

# Biosynthetic Copper Oxide Nanoparticles by Lacto Bacillus and its Anti-Cancer Activity against Human MCF-7 and PC3 Cell Lines

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**Abstract** . Recent development in nanotechnology Metal and metal oxides nanoparticles have a wide range of applications in a variety of fields, research institutes, and industries. Copper oxide nanoparticles are the most common metal oxides. (CuONPs) The good capability of copper-tolerant probiotic of *Lactobacillus* tolerating high concentration of copper<sup>+2</sup> and producing Copper Oxide Nanoparticles, Because of its unique properties and applications, these bacteria have gained more interest as a natural microbial cell nano-factory for a more effective and environmentally friendly method of biosynthesis of these nanoparticles. The morphological and structural properties of CuO NPs were determined by X-ray diffraction, Field Emission Scanning Electron Microscope and Transmission Electron Microscopy (TEM) showed that the synthesized nanoparticles were crystalline, moderately stable, roughly spherical and pure, (CuONPs) biosynthesized by *Lactobacillus plantarum* show activity against human cancerous cell line MCF-7 and PC3 cell lines. MCF-7 cell line showed more activity in less concentration CuONPs on cancer MCF-7 compared to PC3. PC3 was affected in high concentration of 6 CuONPs. The cytotoxic effect of CuO nanoparticles from *Lactobacillus plantarum* on normal cell line (WRL) was small on high concentration CuONPs.

**Keywords:** CuONPs, *Lactobacillus plantarum*, CuONPs Anti-cancer activity against human MCF-7, CuONPs Anti-cancer activity against PC3 cell lines

## 1- Introduction

Nanotechnology is the creation, Materials that are produced, exploited, and synthesized on a scale smaller than 1  $\mu\text{m}$  [1]. The term "nano" comes from a Greek word that means "dwarf" or "extremely thin." Nanoparticles are metal particles with a diameter of 1–100 nm and a variety of shapes such as spherical, triangular, rod, and others (Sau and Rogach (2010)). They are used in biotechnology, nanotechnology, physics, chemistry, and other fields (Rai et al., (2008)). Lactic acid bacteria are Gram-positive, non-spore-forming Rods, Cocci, and Cocco-bacilli that produce lactic acid as one of the main fermentation products by using carbohydrates during fermentation. They are non-respiring but aerotolerant, fastidious, acid resistant, and catalase negative without cytochromes. to the retail [2]. Lactic acid bacteria are one of the most important microbes in the fermentation and processing of food. Lactic acid bacteria are commonly used in the production of fermented foods such as kefir, cheese, butter, yogurt, and beverages, and they help to speed up the fermentation process. During sugar fermentation, LAB produces lactic acid and other organic acids, which lowers the pH of the atmosphere and thus inhibits the growth of unwanted microbial agents. They also help to keep food fresh by developing secondary metabolites such as lactic acid, fatty acid, and bacteriocin, which inhibit the growth of spoilage and pathogenic bacteria [3]. NanoCuO is cheaper than silver, easy to combine with polymers, and relatively stable in terms of chemical and physical properties, according to the limited knowledge available on its potential antimicrobial activity. CuO as well as other highly ionic nanoparticulate metal oxides, which can be prepared with extremely large surface areas and peculiar crystal morphologies, can be especially useful antimicrobial agent [4]. CuO, TiO<sub>2</sub>, Fe<sub>3</sub>O<sub>4</sub>, ZnO, and NiO NPs are transitional metal oxides that have shown to be useful as advanced nano-substances in the fields of energy, biomedicine, and the environment. These NPs' high adsorption potential increases their performance and applications significantly [5]. *Lactobacillus plantarum* is used as an effective reduction and capping agent in the current research, which results in a low-cost, previously unreported, and simple method for the biosynthesis of CuO NPs. *Lactobacillus plantarum* is a gram-positive facultative anaerobic bacteria that is nonpathogenic. It has the largest genome among the lactic acid bacteria. *Lactobacillus plantarum* is

electrokinetically negative CuO NPs are biosynthesised using a potential that absorbs cations easily and acts as a catalyst [6]. In addition to the previously listed applications, CuONPs have shown exclusive anticancer, antimicrobial, and antioxidant efficacy, making them a promising biomedical process [7,8].

