

Analysis of Fatty Acids in Acacia Catechu Seed Extract by Gas Chromatography Mass Spectrometry (Gcms)

Running title: Analysis of fatty acids in acacia catechu seed extract by gas chromatography mass spectrometry (GCMS)

Sindhu priya Kuppusamy

Undergraduate

department of pharmacology

Saveetha dental college

Saveetha university

chennai, 600077, Tamil Nadu, India.

Lakshmi Thangavelu

Assistant Professor of department of pharmacology

Saveetha dental college

Saveetha university

Chennai 600077 , Tamil Nadu, India.

Corresponding author

Lakshmi Thangavelu

Assistant Professor of department of pharmacology

Saveetha dental college

Saveetha university

chennai, 600077, Tamil Nadu, India.

Emails I'd: Lakshmi085@gmail.com

ABSTRACT

INTRODUCTION: Acacia seeds belongs to the family Leguminosae. Whereas, ACACIA catechu belongs to Fabaceae family. They posses various properties, such as; antioxidant, anti-cancer, anti-haemolytic, anti-inflammatory, anti-pyretic, analgesics and anti-depressant. It is used to treat, against, fever, malaria, cholera, dysentery, and even oral thrush. The many

functions are due to the presence of flavonoids, called catechin. And polyphenolic compounds such as; Rutin, caffeine acid, etc.

MATERIALS AND METHOD:

3 standard preparations were made with palmitic, linolenic and stearic acid, to calculate the fatty acid levels in acacia seeds using GCMS analysis.

RESULTS: The amount of fatty acids formed were substantial and showed that we can proceed extracting from Acacia seeds.

CONCLUSION: Fatty acids extracted from Acacia seeds can be used for various properties, such as; anti-inflammatory, malaria, fever, oral thrush, etc.

KEYWORDS: Acacia catechu, medicinal plant, fatty acids, polyphenolic compounds, linolenic acid, stearic acid, Palmitic acid.

INTRODUCTION

Acacia seeds belongs to family Leguminosae^[1] possesses antioxidant, anticancer, anti-haemolytic, anti-inflammatory, antipyretic, analgesic and antidepressant potentials. *Acacia catechu* Willd (*Fabaceae*), commonly known as catechu, cachou, and black cutch, is a moderate size deciduous, thorny tree widely distributed in India.

The name of the plant has recently been changed to *Vachellia karroo*^[2]. The gum produced by *A. karroo* is used against oral thrush and can also be harvested for food during hard times. *Acacia* is also effective against fever, malaria, cholera, diarrhoea, dysentery and high blood pressure. *Acacia* species are rich sources of polyphenolic compounds, known to have strong antioxidant properties that help in the prevention of various oxidative stress. These activities might attribute to the presence of various active secondary metabolites i.e. gallic acid, catechin, rutin, caffeic acid, 7-*O*-galloyl catechin, +catechin and methyl gallate. Flavonoids, a type of water-soluble plant pigments, are the major class of compounds isolated from *Acacia* plants. Catechin is a major flavan in *Acacia* bark and heartwood, found primarily in green tea.

Various parts of this plant have been used since ancient times in Ayurvedic medicine.^[3] Numerous natural bioactive compounds for instance 4-hydroxybenzoic acid, kaempferol, quercetin, 3,4,7-trihydroxyl-3,5-dimethoxyflavone, catechin, rutin, isorhamnetin,

epicatechin, afzelechin, epiafzelechin, mesquitol, ophioglonin, aromadendrin, and phenol have been isolated from heartwood, bark, roots, leaves and stem of *A. catechu* and presence of the above active compounds have been implicated for its myriad biological effects. The phytochemical isolated from this plant have been widely studied for their cytotoxic potentials against variety of cancer cell lines and came out with good results^[4,5,6]. *A. catechu* has been studied for its hepatoprotective, antipyretic, antidiarrheal, hypoglycaemic, anti-inflammatory, immunomodulatory, antinociceptive, antimicrobial, free radical scavenging, and antioxidant activities.

