Physiological Impact of Nicotine on Rats

Hasanain A.J. Gharban ¹, Noor K.H. Al-Quraishy ², Ahmed M.A. Al-Attaby ³, Anmar R.S. Alekabi ⁴, Nabaa J.A. Hijami ⁵, Abeer B.J. Al-Shimmary ⁶

^{1,3,4}College of Veterinary Medicine, University of Wasit, Wasit, Iraq
² University of Wasit, Wasit, Iraq

5,6 Researcher

Emails: hghirban@uowasit.edu.iq ¹, noor.alquraishy@uowasit.edu.iq ², ahmed.surgery.vet@uowasit.edu.iq ³, anmar.rasool@uowasit.edu.iq ⁴, nabaajawher2001@gmail.com ⁵, abeer1999137abeer@gmail.com ⁶

* Corresponding author

Abstract

Nicotine is the well-known chemical compound which responsible for addiction in tobacco smoking and consider as the main cause of mortality worldwide. This study aims to estimate the level of antioxidants and lipid peroxidation in rats exposed to nicotine. Totally, 40 male rats were selected, acclimated and divided equally into two groups; G1 as control and G2 as an experimental group injected subcutaneously with 0.25 ml of nicotine daily for 28 days. Finally, the study animals were served to sampling of blood and lung samples. Results of CAT revealed a significant decreases in G2 (8.84 \pm 0.52 pg/ml) when compared to G1 (18.36 \pm 1.23 pg/ml). Concerning GPx, significant reduction was seen in G2 (215.91 \pm 14.96 pg / ml) in comparison with G1 (418.48 \pm 15.17 pg / ml). For SOD, value of G2 (3.37 \pm 0.1 U / ml) was lowered significantly more than G1 (7.58 \pm 0.17 U / ml). Significantly, higher MDA concentration were seen in G2 (126.25 \pm 5.95 ng / ml) than G1 (53.8 \pm 4.51 ng / ml). In conclusion, we showed that nicotine is a harmful material for lung tissues and has serious systemic side effects which detected by reducing of the antioxidants and induces of furthermore MDA. Moreover studies are of great importance to detect the effect of nicotine on lung development as well as its effect on other body organs.

Keywords: Tobacco, Antioxidants, Lipid peroxidation, Iraq

Introduction

Cigarette smoking is a greater risk for diseases, and a common cause of early deaths in males and females (Kõks et al., 2018). A part from the 21 known illnesses majorly correlated with smoking; many health risks still undifferentiated resulting in various systemic diseases (Carter et al., 2015). Association between the using of tobaccos and health status were initiated based on clinical data concerned to pulmonary tumors that represent 1st problem

definitively related to using of tobaccos (WHO, 2015; Hou et al., 2019). European health experts identified that 7% and 24% of all female and male deaths in developed countries were caused by tobacco consumption (Sinha et al., 2018). In last 50 years, large volume of data was suggested the presence of approximately 4800 different chemical compounds in cigarette involving gases, 10¹⁰ particles of different sizes and several toxics, carcinogenic substances, tumor promoter and mutagenic (Ledda et al., 2017; Nlemedim, 2017). Although, there is a difficulty in determining the active substance, many researchers attributed the tobacco-related health risks to nicotine, nitrosamines and polycyclic hydrocarbons (Etemadi et al., 2023).

Nicotine is a naturally produced alkaloid originated most predominantly from *Duboisia hopwoodii* and tobacco, to be used in almost as an anxiolytic and stimulant (Wylie and Li, 2022). Gastrointestinal tract, respiratory system, urinary bladder and even skin can effectively absorb and metabolize nicotine (Khudhair, 2012; Jaber, 2013; Mishra et al., 2015; Panda and Albano, 2021). Also, nicotine has the ability to crossing the biological membranes as blood brain barrier with binding to the nicotinic cholinergic receptors at nerve terminal, and modulating the release of neurotransmitters like glutamate, serotonin, dopamine, norepinephrine and acetyle choline (Tega et al., 2018; Alhusban et al., 2023). Suppression of apoptosis, inhibition of cell proliferation, induction of chromosome aberration, sister chromatide exchange and increasing the expression of heat shock proteins can be caused by nicotine at the cellular level (Smart et al., 2019).

In Iraq, low available data correlate with the effect of nicotine on the body; hence, the current study was aimed to estimate the levels of antioxidants [catalase (CAT), glutathione perioxidase (GPx) and superoxide dismutase (SOD)] and lipid peroxidation [malondialdehyde (MDA) in rats exposed to nicotine. **Materials and methods**

Ethical approval

Scientific Committee in the College of Medicine (University of Al-Qadisiyah) was approved the work of this study.

Study animals

Totally, 40 male Wistar albino rats of 2 months old and 180-229 gm weight were purchased from the local markets, acclimated for 1 week, and then divided equally and randomly into two groups; G1, in which the study rats were received only normal saline; while rats of G2 were injected subcutaneously the nicotine daily for 28 days at a dose of 0.25 ml. During the

acclimation and experimental periods, the study rats were fed pellet, received tap water and exposed to 12/12 dark / light.

Samples

At the end of experiment, the study rats were euthanized with chloroform and subjected for direct blood sampling into glass gel-free anticoagulant tubes. Post centrifugation (5000 rpm / 3 minutes), the sera were saved frozen in darken containers for serology, and the lung tissues were collected into plastic containers contain 10% neutral buffered saline (NBF) for histology.

