# Microbiological Assssment in Plaque Samples of Patients with Oral Cancer with or without Smoking

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**Aim:** To isolate micro-organisms from the GCF, Serum and saliva in patients with oral malignancy with or without Chronic period on titis. To detect virus from the serum, saliva and GCF in patients with Chronic period ontit is with and without smoking. To evaluate ROM levels from the GCF, saliva and Serum with oral malignancy with and without smoking and Chronic periodontitis.

### Objectives

- To Compare he relation between Chronic Periodontitis with Oral carcinoma
- To compare the relation between Chronic periodontitis without Oral carcinoma
- To compare the relation between Chrionic periodontitis with Smoking
- To compare the relation between Chronic Periodontitis without smoking
- To correlate the relation between Oral carcinoma with smoking
- To correlate the relation between Oral carcinoma without smoking

### **Materials and Methods**

25 patients had been selected for each group and samples were obtained respectively. A single dentist was assigned to collect samples to maintain standards. Patients were in the aged between 30 to 55 years and were obtained from the Department of Periodontics, Thai Moogambigai Dental College, Chennai, TamilNadu, India and Oral cancer patients were taken from MM Hospital Namakkal, and Cancer Institute, Chennai, TATA Memorial Hospital, Mumbai.

### INTRODUCTION

Periodontal diseases are a group of conditions affecting the supporting structures for the dentition. Chronic periodontitis is the result of are sponse of the host to bacterial aggregations on the tooth surfaces. The outcome of this is an irreversible destruction of the connectivet issue attachment, which results in periodontal pocket formation and eventual loss of alveolar bone. Whilegingivitis is known to be a very prevalent condition among children and adolescents, periodontitis is much less common in this group. The occurrence of severe periodontit is in young adults may have a devastating effect on their dentition and in some cases treatment of these forms of periodontal disease can be unsuccessful. Diagnosis of periodontitis and the identification of affected individuals can sometimes be difficult because there may be no selfreported symptoms. Destructive periodontitis has been described as a consequence of the interaction of genetic, environmental, microbial and host factors Loeetal 1994. Hujoel suggests that a hidden periodontitis epidemic related to smoking patterns occurred during the 20<sup>th</sup> Century and that socio demographic shifts in smoking habits will alter the periodontal needs for the future. The precise mechanism where by cigarette smoking exerts an effect on the development of periodontal destruction is unknown. A reduction in clinical signs of gingivitis hasbeen reported in smokers and this effect has been shown to be independent of plaque levels. There have been other reports of less bleeding in smokers with periodontitis, suggesting that nicotine could mediateitsvaso active effects on alocal basis Graneretal, Lindenand Mullally found that young smokers had infact more gingival bleeding than non-smoking regular attenders. The explanation for this finding seemed to be related to the high levels of calculus and plaque reported in this group of young adults. Other studies of older population groups have found little difference in plaque accumulation between smokers and non-smokers.

Nicotine metabolites can concentrate in the periodontium and their effects include the promotion of vasoconstriction, and the impairment of the functional activity of polymorphs and macrophages. The numbers of neutrophils in peripheral blood are also increased by tobacco use and their migration through capillary walls is impaired due also to paralysis of the cell membrane. Cigarette smoking has been demonstrated to activate the release of elastase, which has the capacity to cause tissue damage. The effect of this is particularly well displayed in lung diseases such as emphysema and in animal studies with elastase-deficient models, which do not develop emphysema when exposed to cigarette smoke. In addition there is an increased production of oxygen species, which c tissue levels of alpha- 1 protease inhibitors so enabling elastase and other enzymes with potential for damaging tissue to remain unchecked at active sites Moeker gingival crevicular fluid levels of functional elastase and that complexed with the inhibitor have been demonstrated to be lower in smokers than nonsmoking controls

