

## Isolation, Characterization of *Lactobacillus Delbrueckii* derived Bacteriocin and Assessment of its Bioactive Potential

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### ABSTRACT

A *Lactobacillus delbrueckii* strain that produces bacteriocin was isolated from the vagina of a healthy person. A multitude of bacterial species, including *Pseudomonas*, *Staphylococcus*, *Salmonella*, and *Enterococcus*, were inhibited by the supernatant fluid. Bacteriocin is an antimicrobial peptide which was isolated from *Lactobacillus delbrueckii*. A single four-step process was carried out to purify it to homogeneity. MALDI-TOF investigation was performed with the crude supernatant acquired from an initial stationary phase culture in the MRS broth. Bacteriocin was proteolytic enzyme sensitive, heat resistant, functional over a broad pH range, and drifted as a peptide of 4.5–5.0. kDa in the SDS-PAGE. During exponential growth, the bacteriocin was formed continuously. It was bactericidal to sensitive cells, and cell lysis did not achieve the bactericidal impact. The bacteriocin's amino acid composition was measured, and no modified amino acids were discovered among the residues found.

### Keywords:

*Lactobacillus delbrueckii*, MALDI-TOF analysis, Bacteriocin

### Introduction

Bacteriocins are peptides or proteins that are made by ribosome which has medicament activity against strains from various species or genera (De Vuyst and Leroy, 2007). However, a growing variety of innovative bacteriocins towards foodborne pathogens are well-known (Liu et al., 2015; Yi et al., 2016; Zhao et al., 2016). Bacteriocins developed by lactic acid producing bacterial communities (LAB) have recently received loads of attention because of their potential as safe and economical bio-preservative agents. It's conjointly documented that bacteriocins are proficient of constraining the progression of foodborne pathogens such as *Staphylococcus aureus*, *Escherichia coli*, and *Listeria monocytogenes*.

Bacteriocins are normally used for natural food preservation and protection in several countries also as being safe for humans (Liu et al., 2016, 2017; Yi et al., 2016, Cotter et al., 2013; principle et al., 2014). Since its 1st exploitation in England in 1953, the bacteriocin nisin that was discovered in New Island in 1933 has been approved to be used in forty eight countries (Tagg et al., 1995). Bacteriocins are often classified into 3 main classes based on their structural, physicochemical, and molecular properties. Lantibiotics are small category 1 bacteriocins, i.e. cationic, heat-stable amides, hydro-phobic, and that comprise uncommon amino acids that are synthesized during the post-translation modification. Apart from the cleavage of a frontrunner peptide from the pre-bacteriocin peptide, category two bacteriocins are small, cationic, hydrophobic, heat-stable peptides that don't seem to be post-translationally modified. There are three subclasses beneath this group: Bacteriocins belonging to 2a subclass, or pediocin-like bacteriocins, with an impact on food borne pathogenic communities and the consent sequence Tyr-Gly-Asn-Gly-Val in N-terminus domain; Subclass 2b, which are bacteriocins which need dual polypeptide chains for the maximum action, and Subclass 2c, are the bacteriocins which is

not belonging to subgroups. Bacteriocins of class 3 is a bulky, thermo-labile, and hydrophilic molecules. Therefore in the present investigation, an effort was taken to develop, refine, and characterize bacteriocin from *Lactobacillus delbrueckii*, a human vaginal bacterium. Purified bacteriocin was also put to the test for antimicrobial activity against possible pathogens. Other bioactivities, apart from antimicrobial activity, have been extensively examined. Purified bacteriocin was also put to the test for antimicrobial activity towards possible pathogenic microorganisms. Other bioactivities, in addition to antimicrobial potential, have been extensively examined.

## MATERIAL AND METHODS

**Bacterial Strains used and its culture conditions:** Serial dilution and plating technique in MRS medium was performed to evaluate the growth of *L. delbrueckii*. The *L. delbrueckii* strains were cultured and maintained in the liquid of MRS complex medium (Difco). For the sub culturing, fresh culture cells with the optical density measurement of 0.1 at 600 nm ( $OD_{600}$ ) that were first washed in 0.9% NaCl for the inoculation of liquid cultures in MRS Broth.

**Determination of organic acids by *L. delbrueckii* and acidity:** The acid produced by *L. delbrueckii* was tested for quality and quantity. According to previous studies, the quantities of organic acids released by *L. delbrueckii* were measured by the difference between the volume of organic acid in the supernatants earlier and later of the fermentation process. Each acid's concentrations were measured in millimoles per liter. The experiment was performed in triplicates to conclude the final values. Cells (or cell-free extract) from *L. delbrueckii* was resuspended in the cold phosphate buffer with the molarity of 0.2 M and with a corresponding pH 6.5 with or without 55.5 mM glucose and incubated at 5°C for the defined period. After the incubation period, the cells were centrifuged to be quantified.

