

Molecular Mechanism of Insulin Resistance and Type 2 Diabetes

Nirubama.K

Saveetha Dental College and Hospitals

Saveetha Institute of Medical and Technical Sciences (SIMATS)

Saveetha University

Chennai- 600077,

India.

Email Id- 151801045.sdc@saveetha.com

Dr.Raghu Nandhakumar

Associate Professor

Department of Pharmacology

Saveetha Dental College and Hospitals

Saveetha Institute of Medical and Technical Sciences (SIMATS)

Saveetha University

Chennai-600077,

India.

Email Id- ragunandhakumars.sdc@saveetha.com

Dr.Ezhilarasan,

Associate Professor,

Department of Pharmacology

Saveetha Dental College and Hospitals,

Saveetha Institute of Medical and Technical Sciences (SIMATS)

Saveetha University

Chennai-600077,

India.

Email Id- ezhilarasand.sdc@saveetha.com

Dr.Lakshmi Thangavelu

Associate Professor

Department of Pharmacology

Saveetha Dental College and Hospitals

Saveetha Institute of Medical and Technical Sciences (SIMATS)

Saveetha University

Chennai-600077,

India.

Email Id- lakshmi@saveetha.com

Corresponding Author
Dr.Raghu Nandhakumar,
Associate Professor,
Department of Pharmacology,
Saveetha Dental College and Hospitals,
Saveetha Institute of Medical and Technical Sciences (SIMATS)
Saveetha University
Chennai-600077,
India.
Email Id- ragunandhakumars.sdc@saveetha.com
Phone: 9790270479

ABSTRACT:

Aim: To analyze the relationship between the molecular mechanism of insulin resistance and its associated type-2 diabetes.

Method: Relevant articles were identified via PUBMED, GOOGLE SCHOLAR MeSH, Cochrane, bioRxiv, Semantic scholar search engines using the keywords Insulin receptor, Insulin resistance, signaling, Insulin receptor substrate proteins, Type 2 diabetes mellitus. Period of duration considered was from 1980 to till date.

Discussion: Insulin resistance is a major risk factor for developing type 2 diabetes caused by insulin-target tissue's inability to respond properly to insulin and contributes to obesity morbidity. Insulin action involves a series of signaling cascades initiated by insulin binding to its receptor, causing autophosphorylation of the receptor and activation of the tyrosine kinase receptor, resulting in tyrosine phosphorylation of the insulin receptor substrates (IRSs). Phosphorylation of IRSs contributes to phosphatidylinositol 3-kinase (PI3K) activation and subsequent activation of Akt and its downstream mediator AS160, both of which are critical measures to promote insulin-induced glucose transport.

Conclusion: Identifying signals and understanding the complex relationship of the various factors that modulate insulin sensitivity are important prerequisites for developing new and more specific antidiabetic compounds.. By elucidating the cellular and molecular mechanisms responsible for insulin resistance, these studies provide potential new targets to treat and prevent type 2 diabetes mellitus. This review focuses on the molecular basis of insulin resistance to skeletal muscles and type 2 diabetes.

keywords: Insulin receptor, Insulin resistance, signaling, Insulin receptor substrate proteins, Type 2 diabetes mellitus

INTRODUCTION:

Type 2 DM soon appears as one of 21st century's biggest public health threats. The pathogenesis of Type 2 diabetes includes irregularities in the activity of insulin as well as secretion.(Wild *et al.*, 2004). The relationship between insulin resistance and type-2 diabetes is not only the most

powerful predictor of future development of type-2 diabetes, it is also a therapeutic target once hyperglycemia is present. The natural activity of insulin is begun by establishing its layer receptor that triggers various flagging processes for intercession. Because of the importance of metabolic guidelines and advancing elements of cell development and expansion, insulin activity is exceptionally controlled in order to advance the appropriate balance of metabolic work and vitality. This can prompt a condition known as insulin resistance, which is the result of insufficient insulin flagging caused by transformations or post translational alterations of receptor or effector atoms found downstream. Translocation of glucose transporter 4 (Glut4) the main insulin controlled glucose transportation network, from intracellular vesicles to the plasma membrane and transverse tubules, facilitates the capacity of Insulin to improve glucose transfer in skeletal muscles (2). On a cellular level, it defines the inadequate strength of insulin signaling from the insulin receptor downstream to the final substrates of insulin action involved in various metabolic and mitogenic aspects of cellular function (Kraemer and Ginsberg, 2014). The type 2 pathogenesis involves anomalies in both the action of insulin and its secretion (Saltiel, 2001). Although the precise pathophysiological sequence leading to insulin resistance is still largely unknown, recent studies have contributed to a deeper understanding of the molecular mechanisms. Also expected to trigger a steep rise in complications associated with diabetes, such as ischemic heart disease, stroke, neuropathy, retinopathy, and nephropathy. Besides β cell failure, the major pathophysiological event which is contributing to the development of type 2 DM is target tissues resistance to insulin, which is usually associated with abnormal insulin secretion. necessitates the need to develop new drugs for its effective management. Plants and their bioactive compounds are found to be an alternative therapeutic approach. The *C. fimbriata* extract showed potent inhibitory activity on enzymes of glucose metabolism in a dose-dependent manner (Anitha and Ashwini, 2017; Ashwini, Ezhilarasan and Anitha, 2017). This review discusses the molecular mechanisms of insulin resistance in type 2 diabetes. *Azadirachta indica* is an evergreen tree having potential medicinal values and it has anti mutagenic values (Lakshmi *et al.*, 2015). A detailed understanding of these basic pathophysiological mechanisms is critical to developing novel therapeutic strategies for diabetes. Our team has rich experience in research and we have collaborated with numerous authors over various topics in the past decade (Ariga *et al.*, 2018; Basha, Ganapathy and Venugopalan, 2018; Hannah *et al.*, 2018; Hussainy *et al.*, 2018; Jeevanandan and Govindaraju, 2018; Kannan and Venugopalan, 2018; Kumar and Antony, 2018; Manohar and Sharma, 2018; Menon, Ks, R, *et al.*, 2018; Nandakumar and Nasim, 2018; Nandhini, Babu and Mohanraj, 2018; Ravinthar and Jayalakshmi, 2018; Seppan *et al.*, 2018; Teja, Ramesh and Priya, 2018; Duraisamy *et al.*, 2019; Gheena and Ezhilarasan, 2019a; Hema Shree *et al.*, 2019; Rajakeerthi and Ms, 2019; Rajendran *et al.*, 2019; Sekar *et al.*, 2019; Sharma *et al.*, 2019; Siddique *et al.*, 2019; Janani, Palanivelu and Sandhya, 2020; Johnson *et al.*, 2020; Jose, Ajitha and Subbaiyan, 2020).

