ishiguro, K., Maeda, O., Watanabe, O., Mizuno, T., Fujita, T., Takahashi, H., Noshiro, M. & Goto, H. (2006).** Changes in biochemical parameters related to atherosclerosis after *Helicobacter pylori* eradication. *Aliment. Pharmacol. Ther.* 24, Suppl; 4: 58–64.
4. **Atkinson NS, Braden B (2016).** *Helicobacter Pylori* Infection: Diagnostic Strategies in Primary Diagnosis and After Therapy. *Dig Dis Sci.*; 61(1):19-24.
5. **Dent, J.C. and McNulty, C.A.M. (1988).** Evaluation of new selective media for *Campylobacter pylori*. *Eur. J. Clin. Microbiol. Infect. Dis.* 7:555-68.
6. **Dr. Martin Blaser. (2005),** In endangered species in the stomach. *Scientific American*; Feb. P: 38-45.
7. **Elkind MS, Cole JW. (2006).** Do common infections cause stroke? *Semin Neurol*; 26: 88–99.
8. **Figueiredo C, Machado JC, Yamaoka Y. (2005).** Pathogenesis of *Helicobacter pylori* infection. *Helicobacter*; 10: 14-20.
9. **Franceschi, F., Genta, R.M. & Sepulveda, A.R. (2002).** Gastric mucosa. Long-term outcome after cure of *Helicobacter pylori* infection. *J. Gastroenterol.* 37(suppl.13), 17-23.
10. **Ganji-Arjenaki, M. & Rafieian-Kopaei, M. (2017).** Chronic Kidney Disease and *H. pylori* Prevalence: A Significant Association? *Dig. Dis. Sci.*; 62:2053.
11. **Graham DY. (2014).** History of *Helicobacter pylori*, duodenal ulcer, gastric ulcer & gastric cancer. *World J. Gastroenterol.* 14; 20(18):615-625.