**Hydrogen peroxide production by *L. delbrueckii*:** *L. delbrueckii*'s cell-free supernatant (5 mL) from cells were combined with 1 mL of 0.1 per cent aqueous peroxidase (Horseradish Type VI-A; Sigma Chemical) following that 0.1 mL of 1 % of o-dianisidine aqueous solution was added. As a blank, phosphate buffer (5 mL) was used as an alternative of the experiment sample. The tubes were then further incubated at 37°C for 10 min. To stop the reaction, 0.5 mL of 2N HCl was added to the tubes. The optical density at 400 nm ( $OD_{400}$ ) of the test samples were assessed, and the peroxide quantity was measured by comparing the  $OD_{400}$  nm absorbance with a standard plot (Gilliland, 1969).

**Synthesis of crude Bacteriocin:** In MRS broth, *L. delbrueckii* was cultured aerobically for 24 hours at 35°C (Hi-Media, India; pH 6.5). To remove the cells from the growth media, they were centrifuged in the following condition 8,000 rpm for 30 min at 4 °C and separated with 0.22 µ filters after the incubation period. The supernatant's pH was modified using 1N NaOH to 6.0. The resultant was used as crude bacteriocin.

**Assessment of Bacteriocin Activity:** The agar well diffusion assay was performed to assess bacteriocin activity. *L. delbrueckii* was seeded on Mueller-Hinton agar plates. Wells with a diameter of 6 mm were drilled, and aliquots (20 µL) of the supernatant were inoculated into the wells and kept for 24 hours. The diameter of growth reticence was evaluated after incubation period of 24h. The arbitrary units per milliliter (AU/ml) were used to calculate the activity of the cell-free supernatant. The bacteriocin's unit activity was defined as 1 AU, which is equal to a unit area of zone of inhibition per unit volume, in this case 2 mm/ml (Usmiati & Marwati 2009). The assay was performed as replicate 3 times. The bacteriocin activity was calculated according to the standard formula.

**Partial characterization of bacteriocin:** *L. delbrueckii* was cultured in MRS broth for 24 to 48 h at 37 °C before being partly characterized. The grown cells were centrifuged at 8,000 rpm for 1 h at 4°C after incubation. The precipitation of Bacteriocins was done with ammonium sulfate (80 per cent saturation) and kept overnight at -20 °C for augmented precipitation. The entire mixture was centrifuged after precipitation and pellets were obtained and stored in a potassium phosphate buffer with a pH of 7.2. The precipitate was dialyzed at 4°C for 12 hours against a 20 mM potassium phosphate buffer (pH 7.0). (2010 by Rajaram et al).

**Influence on Bacteriocin Activity by Temperature:** Various heat treatments were used for dialyzed bacteriocin (pH 5.5, 500 l): 30°C to 100°C for 30 min and 121°C for 15 min. *Bacillus cereus* was used as an indicator organism to evaluate the activity of bacteriocin.

**Influence on Bacteriocin Activity by pH:** The effect of pH was investigated using bacteriocin (pH 5.5, 50 l) mixed with 100 l of buffer (0.1M potassium phosphate pH 4 to pH 9) and incubated at 60°C for 1 hour before determining bacteriocin activity against *Bacillus cereus*.

**Influence on Bacteriocin Activity by detergents:** sodium dodecyl sulfate, Tween 80, and Triton-X100 surfactants were applied to the bacteriocin solutions in concentrations of 100 µL or 0.01 g/ml (pH 5.5). Bacteriocin activity against *Bacillus cereus* was determined by incubating these preparations at 60°C for 60 minutes.

**Influence on Bacteriocin Activity by enzymes:** On bacteriocin preparations, the enzymes D-amylase, lysozyme, protease, and proteinase-K were each measured at a final concentration of 1 mg/ml (pH 5.5). It was then incubated

For 2 hours at 60°C to assess bacteriocin activity against the pathogens.

**Assessment of Bacteriocin activity:** The spot-on-lawn approach was used to measure arbitrary units (AU) of bacteriocin activities (Yamamoto et al. 2003). The previously mentioned method was used to obtain cell free supernatant. Filter sterilized 20 mM potassium phosphate buffer (pH 6.5) was used to dilute the resulting sample twofold. After applying 100 µl of *L. delbrueckii* overnight culture, soft agar (20 ml) was solidified in a sterile petri dish. After 30 minutes of drying, 10 µl of each diluted sample was spotted onto the plate and incubated at 37°C overnight. The titer was measured as the reciprocal of the highest dilution (2<sup>n</sup>) that caused the indicator lawn to be inhibited. In consequence, the antibacterial activity per mL was defined in AU as  $2^n \times 1000 \mu\text{L} / 10 \mu\text{l}^{-1}$ .