## 2-Methodology

**Bacterial Isolate:** *Lactobacillus* ssp was preferred as a biological model for CuO NP synthesis because it is more effective. This isolate was collected from dairy products stored at the University of Babylon's Advanced Microbiology Lab. We cultured the isolate for 24 hours on MRS agar at 38°C, and it was diagnosed using the Vitek2 system

**Copper oxide Nanoparticles' Biosynthesis by *Lactobacillus* ssp:** The *Lactobacillus* ssp pure culture was inoculated in a flask containing MRS broth that had been autoclave sterilized, at 37°C incubated for 24 hours at 100 rpm. After the incubation time, we centrifuged the supernatant at 5000 rpm for 25 minutes. To delay the transformation process, the pH of the supernatant was controlled with 0.4 M NaOH (the pH of the supernatant is acidic 4.7 to be neutral we added the NaOH to reach pH 7 to eliminate the influence of organic acids). 20 g. 0.4 M Cu(NO<sub>3</sub>)<sub>2</sub>·H<sub>2</sub>O dissolved in 1000 ml distilled water, was added to 250 ml supernatant, and then heated for 5-10 minutes in a water bath at 85 °C. The transformation process is indicated by the appearance of a black precipitate at the lowest of flask. The flask was then incubated at 37° C for 12 hours, during which time all of the particles collected at the bottom of the flask. To separate the black precipitate, the produce was centrifuged at 10000 rpm for 20 minutes and washed with D.W. The procedure was then repeated three times to obtain pure products, after which the product was dried at 60 °C in a hot air oven for four hours [9].

**The instruments that used in determining the properties of CuO nanoparticles biosynthetic by *Lactobacillus plantarum*:-**

- X-ray diffraction analysis (XRD).
- Field Emission-Scanning Electron Microscopy (FESEM).
- Transmission Electron Microscopy (TEM)

On Tumor Cell Lines, the Cytotoxic Effect of Copper Oxide Nanoparticles Synthesized from *L. plantarum* on (MCF-7 & PC3) cancer human cell and WRL 68 normal cell line. This method was used to investigate the possible cytotoxicity of various concentrations of CuO NPs nanoparticles synthesized from *L. plantarum*. Maintaining Cell Lines [10].

A- After aspirating the growth medium, the cell sheet was washed with PBS.

B- A two to three mL trypsin/EDTA solution was applied to the cell. The vessel was turned over with gentle rocking to fully cover the monolayer. Before being removed from the vessel, the cells were allowed to incubate for 1 to 2 minutes at 37°C.

C- A fresh complete RPMI medium (15-20 ml) was added to the growth medium, and cells from the wedding surface were pipetted in.

D- Cells were redistributed to the target concentration in culture vessels, flasks, or plates, and incubated for 24 hours at 37°C in a 5% CO<sub>2</sub> incubator. A haemocytometer was used to count the cells and assess their concentration. Applying the formula: Total Cell Count/ml: cell count x dilution factor (sample volume) x 10<sup>4</sup>

### MTT Protocol

Copper Oxide and CuO NPs were tested for cytotoxicity using an MTT prepared to use kit (Intron Biotech):

(MTT solution 1 mL x 10 vials, Solubilization solution 50 mL x 2 bottles) are included in the package.

Tumor cells ( $1 \times 10^4$ – $1 \times 10^6$  cells/ml) were grown in a final volume of 200ml complete culture medium for 24 hours per well in 96 flat well micro-titer plates. After being wrapped in sterilized parafilm, the microplates were gently shaken and set at 37°C with 5% CO<sub>2</sub>. After incubation, the medium was removed and the wells were filled with two-fold serial dilutions (25, 50, 100, 200, and 400 mg/ml) of the desired compound. The controls, as well as triplicates of each concentration, were used (cells treated with serum free medium). For the duration of the exposure time, plates were incubated at 37°C with 5% CO<sub>2</sub> (24 hrs). After exposure, 10 ml of MTT solution was added to each well. Plates were then incubated for another 4 hours at 37°C with 5% CO<sub>2</sub>. The media were carefully extracted, and each well received 100ml of solubilization solution for 5 minutes. At a wavelength of 575 nm, an ELISA reader was used to calculate the absorbance. The optical density data was statistically analyzed in order to determine the compound concentration needed. Each cell line's viability has been reduced by 50%. [31,32] through following equation:

$$Y = D + \frac{A - D}{1 + 10^{(x - \log C)B}}$$

The data were processed and analyzed using one way ANOVA in the social science statistical program (SPSS 19), and the results were expressed as (Mean  $\pm$  S.D) [33,34]. The log (Inhibitor) versus response curve is plotted in Graphpad Prism 6 using data analysis of MTT assays carried out in g/ml and log values of g/ml. For the most important IC<sub>50</sub> values, the best values were chosen.