Much evidence on the effect of cigarette smoking on neutrophil activity suggests that these cells may accumulate at the site of inflamed periodontal tissues. As they may fail to migrate through the gingival crevice they can release their enzymes into the surrounding connective tissue therefore contributing directly to tissue destruction. The role of cytokines has also been extensively examined in the pathogenesis of periodontitis. High levels of prostaglandin influence of Tobacco Smoking on the onset of Periodontitis in young persons PGE2 have been associated with aggressive or early onset periodontitis and these elevated concentrations have been related to an increased responsiveness of circulating monocytes to bacterial challenge and their effect on apoptosis of monocytes by nicotine exposure but these investigators did report that when stimulated by LPS monocytes related to limiting the spread of infections and its inhibition by nicotine is more evidence of the complexity of effect of tobacco on this part of the host defence mechanism. Our study aimed to detect micro organisms and evaluate the ROM levels in GCF, saliva and serum of oral malignancy patients with periodontal diseases.

### **Study Population**

The study population consisted of 120 subjects belonging to both sexes and aged between 30-55 years. Chronic Periodontitis subjects were taken from the outpatient clinic of Department of Periodontics, Thai Moogambigai Dental College, Chennai, Tamil Nadu, India. Oral cancer patients were taken from MM Hospital Namakkal, and Cancer Institute, Chennai, TATA Memorial Hospital, Mumbai.

Total of 25 patients had been selected for each group and samples were obtained respectively. Each patients was examined by 1 standard dentist and then assighned to the respective group inorder to maintain levels of standardisation.

### **Inclusion Criteria**

Presence of inflammatory changes in the periodontal tissues

Gingival index  $\geq 1$ 

Probing depth and Clinical attachment loss  $\geq$ 4mm

Radiographic evidence of bone loss

Current smoker (packets >10)

Carcinoma present for more than 8 months

Patients with a history, signs or symptoms of Aggressive Periodontitis, History of periodontal treatment received in the past six months, Under antibiotics and corticosteroidsGross oral pathological findings or history of systemic disease were excluded from the study

#### Sampling

- The site with greatest probing depth was selected for GCF collection. After drying the area with a blast of air, supra-gingival plaque was removed without touching the marginal gingiva and the GCF was collected. A standardized volume of 1µl was collected from each site with an extra-crevicular approach, using volumetric capillary pipettes that were caliberated from 1-5 µl. The collected GCF was transferred immediately to ependorfftubes and stored at -70 c until the time of assay
- Whole saliva (2 mL) was collected in disposable, sterile, clean tubes and centrifuged immediately to remove cell debris
- Venous blood from an anticubital vein was collected in plain tubes without additiveCentrifuged at 3,500rpm for 5 minutes to separate serum

Real-Time PCR, also known as quantitative polymerase chain reaction (qPCR), is a laboratory technique of molecular biology based on the polymerase chain reaction (PCR).PCR was used to determine the prevalence of periopathic bacteria in subgingival plaque samples of individuals with chronic periodontitis and oral cancer

Comparison of Saliva, Serum, GCF\_ROM values among all four groups using oneway ANOVA and Posthocanalysis. Comparison of mean number PI,PG,TF,SM & HP organisms in all four groups using oneway ANOVA and posthoc Analysis. Mean difference among all the variables among gender using Independent T test.

ГS										
					95% Confidenc Me	te Interval for en				
	N	Mean	Deviation	P value	Lower Bound	Upper Bound	Minimum	Maximum		
CP+CA	15	1,8000	0.67612		1.4256	2 1744	1.00	Э		
CP-CA	15	2.0667	0.59362		1.7379	2 3954	1.00	3		
CP+SMOKIN G	15	2.6000	0.50709	0.001	2 3192	2 8808	2.00	з		
CP- SMOKING	15	2.5333	0.51640		2.2474	2,8193	2.00	3		
Total	60	2,2500	0,65419		2.0810	2.4190	1.00	3		