RETRIEVAL OF LITERATURE DATA:

Relevant articles were referred by PUBMED, GOOGLE SCHOLAR MeSH, Cochrane, bioRxiv, Semantic scholar search engines using the keywords Insulin receptor, Insulin resistance, Insulin signaling, Insulin receptor substrate proteins and type 2 diabetes mellitus. Period of duration considered was from 1980 to till date. Out of searched article, 28 articles were selected, out of which 25 are with known concept and 3 articles with recent concept. Our institution is passionate about high quality evidence based research and has excelled in various fields (Pc, Marimuthu

and Devadoss, 2018; Ramesh *et al.*, 2018; Vijayashree Priyadharsini, Smiline Girija and Paramasivam, 2018; Ezhilarasan, Apoorva and Ashok Vardhan, 2019; Ramadurai *et al.*, 2019; Sridharan *et al.*, 2019; Vijayashree Priyadharsini, 2019; Chandrasekar *et al.*, 2020; Mathew *et al.*, 2020; R *et al.*, 2020; Samuel, 2021)

Insulin signaling:

Insulin receptor (IR) is a heterotetramer consisting of two α subunits and two β subunits linked by disulphide bonds (Pedersen *et al.*, 1990). Insulin binds to the insulin receptor α subunit and activates the tyrosine kinase in the β subunit. The activation of tyrosine kinase in the insulin receptor promotes autophosphorylation of the β subunit, where phosphorylation of three tyrosine residues (Tyr-1158, Tyr-1162, and Tyr-1163) is needed to amplify kinase activity (White and Ronald Kahn, 1989). Most of insulin metabolism and antiapoptotic effects are mediated by the signaling pathway of phosphorylation of the insulin receptor substrate (IRS) proteins, and the activation of the phosphatidylinositol (PI) 3-kinase, Akt (also known as protein kinase B), the molecular target of rapamycin (mTOR), and p70 S6 kinase (Guesdon, Waller and Saklatvala, 1994). The insulin receptor tyrosine kinase phosphorylates the IRS proteins, and phosphotyrosine residues on IRS proteins become good targets for the p85 regulatory subunit of PI3-kinase (Shepherd, Navé and Siddle, 1995). The activated PI3-kinase generates 3'-phosphoinositides [phosphatidyl - inositol-3,4-bisphosphate (PIP2) and phosphatidyl-inositol-3,4,5-trisphosphate (PIP3)] (Alessi, 2000) which are bound to the phosphoinositide dependent kinase 1 (PDK1) (Balusamy *et al.*, 2018). The PDKs are the protein kinase B (PKB) and also atypical forms of the protein kinase C (PKC) (Kotani *et al.*, 1998).

Mutation in irs proteins:

Insulin initiates its diverse biological effects by binding to the α -subunit of the insulin receptor, resulting in activation of intrinsic tyrosine kinase activity and rapid phosphorylation of tyrosine residues in three domains, including the juxtamembrane domain, the tyrosine kinase domain, and the COOH-terminal domain of its intracellular β -subunit. Insulin receptor kinase-1 is one of the major substrates of insulin receptor tyrosine kinase and mediates various insulin signals downstream. After insulin stimulation, the insulin receptor undergoes autophosphorylation on tyrosine residues, which in turn activates the tyrosine kinase activity of the receptor towards other substrates. IRS-1 played a significant role in insulin-stimulated glucose uptake, suggesting that the IRS-1 gene may be involved in the development of NIDDM. In humans, insulin resistance is associated with rare mutations of the IRS-1 protein (Whitehead *et al.*, 1998). Disruption of the IRS-1 gene in mice results in insulin resistance, mainly of muscle and fat (Yamauchi *et al.*, 1996). The analysis of IRs in knockout mice shows an important phenotype of single allele IRS-1 gene whereas homozygous disruption of the IRS-1 gene results in a mild form of insulin resistance (Erenler and Karan, 2017). IRS-1 homozygous null mice (IRS-1^{-/-}) do not show a clear diabetic phenotypic expression, presumably because of pancreatic β cell compensation. By comparison, IRS-2^{-/-} mice developed diabetes as a result of severe insulin resistance paired with β cell failure (Araki *et al.*, 1994).