### 3-Results and Discussion

**Copper Oxide Nanoparticles biosynthesis by *Lactobacillus* spp.** The biosynthesis of nanoparticles from *Lactobacillus* spp. The reduction of CuO into CuO NPs during exposure to bacterial extract, accompanied by a shift in color from brown to black over the course of 24 hours, has been confirmed by observing a change in solution color during the synthesis of CuO NPs. The black color is deposited at the bottom of the flask combining *Lactobacillus* spp. (*L. plantarium*, *L. casei*, *L. acidophilus* and *L. bulgarius*) with C (uNO<sub>2</sub> 33)H<sub>2</sub>O. The biosynthesis of CuO NPs by all these strains shows a change in color during the reaction, from light blue to green to dark brown to black at the end [11]. The strains show a change in color from brown to black and precipitate black color shows the ability of all these strains to biosynthesize CuO NPs and completed CuO NPs is characterized by their optical and structural properties. Different ratios of CuO NPs were biosynthesized, but *L. plantarium* was the most active species, according to the findings. The biosynthesis of CuO NPs in *L. plantarium* is agreed with the results described on [12,13]. *Lactobacillus* spp. has a major role in the CuO NPs generation. It's possible that the negative electrokinetic ability of bacteria, which readily absorbs cations and causes nanoparticle synthesis. *Lactobacillus* can also expand even when oxygen is present, allowing for greater metabolic growth potential. Furthermore, the presence of glucose in the MRS media used to synthesize CuO NPs tends to reduce oxidation-reduction potential. All of these factors, including energy-producing glucose, work together to negotiate CuO NPs synthesis in the presence of *Lactobacillus* (which controls the value of rH<sub>2</sub>). medium pH ionic status, and full oxidation reduction potential (rH<sub>2</sub>) partially mediated through the use of sodium hydroxide in the final stage of CuO NPs biosynthesis by *Lactobacillus plantarium* drying these nanoparticles in the oven [14].

#### Morphological & Structural properties of CuO NPs biosynthesized by *Lactobacillus plantarium*:

X-Ray diffraction analysis (hexagonal phase) shows synthesized nanoparticles were crystalline and pure in nature. The diffraction lines of spherical CuO NPs respectively, figure (1). The Scherrer equation was used to calculate the average particle size of CuO NPs [15]. using the full width at half maximum FWHM  $\{16, \} D = 0.9\lambda / \Delta(2\theta) \cdot \cos(\theta) \dots$ . Thus D is the size of the crystallite,  $\lambda$  is the X-ray source's wavelength, and  $\cos 16 = 0.968583$ . The FWHM is  $\beta$ , and the angle of diffraction is  $\theta$ . The average size of copper oxide nanoparticles has been determined using the formula to be the maximum diameter measured. For particles CuO NPs biosynthesized by *L. plantarium* is 18.84 nm, CuO NPs biosynthesized by *L. acidophilus* 17.4 nm and CuO NPs biosynthesized by *L. casei* is 17 nm. Line extension of peaks is a representation that the synthesized materials fall within the nanometer range (figure 1). *L. bulgarius* do not biosynthesize CuO NPs without peak. The obtained nanoparticles

(NPs) have a too small synthesis, which may be due to the biological synthesis method used to prepare them. demonstrating the biosynthesis of copper oxide nanoparticles on pure copper oxide planes The average crystallite size of biosynthesized copper oxide nanoparticles was calculated using the Scherer formula [16].

The *Lactobacillus plantarum* strain was used to make moderately stable CuO NPs. In the nanoparticles, the impact of response time is important. The shape, average size, and particle sizedistribution of copper nanoparticles were determined using the field-emission scanning electron microscopy characteristics of CuO. [20,21]. Some nanopolymers have been confirmed to be damaged by SEM. As a result, the Nanoparticlesneedto be ablewithstand the effects of vacuum pressure formorphological research with efficiency SEM.Microscopyimage of the CuONanoparticles isAmbiguousshows( ni figure.)3

The descriptionsin the number of functional groups involved in biosynthesis of CuO Nanoparticles are accessed using FTIR spectra. CuO NPs from *L. plantarium*, which were collected had prominent peaks in their FTIRspectral data3365 cm refer to Aliphatic primary amine ,1622.16 cm, Alkenyl c=c stretch , 1570 cm Aromatic ring , 1404.22 ,carbonyl compound , 1082 cm CN Stretch and 613 cm Alphatic iodo c-1stretch show fig3[17,18,19 ]