**Bacteriocin Passive Elution from Polyacrylamide Gel:** *Lactobacillus delbrueckii* crude proteins were fractionated using the Schagger and Von Jagow method of Tricine-SDS-PAGE (16 percent polyacrylamide gel). The gels were stained with Coomassie brilliant blue after electrophoresis was completed. The gel was eroded three times in 0.1 percent Tween 80 at 37°C to eliminate SDS and spot the bands of protein with antibacterial activity. Then, using Martinez et al process, soft agar medium (10 ml) with test pathogens (approximately  $1 \times 10^6$  to  $2 \times 10^6$  cells/ml) was shrouded to overlay the entire gel (6 by 9 cm). Subsequently overnight incubation at 30°C, clear zones around the protein bands were found, suggesting that the protein has antibacterial activity.

**Bacteriocin Identification by Mass Spectrometry:** The entire mass spectra of the bacteriocin was identified using a rectilinear MALDI-TOF mass spectrometer with delayed extraction technology with a 125-cm flight tube. For desorption/ionization of the samples, whole spectra were acquired in positive ion linear mode with a nitrogen laser (15 337 nm) and a 20 kV acceleration voltage. The spectra are the product of a sequence of 60 laser shots. Angiotensin II and insulin bovine were used for external mass calibration (MH1 5 5,734.557; Sigma). For each sample preparation phase, the instrument was calibrated.

### **Bioactive potential of bacteriocin isolated**

***In vitro* fungicidal effect of *L. delbrueckii* bacteriocin:** At five and 20°C, the performance of *L. delbrueckii* bacteriocin in opposition to fungal pathogens became evaluated the use of the microdilution approach described through Lavermicocca et al. (2003) with a few modifications. At 80°C, the inventory cultures had been saved as spore or mycelial suspensions in 15% glycerol (v/v). The fungi's conidia had been accumulated from four day-vintage cultures grown at 25°C on potato dextrose agar (PDA). The fungi's conidia had been accumulated from 12-day-vintage cultures grown on Pseudomonas Agar F at 22°C below a 12 h light/darkish cycle. The fungi's conidia had been accumulated from 12-day-vintage cultures grown at 25°C on changed V-8TM medium. Using sterile distilled water, the density of spore suspensions became diluted to two one zero five spores ml<sup>-1</sup>, as calculated through a hemocytometer. In sterile 96-well microdilution plates, microdilution checks had been performed. The microdilution plates had been incubated at both 20 and 5°C after two hundred 1 of take a look at answer containing 185 1 of *L. delbrueckii* bacteriocin and 15 1 of conidial suspension became dispersed into the wells. Using a microtiter plate reader/spectrophotometer, the optical density (OD) became calculated at 580 nm in specific time intervals (Spectra MAX 190, Molecular Devices, CA, USA). The OD of control plates incubated at 20°C became measured at 0, 24, 40, 48, or seventy two hours, at the same time as the OD of tradition plates incubated at five°C became measured at 0, 48, seventy two, 96, or one hundred twenty hours. The test became replicated 3 times. (185 1 MRS broth and 15 1 conidial suspension) and a poor control (185 1 LAB CFS and 15 1 lifeless conidial suspension) had been organized and maintained for every fungus. The test became replicated 3 times.

**Cell culture :** HepG2.2.15 cells were grown in RPMI 1640 complemented by 10% fetal bovine serum and 1% (v/v) penicillin (10,000 U/ml)/streptomycin (10,000 U/ml) under 5% CO<sub>2</sub> at 37°C. Trypsin therapy was used to remove sub-confluent monolayer cells from culture dishes, which were then transferred to fresh cell culture dishes. The blue dye exclusion system of trypan was used to estimate cell number (Strober 2001).