Extensive animal in vivo studies and human clinical trials compositions containing *Acacia* extract indicate that *Acacia* has great potential as a therapeutic agent for inflammatory diseases such as arthritis, irritable bowel syndrome, and inflammatory bowel syndrome^[7]. Catechu black extract has been approved by the US FDA for food use as a natural flavouring substance and/or natural substance used in conjunction with flavour.

Fatty acid is a carboxylic acid with a long aliphatic chain, which is either saturated or unsaturated. Most naturally occurring fatty acids have an unbranched chain of an even number of carbon atoms, from 4 to 28. Fatty acids are usually derived from triglycerides or phospholipids. Two essential fatty acids are linoleic acid (LA) and alpha-linolenic acid (ALA)^[8]. These fatty acids are widely distributed in plant oils. The human body has a limited ability to convert ALA into the longer-chain omega-3 fatty acids—eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), which can also be obtained from fish. Omega-3 and omega-6 fatty acids are biosynthetic precursors to endocannabinoids with antinociceptive, anxiolytic, and neurogenic properties.

Medicinal plants are currently of considerable importance because of their fatty acids which has potential therapeutic value that lead them to the path of development of novel drugs. Presence of beneficial fatty acids and the shift towards natural products in pharmaceutical and cosmeceutical industry made medicinal plant research equally important to conventional drug. methods like conventional solvent extraction, steam distillation, and sublimation, etc., are developed for extraction fatty acids.^[9] However, these methods are based on sequential extraction, including one or more organic solvents. Such phytochemical extracts need to be processed for the removal of traces of the organic solvents.

Furthermore, the mixture has to be purified for individuality. While such methods are useful for extraction and purification of small quantities of fatty acids for research purposes, completely removing the organic solvents from the extracts is a problematic issue. Furthermore, the types and concentrations of organic solvents must be carefully selected to avoid structural changes to the target phytochemical during extraction. Such changes adversely affect one or more of their desirable physical, chemical, and biological properties. Water, is an inexpensive, environment- friendly and an ideal solvent for the industrial extraction of medicinal plants, but its use is limited due to poor extraction efficiency for most organic compounds. The aim of this study is to assess and determine the amount of fatty acids in acacia seed extract.

MATERIALS AND METHOD:

PLANT MATERIAL COLLECTION AND EXTRACTION

Acacia catechu seeds were obtained from Hosur in Tamil Nadu and were processed by Green Chem lab in Bangalore.

PREPARATION OF ACACIA EXTRACT

Seeds were shade dried for a week. Dried seeds were milled to one powder. Powder was passed through 100 mesh sieve and stored in a sealed poly- thene bag 2.5 kg of powdered Acacia catechu seeds were extracted with 10 liters of Ethanol, at 65°C temperature, for 1 hour, in a 20 liter round bottom flask with Graham condenser attached. Condenser was cooled circulating with chilled water. After 1 hour of extraction, round bottom flask was cooled to room temp and the extract were altered and collected. e Marc was extracted repeatedly with 10 liters of Ethanol, twice. The extracts were altered and collected. The combined extracts was evaporated to dryness under reduced pressure in a Buchi Rotary Evaporator (Switzerland) at 65°C, to obtain 150 g of powder extract. The w/w yield of the prepared extract was 6%. The extract were stored at 4°C until used.

CHEMICALS AND REAGENTS

DPPH, BHA, Alpha glucosidase, Folin-Ciocalteu, Alpha amylase, Tris-Hcl buffer, P-nitro phenyl- glucopyranoside, Tryp- sin, starch, dinitrosalicylic acid, were procured from Sigma Chemicals , USA. Other re- agents like sodium carbonate perchloric acid, dimethyl sulfoxide (DMSO), sulphuric acid, ammonium molybdate and sodium phosphate were purchased from Merck, India .