#### RESULTS

Mean Gingiva Index among all four groups was statistically significant

	N		Mean Probing Depth	Std. Deviatio	on va	P value		95% Confidence Interval for Mean		Minim	um l	Maxiı	num
			Depui					ver ind	Upper Bound				
CP-	+CA	15	6.6000	1.0556	0		6.0	154	7.1846	5.00	)	8.0	00
CP	-CA	15	6.5333	1.1254	6		5.91	101	7.1566	5.00	)	9.0	00
CP+S <sub>1</sub>	moking	15	7.3333	0.8997	4 0.0	001	6.83	351	7.8316	6.00	)	9.0	00
CP-Sr	noking	15	7.9333	0.9611	5		7.40	011	8.4656	6.00	)	9.0	00
Tota1		60	7.1000	1.1453	7		6.80	041	7.3959	5.00	)	9.0	00
			Mean Clinical			957	6 Cont fo	fidenc r Mez	e Interval m				
Gr	oups		Attachment	Std.		Lo	wer						
		Ν	loss	Deviation	P value	Bo	und	Upp	er Bound	Minimum	Maxi	mum	
a	P+CA	15	5.6000	1.05560		5	5.0154		6.1846	4.00		7.00	
a	P-CA	15	5.5333	1.12546	5	4	.9101		6.1566	4.00		8.00	
a	P+SMOKING	15	6.3333	.89974	0.00	1 5	.8351		6.8316	5.00		8.00	
a	P-SMOKING	15	6.8667	.91548		6	5.3597		7.3736	5.00		8.00	
Te	otal	60	6.0833	1.12433		5	5.7929		6.3738	4.00		8.00	

Mean probing depth & CAL among all four groups was statistically significant, themean values were higher in CP than in CA

	N	Maan	Std.	Derle	95% Co Interval	nfidence for Mean			
		меан	Deviation <sup>r</sup> value		Lower Bound	Upper Bound		Maximum	
CP+CA	15	289.73	21.07967		278.0598	301.4069	268.00	356.00	
CP-CA	15	289.47	13.94820		281.7424	297.1909	267.00	312.00	
CP+Smoking	15	342.13	56.94216	<0.001	310.5998	373.6669	259.00	468.00	
CP-Smoking	15	348.73	67.29197		311.4683	385.9984	271.00	492.00	
Total	60	317.52	52.85429		303.8630	331.1704	259.00	492.00	

GCF\_ROM values among all four groups was statistically significant. The mean values was slightly higher in smokers than in and was similar in cancer and CP

	(D	Mean Difference	e1	95% Confidence Interval			
(1) groups	(J) groups	(I-J)	sig.	Lower Bound	Upper Bound		
CP+CA	CP-CA	1.06667	1.000	-79.8936	82.0269		
	CP+Smoking	-121.06667*	0.001	-202.0269	-40.1064		
	CP-Smoking	-116.13333*	0.002	-197.0936	-35.1731		
CP-CA	CP+CA	-1.06667	1.000	-82.0269	79.8936		
	CP+Smoking	-122.13333*	0.001	-203.0936	-41.1731		
	CP-Smoking	-117.20000"	0.002	-198.1603	-36.2397		
CP+Smoki ng	CP+CA	121.06667*	0.001	40.1064	202.0269		
	CP-CA	122.13333*	0.001	41.1731	203.0936		
	CP-Smoking	4.93333	0.998	-76.0269	85.8936		
CP- Smoking	CP+CA	116.13333"	0.002	35.1731	197.0936		
	CP-CA	117.20000*	0.002	36.2397	198.1603		
	CP+Smoking	-4.93333	0.998	-85.8936	76.0269		

95% Confidence Std. P Interval for Mean Groups Ν Mean Minimum Maximum Deviation value Upper Lower Bound Bound CP+CA 15 3.5333 1.55226 2.6737 4.3929 1.00 6.00 4.5827 0.489 6.00 CP-CA 15 3.7333 1.53375 2.8840 1.00 1.57963 CP+SMOKING 15 4.0667 3.1919 4.9414 2.00 7.00 CP-SMOKING 15 4.3333 1.39728 3.5595 5.1071 2.00 6.00 3.9167 1.51032 3.5265 4.3068 1.00 7.00 Total 60

Serum ROM levels was statiscally significant in smokers with periodontitis and oral cancer On comparison of mean salivary ROM levels were significant I all 4 groups but was greater in smokers than non smokers

When PI,PG,TF,SM&HP levels were compared among all 4 groups, SM and HP were statistically insignificant