### X-Transmission Electron Microscopy (TEM)

The TEM images CuO NPs nanoparticles shown in Figure (4). If the doping amount of metals increases, the particle size decreases. The size and crystallinity of the synthesized CuO NPs were described using Transmission Electron Microscopy, Dynamic Light Scattering was used to determine the zeta potential of the biosynthesized CuONPs (DLS). DLS was used to determine the average size of the nanoparticles, which was found to be between 20 and 30 nanometers. The average size of biosynthesized CuO NPs was discovered to be nm [22] .The best among other electron microscopy techniques for determining the morphological identities of CuONPs and other metal nanoparticles is transmission electron microscopy (TEM) [23]. In the TEM technique, a beam of energetic electrons is transmitted through an ultra-thin sample and interfaces, resulting in an image. [24].HRTEM (high resolution transmission electron microscopy) is a type of TEM that has a higher resolution and can image crystallographic structures at the nuclear scale. [25].

**elbaT(1): X-Ray diffraction analysis biosynthesis CuO NPs from *Lactobacillus* spp. (*L.plantarum* ,*L.casi* ,*L.acidophilus* and,*L.bulgarius*)**

Sample	Pos. 2θ(deg.)	FWHM	I/I <sub>0</sub>	No. of peak	Average Crystal size (nm)
<b>CuO NPs from</b>	<b>44.0682</b>	<b>0.48880</b>	<b>79</b>	<b>32</b>	
<i>L. plantarium</i>	<b>64.4091</b>	<b>0.48040</b>	<b>99</b>	<b>45</b>	<b>18.84±1.28A</b>
	<b>77.5108</b>	<b>0.56940</b>	<b>100</b>	<b>48</b>	
<b>CuO NPs from</b>	<b>44.0625</b>	<b>0.50540</b>	<b>85</b>	<b>36</b>	
<i>L.acidophilus</i>	<b>64.4106</b>	<b>0.48650</b>	<b>84</b>	<b>47</b>	<b>17.4±0.96B</b>
	<b>77.5353</b>	<b>0.56570</b>	<b>100</b>	<b>51</b>	
<b>CuO NPs from</b>	<b>44.0610</b>	<b>0.53180</b>	<b>74</b>	<b>32</b>	
<i>L.casi</i>	<b>64.4173</b>	<b>0.51050</b>	<b>88</b>	<b>47</b>	<b>17±1.02B</b>
	<b>77.4960</b>	<b>0.53830</b>	<b>100</b>	<b>55</b>	<b>Average Crystal size (nm)</b>
<b>LSD<sub>0.05</sub></b>					<b>1.08</b>

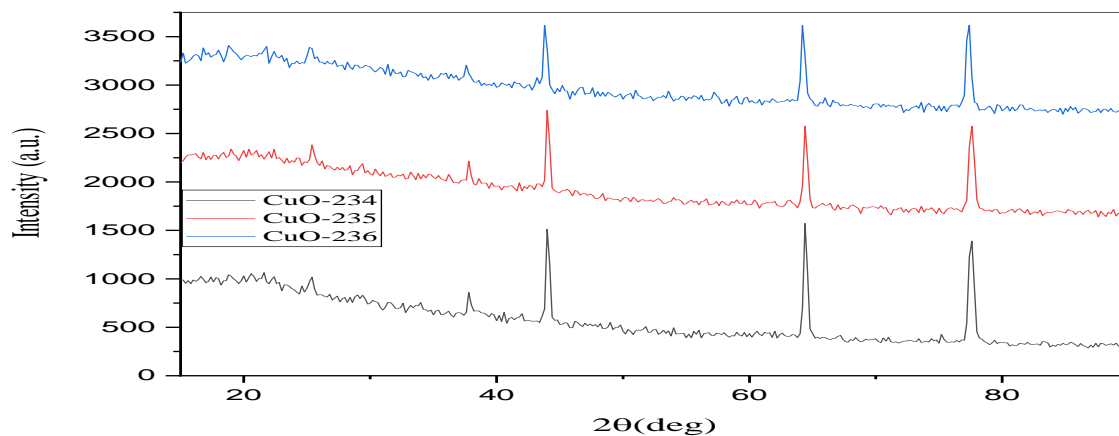


Figure1. XRD analysis for a. CuO NPs biosynthesized by *L. plantarium*. b. CuO NPs biosynthesized by *L. acidophilus*. c. CuO NPs biosynthesized by *L. casi*