12. **Guo, F.H., Yan, X.M., Fan, C.X., Zhao, F., Xiao, D., Zeng, X., Zhang, M.J., He, L.H., Meng, F.L.& Zhang, J.Z.(2007)**, Cross-reactivity of anti-*H. pylori* antibodies with membrane antigens of human erythrocytes. *World J. Gastroenterol.*;13:3742-46.
13. **Guyer MF.(1953)**.Animal microbiology 5th edition.The university of Chicago press. Chicago.
14. **Holt,J.G., Krieg, N.R., Staley, J.T., Williams, S.T.(1994)**.Group 2 aerobic/microaerophilic.Motile, Helical/vibroid gram negative bacteria. In: Bergeys manual of determination bacteriology.PP.42-48 19th edition Williams&Wilkins, USA.
15. **Huang Y, Tian XF, Fan XG, Fu CY& Zhu C.(2009)**. The pathological effect of *Helicobacter pylori* infection on liver tissues in mice. *ClinMicrobiol Infect.*15(9):843-9.
16. **IARC Working group on the evaluation of the carcinogenic risks to humans.(1994)**, *Helicobacter pylori* .Schistosomes, Liver Flukes and *Helicobacter pylori* ;Views and Expert Opinions of an IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Lyon IARC;177-240.
17. **Jamkhande G P. , Gattani G S. , and Farhata. S.(2016)**.*Helicobacter pylori* and cardiovascular complications: a mechanism based review on role of *Helicobacter pylori* in cardiovascular diseases. *Integr Med Res.* 5(4): 244–249.
18. **Jones, K. R., Whitmire, J. M., and Merrell, D. S. (2010)**. A tale of two toxins: *Helicobacter pylori* CagA and VacA modulate host pathways that impact disease. *Front. Microbiol.* 1:115.
19. **Kini D, Agarwal R, SaraswatVa, Naik SR.(2001)**. Role of *Helicobacter pylori* in hyperammonia& subclinical hepaticencephalopathy in cirrhosis of liver.*IndianJ. Gastroenterol.*;20:237-40.
20. **Lee M. , BaekH., Suk ParkJ., KimS., KyungC., BaikSu J., Lee B. K. , Kim J. , AhnC. W. ,Kim K. R. , and Kang S . (2018)**. Current *Helicobacter pylori* infection is significantly associated with subclinical coronary atherosclerosis in healthy subjects: A cross-sectional study. *Plos One*;13(3).
21. **Marshall, B.J., Warren, J.R., Francis, G.J., Langton, S.R., Goodwin, C.S.&Blinow, E.D.(1987)**. Rapid urease test in the management of *campylobacter pyloridis* associated gastritis. *Am.J.Gastroenteriol.*;82:200-10.
22. **Montgomery, E., Martin, D.F. &Peura, D.A.(1988)**. Rapid diagnosis of *Campylobacter pylori* by grams stain .*J .Clin . Pathol.*90:606-9.
23. **Naito Y& Yoshikawa T.(2002)**,Molecular & cellular mechanisms involved in *H. pylori*-induced inflammation & oxidative stress. *Free Radical.Biol.Med.* ;33:323-36).
24. **NIH Consensus conference.(1994)**. *Helicobacter pylori* in peptic ulcer disease .NIH Consensus Development.Panel on *Helicobacter pylori* in peptic ulcer disease.*JAMA*; 272:65-69.
25. **Reyrat, J.M., Pelicic, V., Papini, E., Montecucco, C., Rappuoli, R.&Telford, J.L.(1999)**, Towards deciphering the *Helicobacter pylori*cytotoxin. *Mol. Microbiol.*; 34: 197–204
26. **Sulewska A., Modrzejewski W., Kovalchuk O., Kasacka I., Jackowski R., Hirnle T.(2004)**. Attempts to detect *Helicobacter pylori* in the atherosclerotic plaques. *RoczAkad Med Bialymst.*49:239–241.
27. **Terradot, L., and Waksman, G. (2011)**.Architecture of the *Helicobacter pylori* Cag-type IV secretion system. *FEBS J.* 278, 1213–1222.

28. Vasconez C, Elizalde JI, Liach J, Gines A, dela Rosa C, Fernandez RM, et al.(1999).*H. pylori* ,hyperammonia& subclinicalportal-systemic encephalopathy:effects of eradication.*J.Hepatol.*;30:260-64.
29. Wang, F., Meng, W., Wang, B.&Qiaol, L.(2014). *Helicobacter pylori* induced-gastric inflammation and gastric cancer.*Cancer Lett.* 345(2): 196-202.
30. Wang,X., Sturegrad, E., Rupart, R., Nilsson, H.O., Aleljung, P.A., Garlen, B., Willen, R.&Wadstrom, T.(1997).Infection of BALB/c a mice by spiral and coccoid forms of *H. pylori*.*J.Med. Microbiol.*46:657-63.
31. Wijarnpreecha K, Thongprayoon C, Nissaisorakarn P, Jaruvongvanich V, Nakkala K, Rajapakse R, Cheungpasitporn W.(2017). The association of *Helicobacter pylori* with chronic kidney diseases: a meta-analysis. *Dig Dis Sci.*62(8): 2045-2052.



Figure(1):Kidney in mice, vacular degeneration (star shaped) () with infiltration within glomerulus. () (H&E) (X40).