### DISCUSSION

Periodontitis history is associated with poorly differentiated tumours in the oral cavity. These results support additional, confirmatory basic science, and prospective clinical studies. Although both are called periodontal disease, gingivitis and periodontitis are distinct diseases. Gingivitis is a nondestructive reversible inflammation of the gums strongly associated with poor oral hygiene. On the other hand, only a small subset of the population with poor oral hygiene develops destructive periodontitis, leading to epithelial migration and bone loss. Factors that initiate periodontitis are poorly understood. Smoking reduces gingivitis but it is a strong risk factor for periodontitis. Gingivitis is mostly associated with Gram-positive facultative bacteria, whereas periodontitis with Gram-negative anaerobic. Accumulating evidence supports a role of viruses in the initiation and progression of periodontitis. Recent study also suggests a synergy between chronic periodontitis and oral HPV infection in base of tongue cancers. A link between poor oral hygiene has been suggested. It is possible that, subjects with poor oral hygiene, those who develop periodontitis are at higher risk for developing cancer. Prospective clinical studies including both periodontitis patients and those with gingivitis without a periodontitis history will allow testing this hypothesis. Alveolar bone loss associated with periodontal inflammation is a slow chronic process that is usually irreversible. The rate of bone loss ranges between 0.04 and 0.28 mm annually. In recent studies, panoramic radiographs were taken at admission before the initial cancer diagnosis. It is not likely that cancer preceded periodontitis. The biological mechanism of the association between chronic infection/inflammation and cancer has been described extensively. However, well-designed longitudinal studies are required to prove.

Over a number of years, epidemiological studies established several well-defined risk factors for cancer such as tobacco, diet, age, hereditary. H. pylori became the first bacterial species to be officially recognized by the World Health Organization as a definite cause of cancer in humans. Since then, there has been a growing body of evidence supporting an association between specific microorganisms, including those in the oral cavity, and various types of cancers. In this study herpes simplex viruses were significantly increased in patients with oral cancer and chronic periodontitis. Subsequently, other community constituents, such as Fusobacteriumnucleatum, can become opportunistically pathogenic, and the combined effect of a dysbiotic microbial community along with a dysregulated immune response ultimately causes periodontal disease

Although smoking is a well-recognized risk factor for periodontal attachment loss, smokers often exhibit less gingival bleeding than would be predicted.

Previous studies have shown that the clinical signs of inflammation are less pronounced in smokers when compared with nonsmokers. These observations may be due to alterations in the inflammatory response in smokers, or due to alterations in the vascular response of the gingival tissues. Although no significant differences in the vascular density of healthy gingiva have been observed between smokers and nonsmokers,

the response of the microcirculation to plaque accumulation appears to be altered in smokers when compared with nonsmokers. With developing inflammation, increases in gingival crevicular fluid flow,' bleeding on probing,' and gingival blood vessels' were less in smokers when compared with nonsmokers. In addition, the oxygen concentration in healthy gingival tissues appears to be less in smokers than nonsmokers, although this condition is reversed in the presence of moderate inflammation. Subgingival temperatures are lower in smokers than nonsmokers, and recovery from the vasoconstriction caused by local anaesthetic administration takes longer in smokers

The involvement of ROS and the antioxidant defense mechanisms in human saliva has been demonstrated in various processes of the oral cavity: healing periodontal disease, preventing oral carcinogenesis, reducing oral mucosa inflammatory reactions, and ameliorating metal-based restoration reactions. Oxidative stress is caused by an imbalance between the production of ROS and the body's ability to produce sufficient antioxidants and repair the resulting damage. The action and protective mechanism of a single antioxidant depends on the concentration, specific reactivity of the ROS, and condition of the antioxidant interaction

### CONCLUSION

Microbiological & biochemical parameters are assisted in smoking & non-smoking patients with or without carcinoma were assessed. On assessing the microbiological parameters, Micoplasmic bacteria was significant in chronic periodontitis with carcinoma patients. There was also a significant increase in herpes virus in plaque samples from chronic periodontitis with carcinoma patients when compared with chronic periodontitis. With respect to biochemical parameters, Reactive Oxidase metabolite (ROM) level was highly significant in carcinoma with chronic periodontitis patients. Various etiology with oxidative stress are few of the contributing factors for the malignancy. In the present study there was significant increase with respect to herpes virus and ROM in chronic periodontitis with malignancy patients. There was a relationship with malignancy and chronic periodontitis patients. Further longitudinal studies are needed before we could confirm the relationship with malignancy and chronic periodontitis patients.