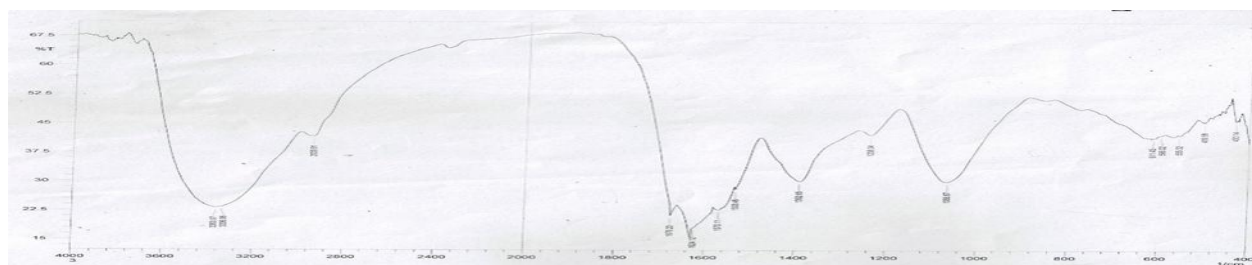


Figure (3) FTIR results biosynthesized CuO NPs From *L. plantarium*.

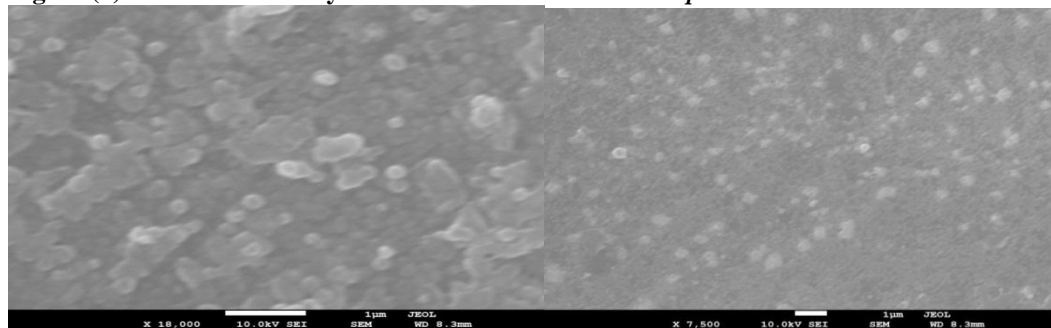


Figure2. Field Emission Scanning Electron CuO NPs biosynthesized by *L. plantarium*