Inhibition of IR gene transcription:

The promoter region of the human IR gene has been established and researched by many groups in order to explain the molecular basis for the modulation of IR expression. Two unique AT-rich sequences, C2 and E3, within the IR gene promoter and both sequences are positively regulated by transcription factor HMGA1 (earlier known as HMG1-Y) (23). HMGA1 interacts with the AT rich regions and regulates multiple gene transcriptional activation by modifying DNA conformation that allows transcriptional factor recruitment to the transcription start site (Bustin and Reeves, 1996). HMGA1 induces human IR gene transcriptional activation by enabling the recruitment of SP1 and cEBP β , the ubiquitously expressed transcription factors, to the promoter region. A recent report demonstrates that a genetic defect that reduces the intracellular expression of HMGA1 protein may adversely affect IR expression in cells and tissues from subjects with insulin resistance and type 2 diabetes (Foti *et al.*, 2005). There is also a possibility that activated PKC ϵ phosphorylates HMGA1, which inhibits its mobilization to the promoter region of IR gene (Sujatha, Asokan and Rajeshkumar, 2018). It has been shown that phosphorylation of the HMGA1 protein reduces its DNA-binding ability (Reeves and Beckerbauer, 2001). There is no specific transcription factor recruitment for the promoter region of the IR gene and therefore no expression of the IR gene without the mobilization of HMGA1 for the IR promoter.

Mitochondrial dysfunction:

For many years, severe mitochondrial dysfunction can result in diabetes (DiMauro and Rustin, 2009). The observation of abnormal mitochondrial function in vitro in type 2 diabetes (Kelley *et al.*, 2002) was soon followed by in vivo demonstration of this abnormality in insulin-resistant, first-degree relatives of people with type 2 diabetes (Petersen *et al.*, 2004). Further reports of a modest defect in muscle mitochondrial function in type 2 diabetes were published shortly thereafter (Szendroedi *et al.*, 2007; Phielix *et al.*, 2008). These studies raised the question of whether type 2 diabetes could be a primary disorder of the mitochondria. However, the study of first-degree relatives tended to be misinterpreted as having shown a major defect in mitochondrial function in type 2 diabetes, although it had studied nondiabetic groups from the opposite ends of the insulin resistance–sensitivity spectrum. Indeed, other studies showed no defect in mitochondrial function in type 2 diabetes (De Feyter *et al.*, 2008; Ee L. Lim *et al.*, 2011a), which led to further confusion. Mitochondrial function was then shown to be acutely modifiable by changing fatty acid availability (Baker *et al.*, 2006; Ee L. Lim *et al.*, 2011b) and that it was affected by ambient blood glucose concentration (Kelley and Mandarino, 1990). When ambient blood glucose levels were near normal in diabetes, no defect in mitochondrial function was apparent. In a study using $^{13}\text{C}/^{31}\text{P}$ MRS, it was found that in healthy lean elderly volunteers with severe muscle insulin resistance, there is ~40% reduction in the rates of oxidative phosphorylation activity associated with increased intramyocellular and intrahepatic lipid content (Lee, Sciamanna and Peterson, 1993). This study indicates that the acquired lack of mitochondrial function associated with aging predisposes elderly subjects to intramyocellular lipid accumulation, which results in insulin resistance. Furthermore, mitochondrial density reduced by 38%, intramyocellular lipid content increased by 60% and serine phosphorylation of

IRS-1 was increased by 50% in the young insulin-resistant offspring of type 2 diabetes. Muscle insulin resistance as determined by the euglycemic-hyperinsulinemic clamp is clearly a risk factor for development of type 2 diabetes (Lillioja *et al.*, 1993). However, the pathophysiology of hyperglycemia in established diabetes relates to hepatic not muscle insulin resistance. This distinction has been elegantly demonstrated in studies of moderate calorie restriction in type 2 diabetes, which resulted in a fall in liver fat, normalization of hepatic insulin sensitivity, and fasting plasma glucose, but no change in muscle insulin resistance (Petersen *et al.*, 2005). More recent work employing severe calorie restriction confirmed previous findings and also demonstrated a longer-term return of normal insulin secretion as intrapancreatic fat content fell (E. L. Lim *et al.*, 2011).

Adipokines:

Insulin has three major target tissues-skeletal muscle, adipose tissue and the liver. Not only is IR overexpressed in these tissues, but they also have three sites where glucose is accumulated and stored, no other tissue may store glucose. About 75% of postprandial insulin-dependent glucose disposal takes place in the skeletal muscle (Klip *et al.*, 1990). Hence, it is a main target organ. Insulin resistance patients with type 2 diabetes frequently display signs of abnormal lipid metabolism, increased circulatory concentration and elevated deposition of lipids in the skeletal muscle. Increase in plasma FFA reduces insulin-stimulated glucose uptake, whereas a decrease in plasma lipid content improves insulin activity in the skeletal muscle cells, adipocytes and liver. Studies have shown that increasing plasma fatty acids in both rodents and humans abolishes insulin activation of IRS-1-associated PI3-kinase activity in skeletal muscle were most prevalent in IRS-1 gene. . Insulin resistance associated with lipid metabolism - has also been shown to be linked to GLUT4 translocation defects. Adipose tissue also acts as an endocrine organ producing adipokines which modulate glucose homeostasis. Many studies are most intensely discussed about the TNF- α , leptin, adiponectin and resistin at the molecular level of insulin resistance and type 2 diabetes mellitus. Recently a study reported that TNF- α increases serine phosphorylation of IRS-1 and down-regulates GLUT4 expression, thereby activating the insulin resistance. Furthermore, mice without functional TNF- α were protected from obesity-induced insulin resistance (Uysal *et al.*, 1997). The role of leptin in regulating food intake and energy expenditure is well known. Humans with leptin deficiency or leptin receptor mutations are severely obese. It also has direct effects on insulin sensitivity and may also reverse insulin resistance in mice with congenital lipodystrophy. Adiponectin has insulin-sensitizing effects, as it facilitates suppression of the synthesis of hepatic glucose and utilization in muscle and fat. The expression of adiponectin is decreased in obese humans and mice (Stumvoll and Häring, 2002). Adiponectin levels were correlated with insulin sensitivity in humans. Because of its insulin-antagonistic effects, adipokine resistance has drawn considerable attention in preclinical research. Resistin decreases in vitro insulin-dependent glucose transport and improves in vivo high blood glucose levels and hepatic glucose production.

Type-2 diabetes association with cancer and its treatment:

Diabetes and cancer are common diseases with tremendous impact on health worldwide. Epidemiologic evidence suggests that people with diabetes are at significantly higher risk for

many forms of cancer. Type 2 diabetes and cancer share many risk factors (Giovannucci *et al.*, 2010). Diabetes (primarily type 2) is associated with increased risk for some cancers (liver, pancreas, endometrium, colon and rectum, breast, bladder). Diabetes is associated with reduced risk of prostate cancer. The association between diabetes and some cancers may partly be due to shared risk factors between the two diseases, such as aging, obesity, diet, and physical inactivity. Possible mechanisms for a direct link between diabetes and cancer include hyperinsulinemia, hyperglycemia, and inflammation. Accordingly, new strategies are evolving to control and to treat cancer and one such strategy could be the use of medicinal plants. Previous studies have reported the anticancer efficacy of medicinal plants against several human in vitro cancer cell lines and came out with promising results (Ezhilarasan, Lakshmi, Vijayaragavan, *et al.*, 2017). Coumarin plays a vital role in the drug discovery process against type-2 diabetes due to its diverse biologically active components (Perumalsamy *et al.*, 2018). The ethanol seed extracts of *A. catechu* were found to be cytotoxic at lower concentrations and induced apoptosis in human pancreatic cells (Ezhilarasan, Lakshmi, Nagaich, *et al.*, 2017). Nanotechnology beholds infinite potential and innovative applications which are being continuously explored for detecting, diagnosing, imaging and treating different types of cancers (Rajeshkumar, Venkat Kumar, *et al.*, 2018; Sharma *et al.*, 2019). Nanoparticles play an important role in the target-specific delivery of drugs. In addition, oligonucleotides also are extensively used for gene transfer in the form of polymeric, liposomal and inorganic carrier materials (Rajeshkumar, Agarwal, *et al.*, 2018; Mehta *et al.*, 2019). Selenium nanoparticles can open ways to new regular strategies for treating illnesses like malignancy, and this audit expresses the reasons why these nano measured medications can be the following huge achievement as chemotherapeutic operators (Karthiga, Rajeshkumar and Annadurai, 2018; Menon, Ks, Santhiya, *et al.*, 2018). There is a significant relationship between oxidative stress and different liver pathogenesis induced by drugs and xenobiotics, focusing upon different chronic liver injury induced by alcohol, antitubercular drugs and hyperactivity of drugs in hyperglycemic patients (Ezhilarasan, 2018; Ezhilarasan, Sokal and Najimi, 2018; Gheena and Ezhilarasan, 2019b). Our institution is passionate about high quality evidence based research and has excelled in various fields ((Pc, Marimuthu and Devadoss, 2018; Ramesh *et al.*, 2018; Vijayashree Priyadharsini, Smiline Girija and Paramasivam, 2018; Ezhilarasan, Apoorva and Ashok Vardhan, 2019; Ramadurai *et al.*, 2019; Sridharan *et al.*, 2019; Vijayashree Priyadharsini, 2019; Chandrasekar *et al.*, 2020; Mathew *et al.*, 2020; R *et al.*, 2020; Samuel, 2021)

CONCLUSION:

In this review, we have summarized the recent findings that leads to our understanding of insulin resistance type 2 diabetes pathogenesis . Identifying signals and understanding the complex relationship of the various factors that modulate insulin sensitivity are important prerequisites for developing new and more specific antidiabetic compounds. About 75% of postprandial insulin-dependent glucose disposal takes place in the skeletal muscle. By elucidating the cellular and molecular mechanisms responsible for insulin resistance, these studies provide potential new targets to treat and prevent type 2 diabetes mellitus.

REFERENCE:

1. Alessi, D. (2000) 'Mechanism and activation and function of protein kinase B',

Toxicology, p. 8. doi: 10.1016/s0300-483x(00)90243-9.