### References

- 1. Arno, Development of a classification system for periodontal diseases and conditions. Ann Periodontol. 2009, Dec;4(1):1-6.
- 2. Bai XT, Socransky SS, Gunsolley JC. Systemic antiinfective periodontal therapy. A systematic review. Ann Periodontol. 2012 Dec;8(1):115-81
- 3. Beralet, Patel M, Socransky SS. Microbiological changes associated with four different periodontal therapies for the treatment of chronic periodontitis. J Evid Based Dent Pract. 1990 Sep;12(3 Suppl):50-60.
- 4. Bouvard: Anti inflammatory effects of macrolide antibiotics. Eur J Pharmacol 2009;429: 209 229,.
- 5. Boyle and Levin, Erakovic, V., Cepelak, I., Barisic, K., Brajsa, K., Ferencic, Z., et al. Azithromycin modulates neutrophil function and circulating inflammatory mediators in healthy human subjects. Eur J Pharmacol 2008; 450: 277–289.
- 6. Chen. H, Waddington RJ, Hall RC, Last KS. Connective tissue elements as diagnostic aids in periodontology. Periodontol 2000. 2003 Oct; 24:193-214.
- 7. Cheng, F. Periodontitis. Ann Periodontol2012;4:32-37.
- 8. D. B. Lowe., & Hand, D. L. Characteristics and mechanisms of azithromycin accumulation and efflux in human polymorphonuclear leukocytes. Int J Antimicrob Agents 2011;18:419-425.
- 9. D. S. Dane, Fu S, Stocker R, Davies MJ. Biochemistry and pathology of radical-mediated protein oxidation. Biochem J 1970; 324: 1-18.
- 10. De martel, Riley DP, Caputi AP, Salvemini D. Antioxidant therapy: a new pharmacological approach in shock,inflammation and ischaemia / reperfusion injury. Pharmacol Rev 2012; 53: 135-159.
- 11. De Oliveira, M.A. Lokuta, H.I.El-Shanthi, L.Muhle, X.Bing. Neutrophil dysfunction in a family with a SAPHO syndrome –like phenotype. Arthiritis and Rheumatism. 2010;58(10):3264-3269.