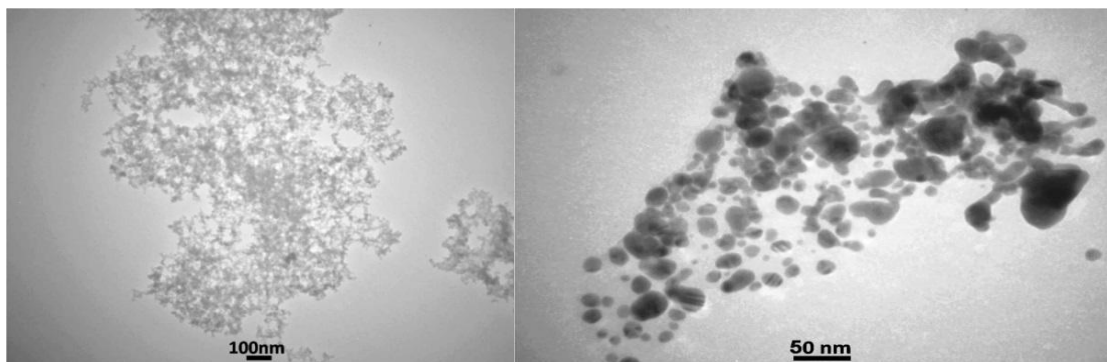


Figure4. TEM micrographs of CuO NPs bio-synthesized by *L. plantarium*

### Anticancer activity of CuO NPs by using MTT analyze

CuO NP was tested for cytotoxicity using the 3-(dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) stain on two cell lines: MCF-7 breast cancer cell line and WRL-68 human hepatic cell line. Using different concentrations of CuO NP on tumor cell lines, this assay was used to determine cell viability and inhibition rate. MTT colorimetric assays were used to assess cell viability at each time point. CuO NPs synthesis by *L.plantarium* at various concentrations (25, 50, 100, 200, and 400)g/ml resulted in a dose-dependent reduction in cell viability on MCF-7 and WRL-68 cells, as shown in table (2) and Fig (5). LSD<sub>0.05</sub> (Fig. 6). The data was analyzed in g/ml, and the log values of g/ml were plotted in Graphpad Prism 5 using the log (Inhibitor) versus response curve. For the most important IC<sub>50</sub> values, the best values were chosen. MTT colorimetric assays were used to assess cell viability at each time point. On a regular cell line, *graveolens* (WRL) The results showed that incubating WRL cells with CuO NPs and a control for 24 hours at different concentrations ranging from 50 to 400g/ml showed low reduction in cell viability as compared to cancer cells, with a dose-dependent pattern in which cell viability decreases as CuO NP concentration rises. [26] Extract CuO NPs at a concentration of 400g/ml had the lowest WRL cell viability (66.86 percent). The concentration 400 g/ml reduced MCF-7 cell viability by 34.07 percent on average, while WRL-68 cell viability was 66.86 percent, and MCF-7 cell viability was (82.45, 63.58, 54.48, and 36.06 percent) respectively. and the viability of the same CuO NPs on WRL-68 are (94.25, 96.18, 87.11 and 72.92%), respectively. The IC<sub>50</sub> for WRL-68 are (153.0), while IC<sub>50</sub> of MCF7 cell lines was 63.64 µg/ml. Table(2): CuO NP synthesis from *L.plantarium* on control human liver cell line (WRL-68) and (MCF-7) cell line toxicity increases with increasing CuO concentration, reaching a limit at 50 g/mL. (85.9%). CuONPs biosynthesized from black bean extract were tested for anticancer activity using the sulforhodamine B analyze, which showed when incubated with CuONPs, there are a few variations in mitochondrial structure. CuONPs also stopped cervical carcinoma cells from growing [27]. According to a study, CuONPs derived from *Ficus religiosa* inhibit the growth of A549 adenocarcinoma cells basal epithelial cells. [28] Based on the mortality data obtained from this study, we were able to predict their capacity as cytotoxic agents. Antitumor activity has been demonstrated for flavonoid compounds, especially quercetin and genistein; these compounds were toxic to cancer cells but had no effect on normal cells [29].

**Table(2): Represents the activities CuO NP synthesis from *L.plantarium* . on control human liver cell line (WRL-68) and breast cancer cell line (MCF-7)**

CuO NPs Conc. µg/ml	Viability % of MFC-7 mean± SD	IC <sub>50</sub> of MFC-7 µg/ml	Viability % of WRL-68 mean± SD	IC <sub>50</sub> of WRL-68 µg/ml
25	82.45±2.53A	63.64	94.25±1.36A	153
50	63.58±3.22B		96.18±1.25B	
100	54.48±3.83C		87.11±3.65C	
200	36.06±4.41D		72.92±2.20D	
400	34.07±2.49D		66.86±3.35E	
LSD <sub>0.05</sub>	2.78		1.65	

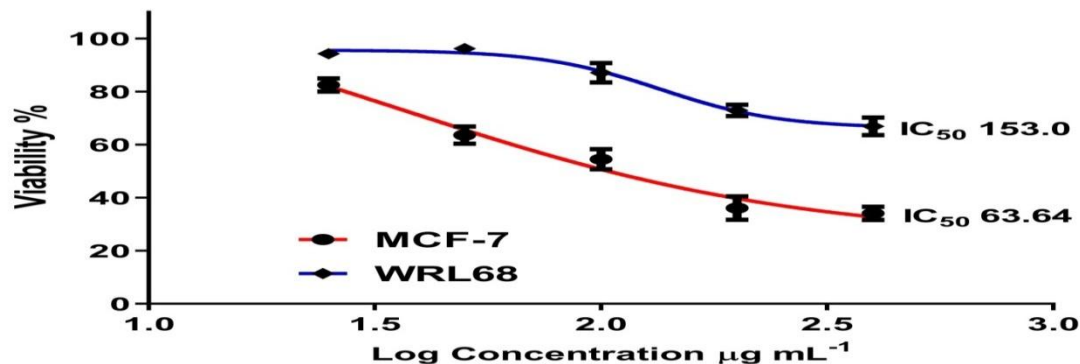


Figure (5): displays the IC<sub>50</sub> of CuO NP synthesis from activities on control WRL-68 and breast cancer cell line MCF-7.