Methodology:

Column Name: Elite-WAX; size: 30m (a film thickness of 1µm df), Inner diameter =0.32 mmID

Carrier gas : Helium (He)

Flow rate : 1.0 ml/min

Split ratio : 1:10

Injection Port : 250°C

Detection : EI- Mass (Temperature: Ion source-230°C; Interface- 250°C)

Column Oven : 80°C

Table 1: Column Temperature Program:

Rate (C/min)	Temperature	Hold Time (min)
-----	80.0	1
10.0	220.0	20

Standard Preparation 1: Weigh accurately about 50mg of Stearic acid (Stearic acid/Ref/01) in a clean Round bottom flask fitted with reflux condenser. Add 5ml of Boron trifluoride solution and reflux for 15 minutes until the solid is dissolved. Cool, transfer the reaction mixture with the aid of 10ml of Hexane in a separator and add 10ml of water and 10ml of saturated Sodium chloride solution. Shake well and allow the layers to separate then drain and discard the lower aqueous layer. Pass the hexane layer through anhydrous Sodium sulphate into a suitable flask.

Standard Preparation 2: Weigh accurately about 50mg of Palmitic acid (Palmitic acid/Ref/01) in a clean Round bottom flask fitted with reflux condenser. Add 5ml of Boron trifluoride solution and reflux for 15 minutes until the solid is dissolved. Cool, transfer the reaction mixture with the aid of 10ml of Hexane in a separator and add 10ml of water and 10ml of saturated Sodium chloride solution. Shake well and allow the layers to separate then drain and discard the lower aqueous layer. Pass the hexane layer through anhydrous Sodium sulphate into a suitable flask.

Standard Preparation 3: Weigh accurately about 50mg of Linoleic acid (Linoleic acid /Ref/01) in a clean Round bottom flask fitted with reflux condenser. Add 5ml of Boron trifluoride solution and reflux for 15 minutes until the solid is dissolved. Cool, transfer the reaction mixture with the aid of 10ml of Hexane in a separator and add 10ml of water and 10ml of saturated Sodium chloride solution. Shake well and allow the layers to separate then drain and discard the lower aqueous layer. Pass the hexane layer through anhydrous Sodium sulphate into a suitable flask.

Sample Preparation: Weigh accurately about 200mg of Acacia catechu seed extract in a clean Round bottom flask fitted with reflux condenser. Add 5ml of Boron trifluoride solution and reflux for 15 minutes until the solid is dissolved. Cool, transfer the reaction mixture with the aid of 10ml of Hexane in a separator and add 10ml of water and 10ml of saturated Sodium chloride solution. Shake well and allow the layers to separate then drain and discard the lower aqueous layer. Pass the hexane layer through anhydrous Sodium sulphate into a suitable flask.

Boron trifluoride preparation: weigh accurately 14gm of Boron trifluoride in a clean conical flask. Add 50ml of Methanol slowly. Mix well and make upto 100ml with Methanol.

Inject 1.0ul of Standard and Sample separately. Calculate the area of the Standard as compared to sample and calculate the assay.

Calculations:

1. Calculation of Stearic acid in *Acaica catechu* seed extract

Standard batch No:Stearic Acid/Std/Ref 01/97.6%

Weight of the Std:50.4mg

Standard Area:17422832

Sample Batch No:Acacia catechu seed extract

Weight of the sample:200.5mg

Sample Area:249495

Extraction solvent:Boron trifluoride solution

Std Assay:97.6%

Calculation:

$$(50.4 / 200.5) \times (249495 / 17422832) \times 97.6\% = 0.35\%$$

2. Calculation of palmitic acid in *Acacia catechu* seed extract

Standard batch No:Palmitic Acid/Std/Ref 01/98.8%

Weight of the Std:57.6mg

Standard Area:98551566

Sample Batch No:Acacia catechu seed extract

Weight of the sample:200.5mg

Sample Area:1280956

Extraction solvent:Boron trifluoride solution

Std Assay:98.8%

Calculation:

$$(57.6 / 200.5) \times (1280956 / 98551566) \times 98.8\% = 0.36\%$$

3. Calculation of Linoleic acid *Acacia catechu* seed extract

Standard batch No:Linoleic Acid/Std/Ref 01/75%

Weight of the Std:50.8mg

Standard Area:26727141

Sample Batch No:Acacia catechu seed extract

Weight of the sample:200.5mg

Sample Area:990825

Extraction solvent:Boron triflouride solution

Std Assay:97.6%

Calculation:

$$(50.8 / 200.5) \times (990825 / 26727141) \times 75\% = 0.70\%$$

RESULTS:

Sample Information

Analyzed by : Admin
 Analyzed : 12/9/2014 12:34:35 PM
 Sample Name : Acacia catechu seed extract
 Sample ID : Acacia catechu seed extract
 Vial # : 2
 Injection Volume : 1.00
 \$EndIf\$Data File : D:\GCMS DATA\Data\FT_29102014\Fatty Acids 17.qgd
 Method File : D:\GCMS DATA\Method\Fatty Acids scan.qgm
 Report File : D:\GCMS DATA\Report\Default.qgr
 Tuning File : D:\GCMS DATA\Tune\Fatty acids tune CID gas off.qgt

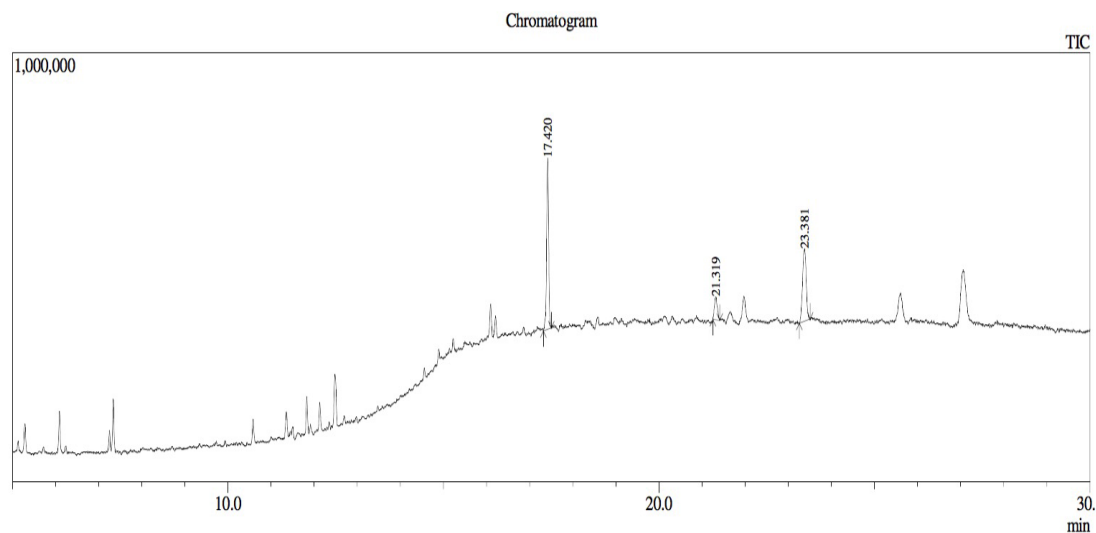


Table 2: peaks showing fatty acid level in *Acacia seeds*, using GCMS

Peak Report TIC								
Peak#	R.Time	I.Time	F.Time	Area	Area%	Height	Height%	Name
1	17.420	17.320	17.505	1280956	50.81	394908	64.20	Methyl palmitate
2	21.319	21.250	21.415	249495	9.90	52896	8.60	Methyl stearate
3	23.381	23.260	23.510	990825	39.30	167304	27.20	Methyl linoleate
				2521276	100.00	615108	100.00	

Figure 1 : Results by GCMS

DISCUSSION

Antioxidant molecules are gaining lot of interest in this respect. Antioxidants are also attracting the attention of the scientific community because of their potential to be used as a drug and supplement to drug in case of many degenerative diseases to minimise the complications. Synthetic antioxidant like butylatedhydroxy anisole (BHA) or butylatedhydroxy toluene (BHT) or propyl gal-late (PG) are supplemented to reduce the oxidative damage but due to side effect these are not well accepted antioxidant.^[10] It is generally assumed that frequent consumption of plant-derived phytochemical from vegetables, fruit, tea and herbs may contribute to shift of balance towards an adequate antioxidant status^[11]. The extraction of these bioactive phytochemical uses organic solvents but the organic solvents lead to the issue of safety and acceptability by human body.