2. Anitha, R. and Ashwini, S. (2017) 'Antihyperglycemic activity of *Caralluma fimbriata*: An In vitro approach', *Pharmacognosy Magazine*, p. 499. doi: 10.4103/pm.pm_59_17.
3. Araki, E. *et al.* (1994) 'Alternative pathway of insulin signalling in mice with targeted disruption of the IRS-1 gene', *Nature*, 372(6502), pp. 186–190.
4. Ariga, P. *et al.* (2018) 'Determination of correlation of width of Maxillary Anterior Teeth using Extraoral and Intraoral Factors in Indian Population: A systematic review', *World journal of dentistry*, 9(1), pp. 68–75.
5. Ashwini, S., Ezhilarasan, D. and Anitha, R. (2017) 'Cytotoxic Effect of *Caralluma fimbriata* Against Human Colon Cancer Cells', *Pharmacognosy Journal*, pp. 204–207. doi: 10.5530/pj.2017.2.34.
6. Baker, D. J. *et al.* (2006) 'The Experimental Type 2 Diabetes Therapy Glycogen Phosphorylase Inhibition Can Impair Aerobic Muscle Function During Prolonged Contraction', *Diabetes*, pp. 1855–1861. doi: 10.2337/db05-1687.
7. Balusamy, S. R. *et al.* (2018) 'Anti-proliferative activity of *Origanum vulgare* inhibited lipogenesis and induced mitochondrial mediated apoptosis in human stomach cancer cell lines', *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*, 108, pp. 1835–1844.
8. Basha, F. Y. S., Ganapathy, D. and Venugopalan, S. (2018) 'Oral hygiene status among pregnant women', *Journal of advanced pharmaceutical technology & research*, 11(7), p. 3099.
9. Bustin, M. and Reeves, R. (1996) 'High-mobility-group chromosomal proteins: architectural components that facilitate chromatin function', *Progress in nucleic acid research and molecular biology*, 54, pp. 35–100.
10. Chandrasekar, R. *et al.* (2020) 'Development and validation of a formula for objective assessment of cervical vertebral bone age', *Progress in orthodontics*, 21(1), p. 38.
11. De Feyter, H. M. *et al.* (2008) 'Early or advanced stage type 2 diabetes is not accompanied by in vivo skeletal muscle mitochondrial dysfunction', *European journal of endocrinology / European Federation of Endocrine Societies*, 158(5), pp. 643–653.
12. DiMauro, S. and Rustin, P. (2009) 'A critical approach to the therapy of mitochondrial respiratory chain and oxidative phosphorylation diseases', *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, pp. 1159–1167. doi: 10.1016/j.bbadis.2008.10.015.
13. Duraisamy, R. *et al.* (2019) 'Compatibility of Nonoriginal Abutments With Implants: Evaluation of Microgap at the Implant-Abutment Interface, With Original and Nonoriginal Abutments', *Implant dentistry*, 28(3), pp.

289–295.

14. Erenler, R. and Karan, T. (2017) 'Screening of norharmane from seven cyanobacteria by high-performance liquid chromatography', *Pharmacognosy Magazine*, p. 723. doi: 10.4103/pm.pm_214_17.
15. Ezhilarasan, D., Lakshmi, T., Vijayaragavan, R., *et al.* (2017) 'Acacia catechu ethanolic bark extract induces apoptosis in human oral squamous carcinoma cells', *Journal of Advanced Pharmaceutical Technology & Research*, p. 143. doi: 10.4103/japtr.japtr_73_17.
16. Ezhilarasan, D., Lakshmi, T., Nagaich, U., *et al.* (2017) 'Acacia catechu ethanolic seed extract triggers apoptosis of SCC-25 cells', *Pharmacognosy Magazine*, p. 405. doi: 10.4103/pm.pm_458_16.
17. Ezhilarasan, D. (2018) 'Oxidative stress is bane in chronic liver diseases: Clinical and experimental perspective', *Arab Journal of Gastroenterology*, pp. 56–64. doi: 10.1016/j.ajg.2018.03.002.
18. Ezhilarasan, D., Apoorva, V. S. and Ashok Vardhan, N. (2019) 'Syzygium cumini extract induced reactive oxygen species-mediated apoptosis in human oral squamous carcinoma cells', *Journal of oral pathology & medicine: official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*, 48(2), pp. 115–121.
19. Ezhilarasan, D., Sokal, E. and Najimi, M. (2018) 'Hepatic fibrosis: It is time to go with hepatic stellate cell-specific therapeutic targets', *Hepatobiliary & Pancreatic Diseases International*, pp. 192–197. doi: 10.1016/j.hbpd.2018.04.003.
20. Foti, D. *et al.* (2005) 'Lack of the architectural factor HMGA1 causes insulin resistance and diabetes in humans and mice', *Nature medicine*, 11(7), pp. 765–773.
21. Gheena, S. and Ezhilarasan, D. (2019a) 'Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells', *Human & experimental toxicology*, 38(6), pp. 694–702.
22. Gheena, S. and Ezhilarasan, D. (2019b) 'Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells', *Human & Experimental Toxicology*, pp. 694–702. doi: 10.1177/0960327119839173.
23. Giovannucci, E. *et al.* (2010) 'Diabetes and Cancer: A consensus report', *Diabetes Care*, pp. 1674–1685. doi: 10.2337/dc10-0666.
24. Guesdon, F., Waller, R. J. and Saklatvala, J. (1994) 'β Casein kinase: Exclusive activation by IL-1 or TNF and substrate specificity', *Cytokine*, p. 558. doi: 10.1016/1043-4666(94)90200-3.