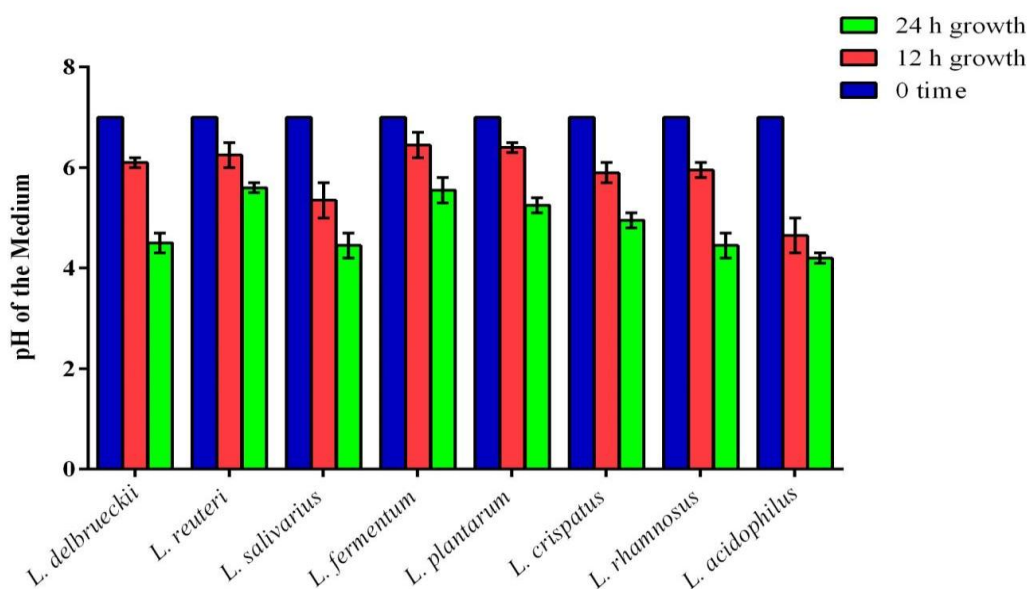
**Anti-cancer activity of bacteriocin:** Five separate human cancer cell lines, including HeLa, MCF-7, HT1080, H1299, and HEK293T, were used to assess the anticancer efficacy of *L. delbrueckii* bacteriocin. The cell lines were incubated with increasing concentrations of bacteriocins (1–20 M) in the specific growth medium, revealing that LS10 has cytotoxic efficacy against all five cancer cell lines in nature. Whole cancer cell lines showed cytotoxicity based on the dose of bacteriocin; highest effect was observed at 10 µM, and the highest activity was observed against the MCF-7 cell line. Although MCF-7 showed 40% cytotoxicity at a 5 µM concentration of LS10, HT1080, HEK293T, and H1299 showed just 20% cytotoxicity. However, at a concentration of 5 µM, no major cytotoxicity was observed in HeLa cells, but at a concentration of 10 µM, cytotoxicity was observed in 80 percent of HeLa and other cell lines.

### **Results and Discussion**

**Measurement of organic acids production and acidity by *L. delbrueckii*:** The results revealed that the complex MRS broth medium yielded the most lactic acid (300–550 mmol l<sup>-1</sup>). It's apparent because of the media's ideal composition, which was designed especially for the *Lactobacillus bacterium* glucose is the major carbohydrate source in the media fermented two pathways by *L. delbrueckii*.

Lactic acid is the major product when glucose is used as the main substrate in the metabolic pathways shown by our findings. Adding to that, glycolysis generates 2 mol of lactic acid is formed from 1 mol of glucose, while the 6-PG/PK pathway produces just 1 mole of lactic acid from 1 mol of glucose. Based on our findings from this study, the strain *L. delbrueckii* appears to

be hetero-fermentative in several conditions, but certain other factors can play a role. Figure 1 the result shows the quantification of lactic acid produced by the isolates. *L. acidophilus* yielded the huge amount of lactic acid after 24 hours, according to the results.



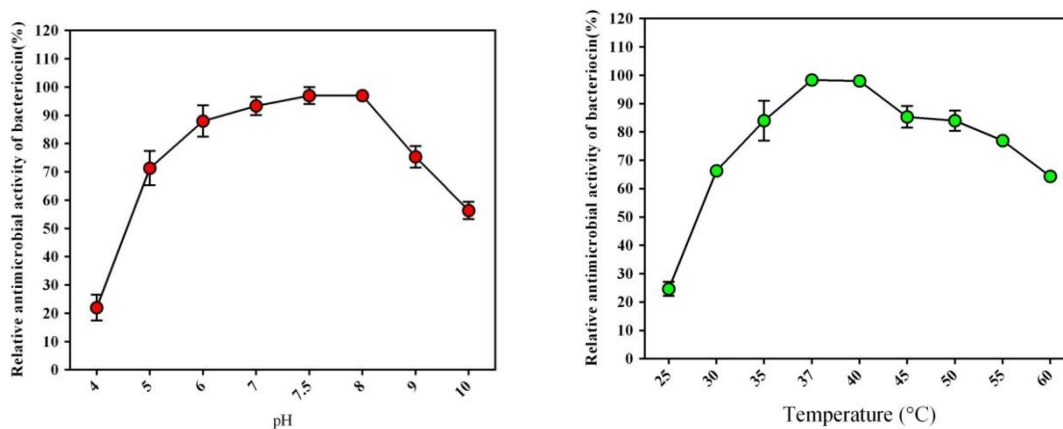
**Figure 1 –production of organic acid by the isolates in the 24 h of incubation**

**H<sub>2</sub>O<sub>2</sub> production by the isolate:** Hydrogen peroxide, an antagonistic chemical, is also used in lactobacilli. Quantitative approaches are typically used to assess the performance. In the current study, the isolate *L. delbrueckii* produced the highest amount of H<sub>2</sub>O<sub>2</sub>, 3.18 mg/ml. According to the findings, lactic acid production improved as the incubation time increased from 24 to 72 hours.