A.catechu has a high therapeutic value as it helps to relieve the symptoms of diseases like lymphatic and venous insufficiency, hemorrhagic diseases, and hypertension. *A.catechu* was found to induce apoptosis in LNCaP cells (Prostate cells) at low concentration and in HepG2 cells (human hepatoma) at very high concentration by acting as pro-oxidant instead of an antioxidant which resulted in programmed cell death. It inhibited the tumour cell growth by blocking cells amelioration through G0-G1 transition which is mediated by down regulation of cyclin D1 and E. *A.catechu* also found to exhibit excellent antibacterial property against *E. coli* strain (incorporated with *envA1* allele) by inhibiting the action of enzyme type II topoisomerase as well as its selective promotion of *E. coli* topoisomerase IV-dependent DNA cleavage which is very essential for the survival of the cell.

Further, *A. catechu* is proven for its cytotoxic potentials against various cell lines.^[12] These findings suggest the fact that active phytochemical components present in *A. catechu* are responsible for the mechanism by which they induce cytotoxicity and our current results are in agreement with these reports. Staining of apoptotic cells with fluorescent dyes such as AO/EB is considered one of the methods for evaluating the nuclear morphology changes.^[13,14] Previous studies have performed AO/EB staining and reported that early apoptotic cells had fragmented DNA which exhibited intense green coloured nuclei. Late apoptotic and necrotic cells DNA were fragmented and stained orange and red.^[15] In light of

the above reports, the presence of high orange stain intensity in cells treated with high concentration of ACB extract further confirm the DNA fragmentation and apoptosis.

A.catechu has been previously reported for its anti-HIV activity.^[16] The present authors have previously demonstrated that *A. mellifera* ethyl acetate, n-butanol and aqueous extracts also have hepatoprotective and anti-HBV effects.^[17] In the present study, anti-HBV evaluation of *A. oerfota* extracts at a non-cytotoxic dose showed its association with the methanol and aqueous extracts. Furthermore, the highest anti-HBV activity of *C. grandis* and *C. epigeus* was associated with the crude ethanolic-extract, indicating the possibility of synergy among the antiviral phytochemical constituents of the extract. Synergistic activity of antiviral components of plant extracts that act by different mechanisms has been reported previously.^[18]

The antimalarial activity of *A. Catechu* extracts (leaves, pods and bark) were determined by the schizont maturation inhibition assay. Ethanol extract S of leaves, pods and bark were screened for antimalarial activity against an artesunate sensitive strain of *P. falciparum* (3D7). The schizonticide activities were expressed in terms of IC₅₀ values, defined as minimum concentration of plant extracts required to inhibit 50% of schizont maturation. The schizont maturation inhibition results at 48 h and 96 h by artesunate, leaves, pods and bark extracts. After 48 h of incubation, leaves were found more potent than pods and bark against the plasmodium parasite with an IC₅₀ of 1.29 (1.08–1.49) µg/ml.^[20]

CONCLUSION:

Fatty acids extracted from Acacia seeds can be used for various properties, such as; anti-inflammatory, malaria, fever, oral thrush, etc.

REFERENCE:

1. Ambasta SP. The useful plants of India, publication and information directorate. New Delhi: Council of Scientific & Industrial Research; 1994.4(5) 113-187
2. Maldini M, Montoro P, Hamed AI, Mahalel UA, Oleszek W, Stochmal A, Piacente S. Strong antioxidant phenolics from *Acacia nilotica*: profiling by ESI-MS and qualitative–quantitative determination by LC–ESI-MS. J Pharm Biomed Anal. 2011;56(2):228–239.