25. Hannah, R. *et al.* (2018) 'Awareness about the use, ethics and scope of dental photography among undergraduate dental students dentist behind the lens', *Journal of advanced pharmaceutical technology & research*, 11(3), p. 1012.
26. Hema Shree, K. *et al.* (2019) 'Saliva as a Diagnostic Tool in Oral Squamous Cell Carcinoma - a Systematic Review with Meta Analysis', *Pathology oncology research: POR*, 25(2), pp. 447–453.
27. Hussainy, S. N. *et al.* (2018) 'Clinical performance of resin-modified glass ionomer cement, flowable composite, and polyacid-modified resin composite in noncarious cervical lesions: One-year follow-up', *Journal of conservative dentistry: JCD*, 21(5), pp. 510–515.
28. Janani, K., Palanivelu, A. and Sandhya, R. (2020) 'Diagnostic accuracy of dental pulse oximeter with customized sensor holder, thermal test and electric pulp test for the evaluation of pulp vitality: an in vivo study', *Brazilian dental science*, 23(1). doi: 10.14295/bds.2020.v23i1.1805.
29. Jeevanandan, G. and Govindaraju, L. (2018) 'Clinical comparison of Kedo-S paediatric rotary files vs manual instrumentation for root canal preparation in primary molars: a double blinded randomised clinical trial', *European archives of paediatric dentistry: official journal of the European Academy of Paediatric Dentistry*, 19(4), pp. 273–278.
30. Johnson, J. *et al.* (2020) 'Computational identification of MiRNA-7110 from pulmonary arterial hypertension (PAH) ESTs: a new microRNA that links diabetes and PAH', *Hypertension research: official journal of the Japanese Society of Hypertension*, 43(4), pp. 360–362.
31. Jose, J., Ajitha and Subbaiyan, H. (2020) 'Different treatment modalities followed by dental practitioners for Ellis class 2 fracture – A questionnaire-based survey', *The open dentistry journal*, 14(1), pp. 59–65.
32. Kannan, A. and Venugopalan, S. (2018) 'A systematic review on the effect of use of impregnated retraction cords on gingiva', *Journal of advanced pharmaceutical technology & research*, 11(5), p. 2121.
33. Karthiga, P., Rajeshkumar, S. and Annadurai, G. (2018) 'Mechanism of Larvicidal Activity of Antimicrobial Silver Nanoparticles Synthesized Using Garcinia mangostana Bark Extract', *Journal of Cluster Science*, pp. 1233–1241. doi: 10.1007/s10876-018-1441-z.
34. Kelley, D. E. *et al.* (2002) 'Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes', *Diabetes*, 51(10), pp. 2944–2950.
35. Kelley, D. E. and Mandarino, L. J. (1990) 'Hyperglycemia normalizes insulin-stimulated skeletal muscle glucose oxidation and storage in noninsulin-dependent diabetes mellitus', *Journal of Clinical Investigation*, pp. 1999–2007. doi: 1112

10.1172/jci114935.

36. Klip, A. *et al.* (1990) 'Recruitment of GLUT-4 glucose transporters by insulin in diabetic rat skeletal muscle', *Biochemical and biophysical research communications*, 172(2), pp. 728–736.
37. Kotani, K. *et al.* (1998) 'Requirement of atypical protein kinase clambda for insulin stimulation of glucose uptake but not for Akt activation in 3T3-L1 adipocytes', *Molecular and cellular biology*, 18(12), pp. 6971–6982.
38. Kraemer, F. B. and Ginsberg, H. N. (2014) 'Gerald M. Reaven, MD: Demonstration of the central role of insulin resistance in type 2 diabetes and cardiovascular disease', *Diabetes care*, 37(5), pp. 1178–1181.
39. Kumar, D. and Antony, S. D. P. (2018) 'Calcified canal and negotiation-A review', *Journal of advanced pharmaceutical technology & research*, 11(8), p. 3727.
40. Lakshmi, T. *et al.* (2015) 'Azadirachta indica: A herbal panacea in dentistry - An update', *Pharmacognosy reviews*, 9(17), pp. 41–44.
41. Lee, C. P., Sciamanna, M. and Peterson, P. L. (1993) 'Intact Rat Brain Mitochondria from a Single Animal: Preparation and Properties', *Mitochondrial Dysfunction*, pp. 41–50. doi: 10.1016/b978-0-12-461205-1.50010-2.
42. Lillioja, S. *et al.* (1993) 'Insulin Resistance and Insulin Secretory Dysfunction as Precursors of Non-Insulin-Dependent Diabetes Mellitus: Prospective Studies of Pima Indians', *New England Journal of Medicine*, pp. 1988–1992. doi: 10.1056/nejm199312303292703.
43. Lim, E. L. *et al.* (2011a) 'Effects of raising muscle glycogen synthesis rate on skeletal muscle ATP turnover rate in type 2 diabetes', *American journal of physiology. Endocrinology and metabolism*, 301(6), pp. E1155–62.
44. Lim, E. L. *et al.* (2011b) 'Inhibition of lipolysis in Type 2 diabetes normalizes glucose disposal without change in muscle glycogen synthesis rates', *Clinical science*, 121(4), pp. 169–177.
45. Lim, E. L. *et al.* (2011) 'Reversal of type 2 diabetes: normalisation of beta cell function in association with decreased pancreas and liver triacylglycerol', *Diabetologia*, 54(10), pp. 2506–2514.
46. Manohar, M. P. and Sharma, S. (2018) 'A survey of the knowledge, attitude, and awareness about the principal choice of intracanal medicaments among the general dental practitioners and nonendodontic specialists', *Indian journal of dental research: official publication of Indian Society for Dental Research*, 29(6), pp. 716–720.
47. Mathew, M. G. *et al.* (2020) 'Evaluation of adhesion of Streptococcus mutans, plaque

accumulation on zirconia and stainless steel crowns, and surrounding gingival inflammation in primary molars: Randomized controlled trial', *Clinical oral investigations*, pp. 1–6.