**Determination of bacteriocin activity:** Since During the mid phase, the highest activity was found, and the bacteriocin was isolated from *Lactobacillus delbrueckii* culture during this period. The secreted bacteriocin's highest antimicrobial activity was strain-specific, ranging from 350 to 520 AU/ml. When trypsin loses its bactericidal function, it indicates that the activity is attributed to a protein. As a result, using commercial nisin as a reference, the bacteriocin activity was measured in Arbitrary Units/ml (AU/ml). Figure 3.2 depicts the normal graph for nisinbacteriocinogenic activity. The only disadvantage is that, because of its hydrophobic nature, nisin diffusion inside agar media is only corresponding to its concentration over a narrow linear array (Chandrapati et al., 1998). The non-linear norm chart, on the other hand, is also useful for assessing other bacteriocinogenic behavior.

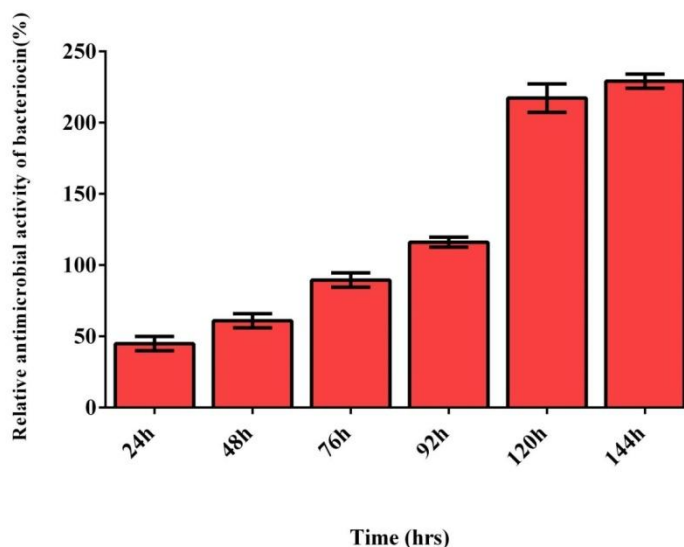
**Sensitivity of bacteriocin to pH, enzymes and temperature:** When incubated for 15 and 30 minutes at 60, 80, and 100 degrees Celsius, no substantial variations in antimicrobial activity were observed (P 0.05). (Table 4.23).Bacteriocin were stable at 37°C with pH of 2.0 to 10.0 after incubation (P0.05) showed that bacteriocin be able to treat both acidic and non-acidic foods. Bacteriocin's antimicrobial efficacy was not inactivated at high temperatures, such as boiling (60°C, 30 minutes) or boiling (100°C, 45 minutes). Catalase therapy did not result in a loss of bacteriocin function, suggesting that H<sub>2</sub>O<sub>2</sub> was not the source of reticence activity. Bacteriocin treatment with enzymes like proteinase K and pepsin forms a absolute loss of bacteriocin activity, suggesting that the effective portion of bacteriocin was protein in nature, while non-treated bacteriocin had a substantial (P0.01) bacteriocin activity. The bacteriocin action was slightly

reduced by the presence of -amylase, DNase, and RNase. These findings indicate that the biocontrol agent was protein in nature and that its action was solely based on glycosylation, implying that it was active in cooperation with the glycol and protein portions. Sodium dodecyl sulphate (SDS), Tween 80, EDTA, Tween 20, and urea were used as detergents. Tween 80, SDS, and Tween 20 have no effect on bacteriocin activity, which means that these detergents have no effect on bacteriocin activity. The bacteriocin activity was greatly decreased when the assay was done after treatment (P0.05). (See Figure).



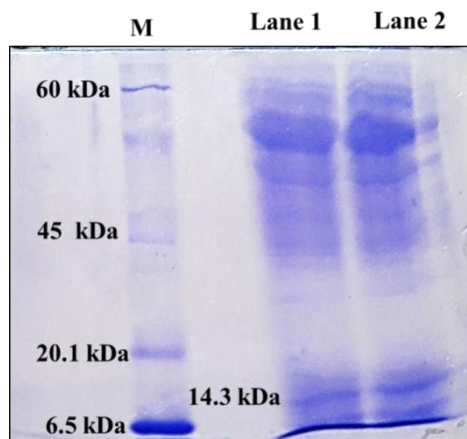
**Figure 2 A - Stability of bacteriocin at various pH Figure 2 B - Temperature stability of the bacteriocin**

The lack of bacteriocin activity reduction or loss after detergent treatment may be attributed to non-denaturation of the bacteriocin's connection with other molecules, which stabilizes bacteriocin activity. Bacteriocin action was steady at 37°C with pH of 2.0 to 10.0 after incubation (P0.05). This shows that bacteriocin be able to treat both acidic and non-acidic foods. Bacteriocin's antimicrobial efficacy was not inactivated at high temperatures, such as boiling (60°C, 30 minutes) or boiling (100°C, 45 minutes).