3. Hussein G, Miyashiro H, Nakamura N, Hattori M, Kakiuchi N, Shimotohno K. Inhibitory effects of Sudanese medicinal plant extracts on hepatitis C virus (HCV) protease. *Phytother Res.* 2000;14(7):510–516.
4. Ahmad I., Beg A.Z. Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi drug resistant human pathogens. *J. Ethnopharmacol.* 2001;74:113–123.
5. Alam, P., Alajmi, M.F., Arbab, A.H., Parvez, M.K., Siddiqui, N.A., Alqasoumi, S.I., Al-Rehaily, A.J., Al-Dosari, M.S., Basudan, O.A., 2017. Comparative study of antioxidant activity and validated RP-HPTLC analysis of rutin in the leaves of different *Acacia* species grown in Saudi Arabia. *Saudi Pharm. J.* 25 (5), 715–723.
6. Alam P., Alajmi M.F., Siddiqui N.A., Al-Rehaily A.J., Alharbi H., Basudan O.A., Hussain A. Densitometric validation and analysis of biomarker β -amyrin in different *Acacia* species (leaves) grown in Kingdom of Saudi Arabia by high performance thin-layer chromatography. *Pak J Pharm Sci.* 2015;28:1485–1491.
7. Bouhlef I., Limem I., Skandrani I., Nefatti A., Ghedira K., Dijoux-Franca M.G., Leila C.G. Assessment of isorhamnetin 3-O-neohesperidoside from *Acacia salicina*: protective effects toward oxidation damage and genotoxicity induced by aflatoxin B1 and nifuroxazide. *J. Appl. Toxicol.* 2010;30:551–558.
8. Chatterjee A., Pakrashi S.C. NISCAIR Press; New Delhi: 2000. The Treatise on Indian Medicinal Plants; 2015;42:51–53.
9. Chatti I.B., Boubaker J., Skandrani I., Bhourri W., Ghedira K., Chekir G.L. Antioxidant and antigenotoxic activities in *Acacia salicina* extracts and its protective role against DNA strand scission induced by hydroxyl radical. *Food Chem. Toxicol.* 2011;49(8):1753–1758.
10. Husain F.M., Ahmad I., Khan M.S., Al-Shabib N. *Trigonella foenum-graceum* (Seed) extract interferes with quorum sensing regulated traits and biofilm formation in the strains of *Pseudomonas aeruginosa* and *Aeromonas hydrophila*. *Evid. Based Complement Alternat. Med.* 2015;9(7):332-350
11. Joy P.P., Thomas J., Mathew S., Skaria P. Medicinal Plants Kerala Agricultural University Publications; Kerala, India: 1998. 51, 334-356
12. Kritkar K.R., Basu B.D. Indian Medicinal Plants with Illustrations. Uttaranchal Oriental Press; 2003. 4(5)1289–1292.
13. Kumar B.P., Singh R. Antidiabetic activity of *Acacia tortilis* (Forsk.) Hayne ssp. *raddiana* polysaccharide on streptozotocin-nicotinamide induced diabetic rats. *Biomed. Res. Int.* 2014;20, 503-517.

14. Musthafa K.S., Ravi A.V., Annapoorani A., Packiavathy I.S.V., Pandian S.K. Evaluation of anti-quorum-sensing activity of edible plants and fruits through inhibition of the N-acyl-homoserine lactone system in *Chromobacterium violaceum* and *Pseudomonas aeruginosa*. *Chemotherapy*. 2010;56:333–339.
15. Nadkarni K.M. The Indian Plants and Drugs. New Delhi: Shrishti Book Distributors. 2005;4, 445-507
16. O'Toole G.A., Kolter R. Initiation of biofilm formation in *Pseudomonas fluorescens* WCS365 proceeds via multiple, convergent signaling pathways: a genetic analysis. *Mol. Microbiol.* 1998;28:449–461.
17. Perez C., Pauli M., Bazerque P. An antibiotic assay by the well agar method. *Acta Biol. Med. Exp.* 1990;15:113–115.
18. Nadumane VK, Nair S. Evaluation of anticancer and cytotoxic potentials of *Acacia catechu* extracts *in vitro*. *J Nat Pharm.* 2011;2:190–5.
19. Lakshmi T, Ezhilarasan D, Upendra N, Vijayaragavan R. *Acacia catechu* ethanolic seed extract triggers apoptosis of SCC-25 cells. 2013;45;1112-1115
20. Savitskiy VP, Shman TV, Potapnev MP. Comparative measurement of spontaneous apoptosis in pediatric acute leukemia by different techniques. *Cytometry B Clin Cytom.* 2003;56:16–22.

Table legends:

Table 1 –column temperature program

Table 2 - peaks showing the fatty acid level in *Acacia seeds*, using GCMS analysis

Figure 1 - peaks showing the fatty acid level in *Acacia seeds*, using GCMS analysis