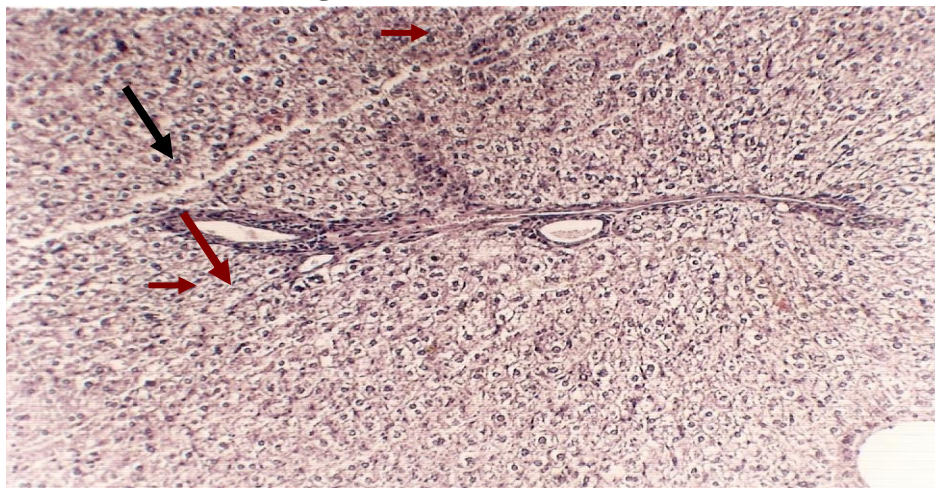


Figure (2):Liver in mice, Vacular degeneration () & perivascular leukocyte cuffing (). (H&E) (X10).

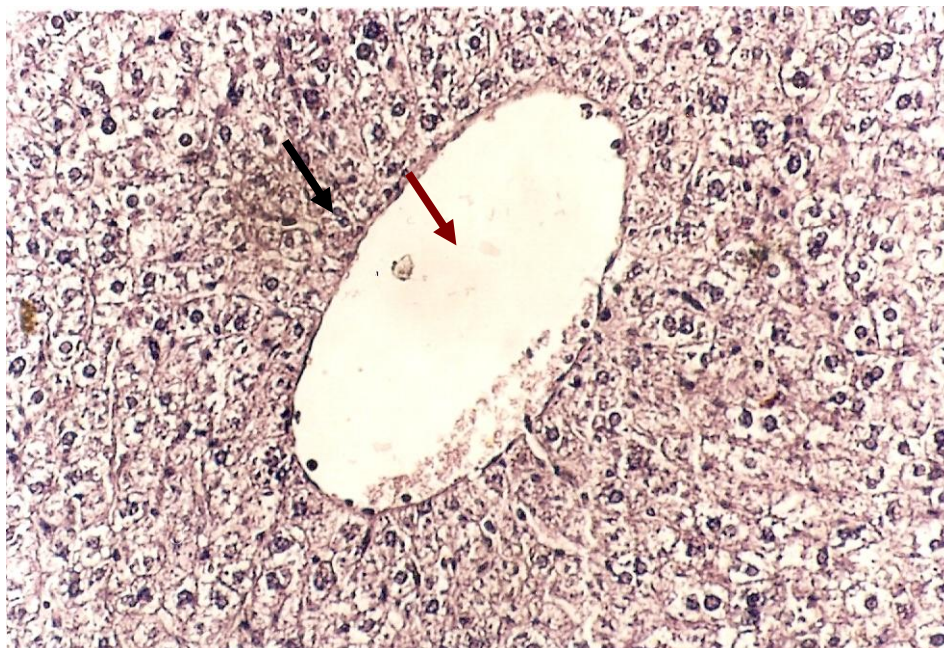
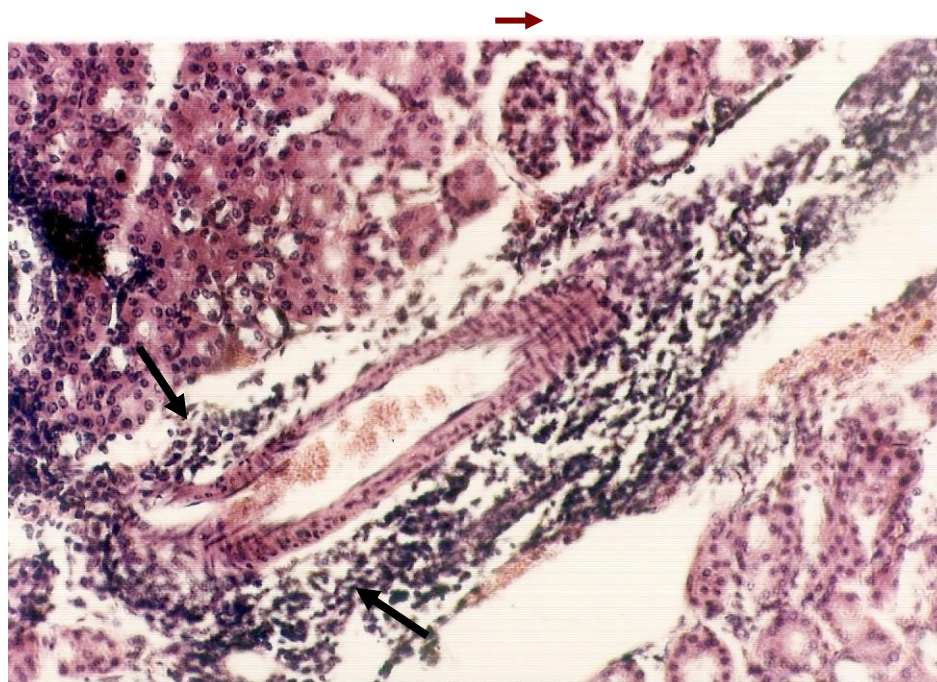