48. Mehta, M. *et al.* (2019) 'Oligonucleotide therapy: An emerging focus area for drug delivery in chronic inflammatory respiratory diseases', *Chemico-biological interactions*, 308, pp. 206–215.
49. Menon, S., Ks, S. D., R, S., *et al.* (2018) 'Selenium nanoparticles: A potent chemotherapeutic agent and an elucidation of its mechanism', *Colloids and surfaces. B, Biointerfaces*, 170, pp. 280–292.
50. Menon, S., Ks, S. D., Santhiya, R., *et al.* (2018) 'Selenium nanoparticles: A potent chemotherapeutic agent and an elucidation of its mechanism', *Colloids and Surfaces B: Biointerfaces*, pp. 280–292. doi: 10.1016/j.colsurfb.2018.06.006.
51. Nandakumar, M. and Nasim, I. (2018) 'Comparative evaluation of grape seed and cranberry extracts in preventing enamel erosion: An optical emission spectrometric analysis', *Journal of conservative dentistry: JCD*, 21(5), pp. 516–520.
52. Nandhini, J. S. T., Babu, K. Y. and Mohanraj, K. G. (2018) 'Size, shape, prominence and localization of gerdy's tubercle in dry human tibial bones', *Journal of advanced pharmaceutical technology & research*, 11(8), p. 3604.
53. Pc, J., Marimuthu, T. and Devadoss, P. (2018) 'Prevalence and measurement of anterior loop of the mandibular canal using CBCT: A cross sectional study', *Clinical implant dentistry and related research*. Available at: <https://europepmc.org/article/med/29624863>.
54. Pedersen, O. *et al.* (1990) 'Evidence against altered expression of GLUT1 or GLUT4 in skeletal muscle of patients with obesity or NIDDM', *Diabetes*, 39(7), pp. 865–870.
55. Perumalsamy, H. *et al.* (2018) 'In silico and in vitro analysis of coumarin derivative induced anticancer effects by undergoing intrinsic pathway mediated apoptosis in human stomach cancer', *Phytomedicine: international journal of phytotherapy and phytopharmacology*, 46, pp. 119–130.
56. Petersen, K. F. *et al.* (2004) 'Impaired Mitochondrial Activity in the Insulin-Resistant Offspring of Patients with Type 2 Diabetes', *New England Journal of Medicine*, pp. 664–671. doi: 10.1056/nejmoa031314.
57. Petersen, K. F. *et al.* (2005) 'Reversal of nonalcoholic hepatic steatosis, hepatic insulin resistance, and hyperglycemia by moderate weight reduction in patients with type 2 diabetes', *Diabetes*, 54(3), pp. 603–608.
58. Phielix, E. *et al.* (2008) 'Lower intrinsic ADP-stimulated mitochondrial respiration underlies in vivo mitochondrial dysfunction in muscle of male type 2 diabetic patients',

Diabetes, 57(11), pp. 2943–2949.

59. Rajakeerthi and Ms, N. (2019) 'Natural Product as the Storage medium for an avulsed tooth – A Systematic Review', *Cumhuriyet Üniversitesi Diş Hekimliği Fakültesi dergisi*, 22(2), pp. 249–256.
60. Rajendran, R. *et al.* (2019) 'Comparative evaluation of remineralizing potential of a paste containing bioactive glass and a topical cream containing casein phosphopeptide-amorphous calcium phosphate: An in vitro study', *Pesquisa brasileira em odontopediatria e clinica integrada*, 19(1), pp. 1–10.
61. Rajeshkumar, S., Venkat Kumar, S., *et al.* (2018) 'Biosynthesis of zinc oxide nanoparticles using *Mangifera indica* leaves and evaluation of their antioxidant and cytotoxic properties in lung cancer (A549) cells', *Enzyme and Microbial Technology*, pp. 91–95. doi: 10.1016/j.enzmictec.2018.06.009.
62. Rajeshkumar, S., Agarwal, H., *et al.* (2018) 'Brassica oleracea Mediated Synthesis of Zinc Oxide Nanoparticles and its Antibacterial Activity against Pathogenic Bacteria', *Asian Journal of Chemistry*, pp. 2711–2715. doi: 10.14233/ajchem.2018.21562.
63. Ramadurai, N. *et al.* (2019) 'Effectiveness of 2% Articaine as an anesthetic agent in children: randomized controlled trial', *Clinical oral investigations*, 23(9), pp. 3543–3550.
64. Ramesh, A. *et al.* (2018) 'Comparative estimation of sulfiredoxin levels between chronic periodontitis and healthy patients - A case-control study', *Journal of periodontology*, 89(10), pp. 1241–1248.
65. Ravinthar, K. and Jayalakshmi (2018) 'Recent advancements in laminates and veneers in dentistry', *Journal of advanced pharmaceutical technology & research*, 11(2), p. 785.
66. Reeves, R. and Beckerbauer, L. (2001) 'HMGI/Y proteins: flexible regulators of transcription and chromatin structure', *Biochimica et biophysica acta*, 1519(1-2), pp. 13–29.
67. R, H. *et al.* (2020) 'CYP2 C9 polymorphism among patients with oral squamous cell carcinoma and its role in altering the metabolism of benzo[a]pyrene', *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*, pp. 306–312. doi: 10.1016/j.oooo.2020.06.021.
68. Saltiel, A. R. (2001) 'New perspectives into the molecular pathogenesis and treatment of type 2 diabetes', *Cell*, 104(4), pp. 517–529.
69. Samuel, S. R. (2021) 'Can 5-year-olds sensibly self-report the impact of developmental enamel defects on their quality of life?', *International journal of paediatric dentistry / the British Paedodontic Society [and] the International Association of Dentistry for Children*, 31(2), pp. 285–286.