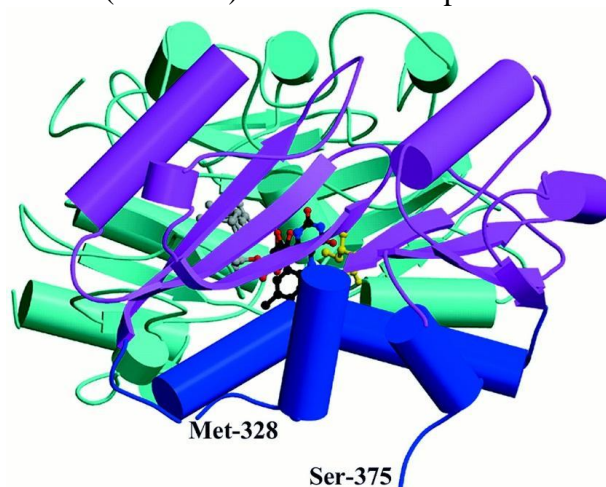
**Figure 2 C- Stability of bacteriocins with time interval**

**Molecular weight determination of bacteriocin:** The bacteriocin's molecular weight was measured by Tricine-SDS PAGE (Fig. 4.8). The protein band was visible when dyed with Coomassie blue, and bacteriocin's antimicrobial activity was linked to a band with the respective molecular weight of around 14.5kDa.



**Figure 3 - Tricine SDS-PAGE analysis of bacteriocin: Lane1 indicates the marker and lane 2, 3 indicates bacteriocin.**

**Amino acid sequence of bacteriocin:**The N terminus of distilled bacteriocin begins with valine and ends with lysine, according to the amino acid series. This band's amino acid analysis yielded the following partial sequence: "VYAEPQAQNIPWVK." The molecular mass was found to be 14300 Da, with a pH of 8.77 as the isoelectric point. The bacteriocin had four hydrophobic residues and a total charge of two. The Bactibase Database was used to determine the theoretical isoelectric point 5.97 (pI), molecular weight, amino acid composition, grand average of hydropathicity of the peptide chain (GRAVY) and atomic composition.



**Figure 4 –The structure prediction of bacteriocin from *Lactobacillus delbrueckii***

**Bacteriocin Identification by MALDI/ToF:**Membranes to adsorb bacteriocins were successfully analyzed using MALDI-TOF MS. The on-target washing process, on the other hand, has been shown to be a useful and efficient technique for the revealing of bacteriocins by MALDI-TOF MS. Figure 4 displays the spectrum of supernatant taken from the culture of *Lactobacillus delbrueckii*. Some pollutants were removed with a ten-second wash with milli-Q

water; however, a 30-second wash was found to be the most effective, resulting in a peptide signal with greater power, improved resolution, and less noise. The 60-second wash eliminated the pollutants as well, but it seemed to weaken the sample signal. Despite the fact that some of the samples had low signal strength and resolution. With signals ranging from 3,350 to 3,360 Da, the m/z ratios of all the samples were close to the mass predicted for nisin (MH<sup>+</sup> = 3,354 0.1 percent). The single band on native-PAGE illustrated the homogeneity of the distilled bacteriocin. Gel filtration was used to assess the molecular mass of bacteriocin, which was found to be 14.3 kDa. The monomer bacteriocin has a molecular mass of 14.3 kD, as calculated by MALDITOF-MS fingerprinting, SDS-PAGE, and gel filtration analysis. The molecular mass of bacteriocin was found to be close to that of other bacteriocins reported from Bacilli, such as Colicins, a large bacteriocin with a various molecular weight (25-80kDa) that has also been identified as an endopeptidase that exerts its inhibitory action by damaging the cell wall of sensitive bacteria, thus being classified as a cell wall degrading bacteriocin. Using the MASCOT and PROWL servers, the observed mass and relative intensities obtained from peptide mass fingerprinting were used to classify the parent sequence against profiles of recognized sequences (Figure 5.3-5.4). The isolated bacteriocin was found to be identical to *Lactobacillus delbrueckii* uracil-5-methyltransferase (MASCOT search score: 68) with a molecular weight of 50kDa. A number of high molecular weight bacteriocins that target sensitive bacteria's cell walls were also well defined. These bacteriocins, also known as endopeptidase or bacteriocins, were graded as class III bacteriocins (Clara et al., 2012). FemABX-like proteins function as peptidyltransferases, allowing amino acids to be incorporated into the peptidoglycan's interchange peptide connection, resulting in the targeted bacterial indicator's cell wall lysis. The amino acid sequence of bacteriocin was compared to the protein sequence of *Lactobacillus delbrueckii* uracil-5-methyltransferase using MALDI-TOF-MS. The incomplete N terminal sequence of bacteriocin (the first 14 amino acid residues) is VYAEPQAQNIPWVK.