Figure (3): Liver,diltation of the central vein (),enlargement of hepatocytes (vacuolar degeneration)(),depletion of the sinusoids.



Figure(4):perivascular leukocyte cuffing(lymphocytes)() & vacuolar degeneration.(H&E) (X20).

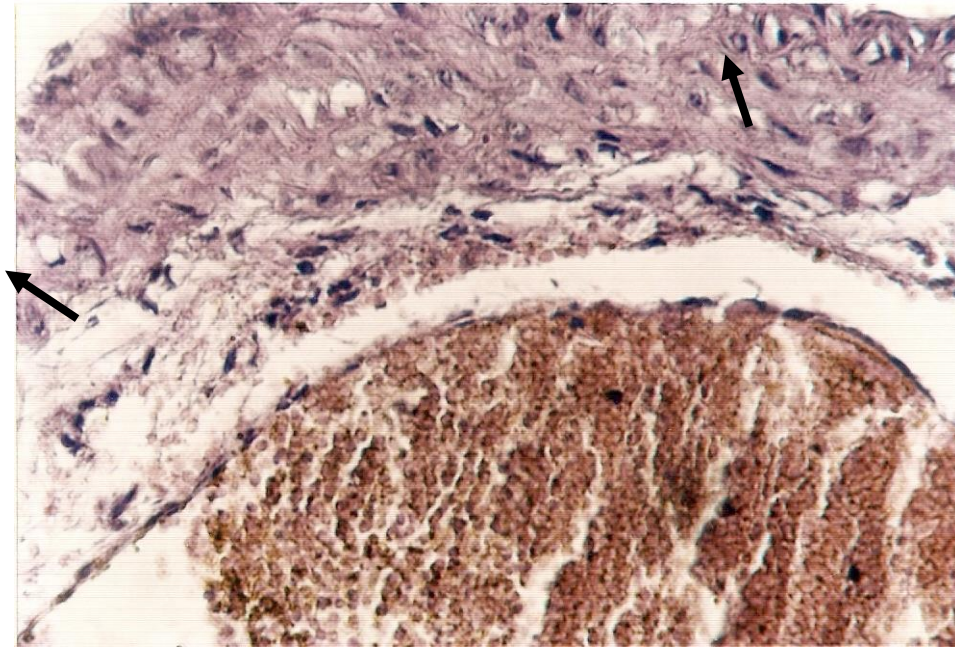


Figure (5): Heart in mice, Show thrombosis, perivascular leukocyte cuffing & edema with vacuolar degeneration() of cardiac muscle fibers.(H&E) (X40).