- 12. Eggo, PJ & Cutler, CW 2003, 'Microorganisms as risk indicators for periodontal disease'. Periodontology 2000, vol. 32, pp. 24-35.
- 13. Ehrlich, J, Cohen, GH & Hochman, N 1983, 'Specific herpes simplex virus antigen in human gingiva', Journal of Periodontology vol. 54, pp. 357-360.
- 14. Ellermann and Bang, Bullon P, Wilson M, Newman H. Oxidative injury and inflammatory periodontal diseases: the challenge of anti-oxidants to free radicals and reactive oxygen species. Crit Rev Oral Biol Med 1908; 10: 458-476.
- 15. Erlich, HA, Gelfand, D & Shinski, JJ 1991, Recent advances in the polymerase chain reaction (review) science 1991: 252: 1643-51.
- Grande, SR, Imbronito, AV, Okuda, OS, Lotufo, RFM, Magalhaes, MHG &Nunes, FD 2008, 'Herpes viruses in periodontal compromised sites: comparison between HIV-positive and -negative patients', Journal of Clinical Periodontology, vol. 35, pp. 838-845.
- 17. Graner, De Couto Pita AK, Busch L, Sánchez GA. Parameters of oxidative stress in saliva from patients with aggressive and chronic periodontitis. Redox Rep, 1999; Jun, 20:1-8.
- 18. Greenberg, M.S 1996, 'Herpesvirus infections', Dental Clinics of North America, vol. 40, pp. 359-368.
- 19. Halliwell B. Reactive oxygen species in living systems: source, biochemistry, and role in human disease. Am J Med 1991; 91 (Suppl. 3C): 14S-22S.
- 20. Hayward, S. D, Yashima A, Nagano T, Kanazashi M, Maeda N, Arai T. Effects of full-mouth scaling and root planing in conjunction with systemically administered azithromycin. J Periodontol. 2006 Mar;78(3):422-9.
- 21. John Burdell, Berk BC. Fyn and JAK2 mediate Ras activation by reactive oxygen species. J BiolChem 1848; 274: 21003-21010.
- 22. Kaldahl WB, Kalkwarf KL, Patil KD, Dyer JK, Bates RE Jr. Evaluation of four modalities of periodontal therapy. Mean probing depth, probing attachment level and recession changes. J Periodontol. 1988 Dec;59(12):783-93.
- 23. Kamma, JJ, Contreras, A & Slots, J 2001, 'Herpes viruses and periodontopathic bacteria in early-onset periodontitis', Journal of Clinical Periodontology, vol. 28, pp. 879-885.
- 24. Kling, I. Nakagawa, N. Okahashi, and N. Hamada, "Variations of Porphyromonasgingivalis fimbriae in relation to microbial pathogenesis," Journal of Periodontal Research, 1879; vol. 39, no. 2: 136-142.
- 25. Kubar, A, Saygun, I, Yapar, M, Ozdemir, A & Slots, J 2004, 'Real-time PCR quantification of cytomegalovirus in aggressive periodontitis lesions using Taq Man Technology', Journal of Periodontal Research vol. 39, pp. 81-86.
- 26. Kumar.V, Michelinaki M., &Kalpaxis, D. L. Insights into the mechanism of azithromycin interaction with an Escherichia coli functional ribosomal complex. MolPharmacol, 2004;59: 1441-1445
- 27. Loe, "Disruption of epithelial barrier and impairment of cellular function by Porphyromonasgingivalis," Frontiers in Bioscience, 1994; vol. 12, no. 10: 3965- 3974.
- 28. Loesche WJ, Giordano JR. Metronidazole in periodontitis V: debridement should precede medication. Compendium.1 994 Oct;15 (10):1198-1201.
- 29. Lundquist H. Isoluminol enhanced chemiluminescence: A sensitive method to study the release of superoxide anion from human neutrophils. Free Radical bio med. 1996;20:785-792.
- 30. Ly M. Free radicals, antioxidants and human disease : curiosity, cause or consequence. Lancet, 2014; 344: 721-724.
- Mc Laughlin drubin, Matsushima Y, Ujiie Y, Shirakawa S, Nagano T, Kanazashi M, Yashima A. Full-mouth scaling and root planing combined with azithromycin to treat peri-implantitis. J Periodontol. 2013 Nov;81(11):1555-63
- 32. Moeker, du Bois AH, Gannon S, Haynes DR, Hirsch RS. Antibacterial and immunomodulatory properties of azithromycin treatment implications for periodontitis. Inflammopharmacology. 2011 Aug;21(4):321-38.
- 33. Pauletto, du Bois AH, Gannon S, Haynes DR, Hirsch RS. Antibacterial and immunomodulatory properties of azithromycin treatment implications for periodontitis. J ClinPeriodontol. 2011 Feb;41(2):164-71.
- PetrovicSM ,Özdemir G, Tervahartiala T, Vural C, Atilla G, Baylas H, Sorsa T. Azithromycin as an adjunctive treatment of generalized severe chronic periodontitis: clinical, microbiologic, and biochemical parameters. J Periodontol. 2014 Dec;83(12):1480-91.

- 35. Poreba. I, Britigan B, Nakayama K, Grenier D. Cleavage of human transferrin by Porphyromonasgingivalisgingipains promotes growth and formation of hydroxyl radicals. Infect Immun. 2012 Aug; 72(8):4351-6.
- 36. Prato G.P, Gutteridge JMC. Free radicals in biology and medicine. Br J ExpPathol. 2002 Dec;70(6):737-57.