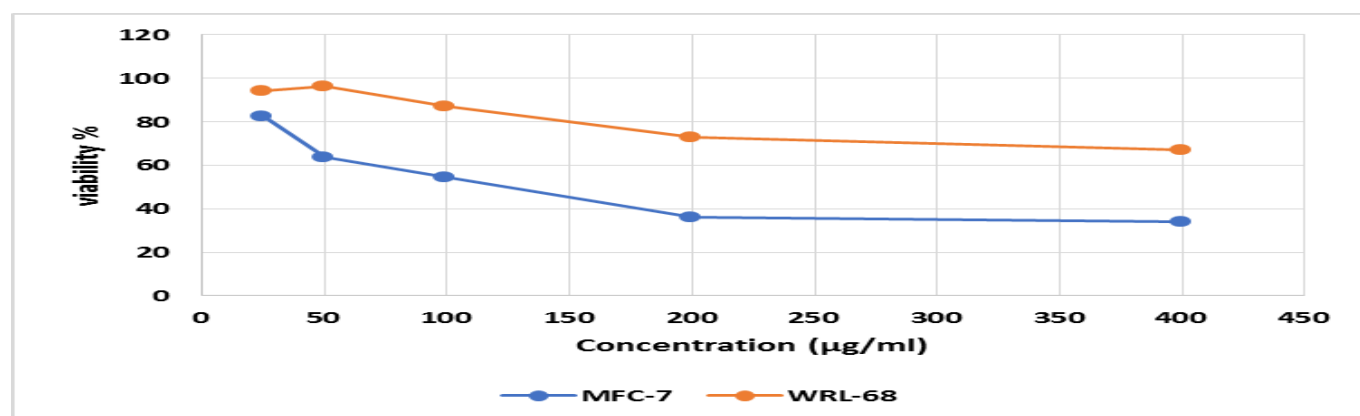
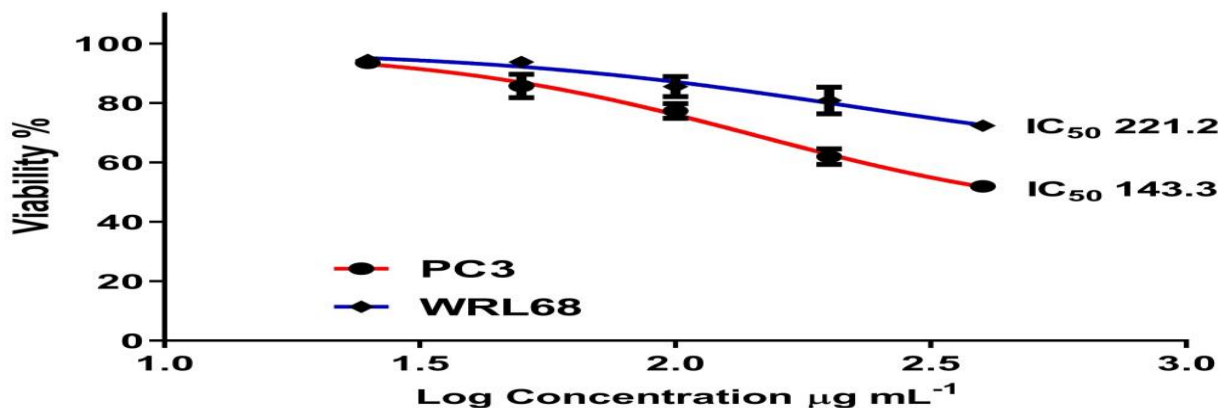


Figure (6) viability cell percentage of different concentrations of CuO NPs on MFC-7 and WRL-68 cell line.