70. Sekar, D. *et al.* (2019) 'Methylation-dependent circulating microRNA 510 in preeclampsia patients', *Hypertension research: official journal of the Japanese Society of Hypertension*, 42(10), pp. 1647–1648.
71. Seppan, P. *et al.* (2018) 'Therapeutic potential of *Mucuna pruriens* (Linn.) on ageing induced damage in dorsal nerve of the penis and its implication on erectile function: an experimental study using albino rats', *The aging male: the official journal of the International Society for the Study of the Aging Male*, pp. 1–14.
72. Sharma, P. *et al.* (2019) 'Emerging trends in the novel drug delivery approaches for the treatment of lung cancer', *Chemico-biological interactions*, 309, p. 108720.
73. Shepherd, P. R., Navé, B. T. and Siddle, K. (1995) 'Insulin stimulation of glycogen synthesis and glycogen synthase activity is blocked by wortmannin and rapamycin in 3T3-L1 adipocytes: evidence for the involvement of phosphoinositide 3-kinase and p70 ribosomal protein-S6 kinase', *Biochemical Journal*, pp. 25–28. doi: 10.1042/bj3050025.
74. Siddique, R. *et al.* (2019) 'Qualitative and quantitative analysis of precipitate formation following interaction of chlorhexidine with sodium hypochlorite, neem, and tulsi', *Journal of conservative dentistry: JCD*, 22(1), pp. 40–47.
75. Sridharan, G. *et al.* (2019) 'Evaluation of salivary metabolomics in oral leukoplakia and oral squamous cell carcinoma', *Journal of oral pathology & medicine: official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*, 48(4), pp. 299–306.
76. Stumvoll, M. and Häring, H. (2002) 'Resistin and adiponectin--of mice and men', *Obesity research*, 10(11), pp. 1197–1199.
77. Sujatha, J., Asokan, S. and Rajeshkumar, S. (2018) 'Phytochemical analysis and antioxidant activity of chloroform extract of *Cassia alata*', *Research Journal of Pharmacy and Technology*, p. 439. doi: 10.5958/0974-360x.2018.00081.1.
78. Szendroedi, J. *et al.* (2007) 'Muscle mitochondrial ATP synthesis and glucose transport/phosphorylation in type 2 diabetes', *PLoS medicine*, 4(5), p. e154.
79. Teja, K. V., Ramesh, S. and Priya, V. (2018) 'Regulation of matrix metalloproteinase-3 gene expression in inflammation: A molecular study', *Journal of conservative dentistry: JCD*, 21(6), pp. 592–596.
80. Uysal, K. T. *et al.* (1997) 'Protection from obesity-induced insulin resistance in mice lacking TNF-alpha function', *Nature*, 389(6651), pp. 610–614.
81. Vijayashree Priyadharsini, J. (2019) 'In silico validation of the non-antibiotic drugs acetaminophen and ibuprofen as antibacterial agents against red complex pathogens', *Journal of periodontology*, 90(12), pp. 1441–1448.

82. Vijayashree Priyadharsini, J., Smiline Girija, A. S. and Paramasivam, A. (2018) 'In silico analysis of virulence genes in an emerging dental pathogen *A. baumannii* and related species', *Archives of oral biology*, 94, pp. 93–98.
83. Whitehead, J. P. *et al.* (1998) 'Molecular scanning of the insulin receptor substrate 1 gene in subjects with severe insulin resistance: detection and functional analysis of a naturally occurring mutation in a YMXM motif', *Diabetes*, pp. 837–839. doi: 10.2337/diabetes.47.5.837.
84. White, M. F. and Ronald Kahn, C. (1989) 'Cascade of autophosphorylation in the β -subunit of the insulin receptor', *Journal of Cellular Biochemistry*, pp. 429–441. doi: 10.1002/jcb.240390409.
85. Wild, S. *et al.* (2004) 'Global Prevalence of Diabetes: Estimates for the year 2000 and projections for 2030', *Diabetes Care*, pp. 1047–1053. doi: 10.2337/diacare.27.5.1047.
86. Yamauchi, T. *et al.* (1996) 'Insulin signalling and insulin actions in the muscles and livers of insulin-resistant, insulin receptor substrate 1-deficient mice', *Molecular and cellular biology*, 16(6), pp. 3074–3084.