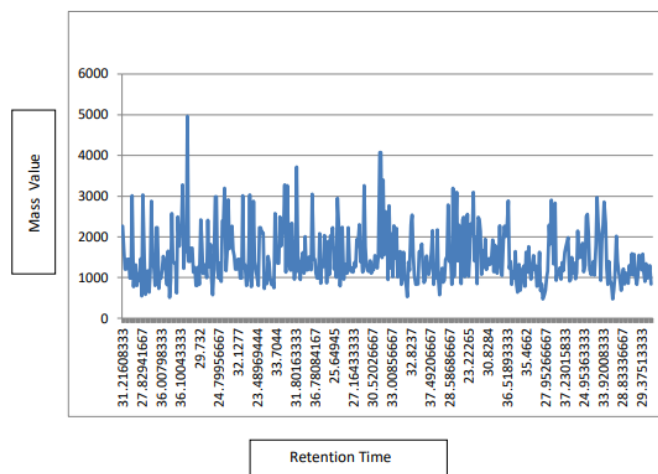


Figure – 5 MALDI-TOF spectrum of bacteriocin

### Bio assays of bacteriocin

**Antifungal efficacy:** Thus, the isolate *Lactobacillus delbrueckii*'s wide spectrum antimicrobial effect against common pathogenic bacteria suggested that this isolate could be useful and could be used to isolate and purify bacteriocin or bacteriocin-like inhibitory molecules. The isolated



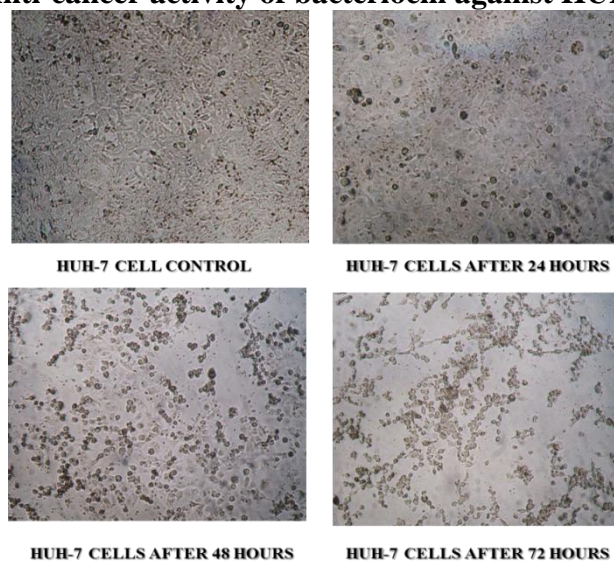
strain and eventually purified bacteriocin showed significant reticent effects on *Candida albicans* in the current research. Highest antifungal efficacy was observed in *Lactobacillus delbrueckii*. This showed that bacteriocin derived from *Lactobacillus delbrueckii* had a wide spectrum of action against *Candida albicans*. *Lactobacillus delbrueckii* metabolites had inhibitory zones ranging from 16 to 22 mm. Inhibitory zones of 18 and 17 mm, respectively were recorded in *Aspergillus niger* culture when indicted against bacteriocin but only 12 mm zone was observed against the same fungus with fluconazole anti-fungal after the incubation of 72 h.



**Figure -6 Antifungal activity – Growth inhibition**

**Anti-cancer potential of bacteriocin peptides:** Five various human cancer cells, including MCF-7, HeLa, HT1080, HEK293T and H1299 were used to assess the anticancer efficacy of bacteriocin LACDEL-14.3. The cell lines were incubated with higher concentrations of the bacteriocin LACDEL-14.3 in the specific growth medium which shows that LACDEL-14.3 has a natural cytotoxic effect against whole cancer cell lines studied. Whole cancer cell lines displayed cytotoxicity which depends on the bacteriocin concentration. The highest activity was observed against MCF-7 cells at a concentration of 10  $\mu\text{M}$ . At a 10  $\mu\text{M}$  concentration LS10, MCF-7 showed 60% cytotoxicity, while HT1080, HEK293T, and H1299 exhibited just 40% cytotoxicity. However, at a concentration of 10  $\mu\text{M}$ , there was no significant cytotoxicity in HeLa cells, but at a concentration of 100  $\mu\text{M}$ , there was 50% cytotoxicity in HeLa and other cell lines.