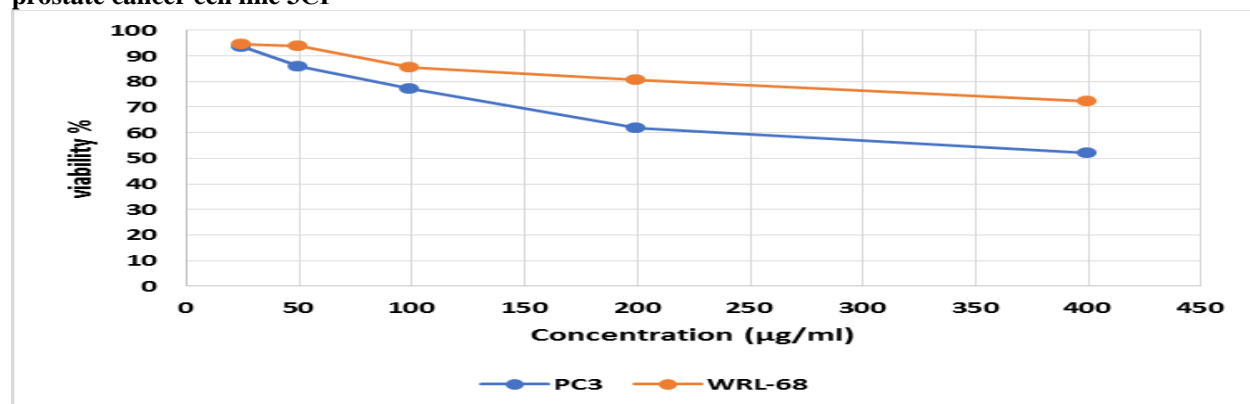
Cuo NP was tested for cytotoxicity using the 3-(dimethylthiazol-2-yl)-2,5- diphenyl tetrazolium bromide (MTT) stain on two categories of cell lines: prostate cancer cell line (PC3) & human hepatic cell line WRL-68. By applying different concentrations of Cuo NPs to tumor cell lines, this assay was used to determine cell viability and reserve rate. Cuo NP synthesis by *L.plantarum* used in different concentration (25, 50, 100, 200 and 400) µg/ml. In a dose-dependent manner, induced a decrease in cell viability on PC3 and WRL-68 to showed the viability and cytotoxicity of Cuo NPs as in table (3) and Fig.(7) Fig (8) LSD<sub>0.05</sub>. The results showed that incubating WRL cells with CuO NPs and a control at different concentrations ranging from 50 to 400 µg/ml for 24 hours resulted in a low reduction in cell viability as compared to cancer cells in a dose-dependent fashion, with cell viability decreasing as the concentration of CuO NPs increased. Extract CuO NPs at a concentration of 400 µg/ml had the lowest WRL cell viability (66.86 percent). The viability of the PC3 cell line is 51.94 percent, while the viability of the WRL-68 cell line is 72.34 percent, and the viability of the other concentrations is unknown. (25, 50, 100 and 200) µg/ml on MCF-7 cell line is (93.52, 85.69, 77.28 and 61.92 %) respectively, and the viability of the same CuO NPs on WRL-68 are (94.44, 93.79, 85.49 and 80.80 %), respectively. The IC<sub>50</sub> for WRL-68 are (221.2), while IC<sub>50</sub> of PC3 cell lines was 143.3 µg/ml, The antiproliferative profile was improved by increasing the CuNPs concentration [30]. The antitumor activity of Cu NPs was studied in the A549 cell line, and it was found to have excellent antiproliferative activity with an IC<sub>50</sub> of 120 µg/mL CuNPs concentration. Biological and nonbiologically synthesized NPs IC<sub>50</sub> values for cervical HeLa human cancer cells were 78.9, 66.4, 71.8, and 85.5 µg/mL, respectively, and 88.6, 82.9, 77.5, and 91.7 µg/mL for PC3 human prostate cancer cells were 88.6, 82.9, 77.5, and 91.7 µg/mL, respectively. were found for nonbiologically Nanoparticles of Ag, Au, Co, and Cu were synthesized [29].

**Table(3):** Represents the activities CuO NPssynthesis from *L.plantarum* . on control human liver cell line (WRL-68) and prostate cancer cell line (PC3).

CuO NPs Conc. $\mu\text{g/m}$	Viability % of PC3 mean $\pm$ SD	IC <sub>50</sub> of PC3 $\mu\text{g/ml}$	Viability % of WRL-68 mean $\pm$ SD	IC <sub>50</sub> of WRL-68 $\mu\text{g/ml}$
25	93.52 $\pm$ 1.1A	221.2	94.44 $\pm$ 0.31A	143.3
50	85.69 $\pm$ 3.94B		93.79 $\pm$ 0.93A	
100	77.28 $\pm$ 2.49C		85.49 $\pm$ 3.37B	
200	61.92 $\pm$ 2.61D		80.8 $\pm$ 4.47C	
400	51.94 $\pm$ 0.85E		72.34 $\pm$ 0.69D	
LSD <sub>0.05</sub>	2.78		1.65	



**Figure (7):** displays the IC<sub>50</sub> of CuO NPssynthesis from activities on control WRL-68 and prostate cancer cell line 3CP



**Figure (8)** viability cell percentage of different concentrations of CuO NPs on PC3 and WRL-68 cell line.

#### 4-Conclusion

The method used in the biological and catalytic applications of *Lactobacillus* spp in the synthesis of CuONPs, which provides a great opportunity to medicinal institutes biological activity and mode of synthesis from *Lactobacillus* spp, which provides medicinal institutes with a great opportunity. Our actual situation research into the biosynthesis of CuO NPs by *Lactobacillus Plantarum* and the use of CuO NPs as anticancer has yielded excellent results with few complications and a low cost. It is well highlight the role of recent characterization techniques in evaluating the identities of CuONPs. CuO-NPs have a greater effect on MCF7 cell lines than PC3 cell lines, and CuONPs have more activity on MCF7 cancer cell lines. On high concentration CuONPs, the cytotoxic effect of CuO nanoparticles from *Lactobacillus planetarium* on a normal cell line (WRL) was minimal. The synthesis and toxicity of the compound are

well explained. More research should be done to improve CuONPs' biological applications are being explored by focusing on ways to reduce CuO-NPs' toxicity while maintaining and improving their biological quality.

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