**Figure – 7 Anti-cancer activity of bacteriocin against HUH -7 cell lines**



**Figure – 8 Anti-cancer activity of bacteriocin against HUH -7 cell lines morphology**

**Conclusion:** These findings help *Lactobacillus delbrueckii*'s probiotic efficacy in vaginal application and further pathologies. In fact, our current study and findings are the first to identify a new bacteriocin which was produced by *Lactobacillus delbrueckii* which molecular weight was predicted as 13 kDa and antimicrobial efficacy against vaginal pathogenic microorganisms, specifically *Candida* and the most common pathogens associated with vaginal infections. These findings support the use of this probiotic strain in clinical practice for the treatment and prevention of infections by establishing a novel mechanism of action. More research into bacteriocin synthesis, isolation, and purification, as well as its possible mechanism of action, is needed.

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**CONFLICT OF INTEREST:** The authors declare no conflict of interest.

**REFERENCES:**

- [1] Maldonado-Barragán A, West SA. The cost and benefit of quorum sensing-controlled bacteriocin production in *Lactobacillus plantarum*. *Journal of Evolutionary Biology*. 2020 Jan;33(1):101-11.
- [2] Goel A, Halami PM, Tamang JP. Genome Analysis of *Lactobacillus plantarum* isolated From Some Indian Fermented Foods for Bacteriocin Production and Probiotic Marker Genes. *Frontiers in microbiology*. 2020 Jan 29;11:40.
- [3] Kheirollahi F, Anvari M. Bacteriocin production by *Lactobacillus* spp. And study on it's antimicrobial activity against some pathogenic microorganisms. *Majallah-ipizishki-iDanishgah-iUlum-iPizishkivaKhadamat-iBihdashti-iDarmani-i Tabriz*. 2019 Aug 1;41(3):53-60.
- [4] Zhang J, Yang Y, Yang H, Bu Y, Yi H, Zhang L, Han X, Ai L. Purification and partial characterization of bacteriocin Lac-B23, a novel bacteriocin production by *Lactobacillus plantarum* J23, isolated from Chinese traditional fermented milk. *Frontiers in microbiology*. 2018 Oct 1;9:2165.
- [5] Sabo SS, Converti A, Ichiwaki S, Oliveira RP. Bacteriocin production by *Lactobacillus plantarum* ST16Pa in supplemented whey powder formulations. *Journal of dairy science*. 2019 Jan 1;102(1):87-99.
- [6] Mohammadi F, Eshaghi M, Razavi S, Sarokhalil DD, Talebi M, Pourshafie MR. Characterization of bacteriocin production in *Lactobacillus* spp. isolated from mother's milk. *Microbial pathogenesis*. 2018 May 1;118:242-6.
- [7] Maldonado-Barragán A, West SA. The cost and benefit of quorum sensing-controlled bacteriocin production in *Lactobacillus plantarum*. *Journal of Evolutionary Biology*. 2020 Jan;33(1):101-11.
- [8] Turgis M, Vu KD, Millette M, Dupont C, Lacroix M. Influence of environmental factors on bacteriocin production by human isolates of *Lactococcus lactis* MM19 and *Pediococcus acidilactici* MM33. *Probiotics and antimicrobial proteins*. 2016 Mar 1;8(1):53-9.
- [9] Gordon DM, O'Brien CL. Bacteriocin diversity and the frequency of multiple bacteriocin production in *Escherichia coli*. *Microbiology*. 2006 Nov 1;152(11):3239-

44.

- [10] Lakshminarayanan B, Guinane CM, O'Connor PM, Coakley M, Hill C, Stanton C, O'Toole PW, Ross RP. Isolation and characterization of bacteriocin-producing bacteria from the intestinal microbiota of elderly Irish subjects. *Journal of applied microbiology*. 2013 Mar;114(3):886-98.
- [11] Corr SC, Li Y, Riedel CU, O'Toole PW, Hill C, Gahan CG. Bacteriocin production as a mechanism for the antiinfective activity of *Lactobacillus salivarius* UCC118. *Proceedings of the National Academy of Sciences*. 2007 May 1;104(18):7617-21.
- [12] Birri DJ, Brede DA, Tessema GT, Nes IF. Bacteriocin production, antibiotic susceptibility and prevalence of haemolytic and gelatinase activity in faecal lactic acid bacteria isolated from healthy Ethiopian infants. *Microbial ecology*. 2013 Feb 1;65(2):504-16.