Serology

Following the manufacturer instructions (SunLong Biotech, China) of the CAT (Cat.No: SL1084Ra), GPx (Cat.No: SL1033_1Ra), SOD (Cat.No: SL1341Ra) and MDA (Cat.No: SL0475Ra) kits, the samples and Standards of each kit were prepared, processed, and the optical density (OD) were measured at 450 nm using the Microplate Reader (BioTek, USA). After setting the blank control at zero, the concentrations of the samples in each parameter were detected using the Standard Curve.

Statistical analysis

The GraphPad Prism Software was served for identification of significant differences in values of serology among G1 and G2 groups at P<0.05 (Gharban, 2022; Gharban et al., 2023). Values were represented as Mean ± Standard Errors (M±SE).**Results**

Serology

The findings of CAT revealed a significant decreases (P<0.0001) in values of G2 (8.84 \pm 0.52 pg/ml) when compared to G1 (18.36 \pm 1.23 pg/ml), (Figure 1). Concerning GPx, there was significant reduction (P<0.0001) in value of G2 (215.91 \pm 14.96 pg / ml) in comparison with those of G1 (418.48 \pm 15.17 pg / ml), (Figure 2).

For SOD, value of G2 (3.37 \pm 0.1 U / ml) was lowered significantly (P<0.0001) more than value of G1 (7.58 \pm 0.17 U / ml), (Figure 3). Significantly, higher value of MDA was seen in G2 (126.25 \pm 5.95 ng / ml) than G1 (53.8 \pm 4.51 ng / ml), (Figure 4).

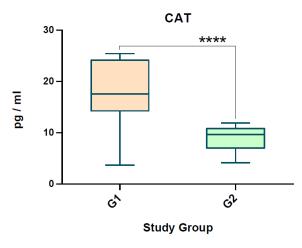


Figure (1): Concentration of CAT among study groups; G1 and G2

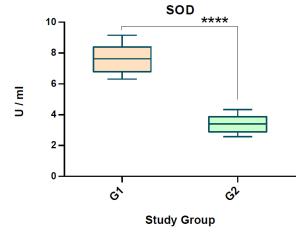


Figure (3): Concentration of SOD among study groups; G1 and G2

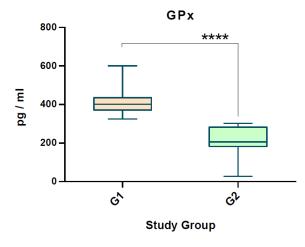


Figure (1): Concentration of GPx among study groups; G1 and G2

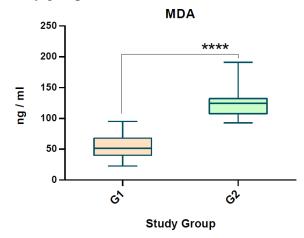


Figure (4): Concentration of MDA among study groups; G1 and G2

Discussion

Nicotine is one of the greatly toxic chemical substances which identified as a major risk factor for lung-related diseases and can potentially cause adverse effects or even death at high dose; 6 mg for children and 50-60 mg for adult (Mayer, 2014; Rowell and Tarran, 2015). The damage to the tissues by nicotine is evidenced in this study by the reduction of antioxidants (CAT, GPx and SOD) and increasing lipid peroxidation (MDA) in blood. Sies (1997) was the first researcher introduced the phrase of "oxidative stress" defining it as a disturbance in the pro-oxidant-antioxidant balance that causes either reduction for antioxidant concentrations or increasing the production the highly diffusible reactive oxygen species (ROS) which

manifests its deleterious effects (Al-Khafaji, 2015; Bardaweel et al., 2018). Some authors have reported that smokers have poorer dietary habits and this result in observable consuming of ascorbic acid which may be the reason for the significant increase of oxidative stress (Shah et al., 2015; Ahmadi-Motamayel et al., 2017; Singh et al., 2019).

CAT is one of the most important antioxidant enzymes, which represent in almost all aerobic organisms. This enzyme is produced by numerous metabolic reactions and decompose the hydrogen peroxide to oxygen and water (Gebicka and Krych-Madej, 2019). Several studies demonstrated that catalase deficiency or malfunctioning is associated with many diseases and can occur as a result of inherited or genetic disorders (Dai et al., 2017; Maciejczyk et al., 2017; Nandi et al., 2019). Other authors found that CAT activity decreased significantly in the blood of smokers attributing this effect to increasing the ROS production and inhibition of CAT activity by nicotine which can result later in mutagenic and cytotoxic effects (Rickert et al., 2011; Crooks et al., 2018; Smart et al., 2019). Raddam et al. (2017) suggested that the decreased activity of CAT is attributed to inactivation of CAT by the cross linking or impairment of nitric oxide synthesis which can bind reversible to ferric iron and inhibiting afterwards CAT activity (Noichri et al., 2013; Silambarasan et al., 2014). Our results were in contrast with previous studies which detected an increasing of CAT levels in smokers (Garg et al., 2006; Reejamol and Swaminathan, 2013).

GPx is a systolic enzyme that catalyzes the reduction hydrogen peroxide radical to alcohol and oxygen. This enzyme has the ability to cure several diseases since increasing the endogenous levels of GPx and resolving of ROS-induced pathology (Prasad et al., 2018; Joshi et al., 2020; Zhang et al., 2021). We agrees the major worldwide reports that revealed the reduction of GPx levels in smokers (Agarwal et al., 2019; Oladunjoye et al., 2022). This reduction could be resulted by fluxing of more hydroxyl radicals and hydrogen peroxide as a result of smoking. In normal conditions, SOD considers as one the most active important antioxidant enzyme as free radical scavenger. Our results showed a significant reduction in concentration of SOD in experimentally G2 when compared to control G1 which agrees with other findings by other authors (Jain and Flora, 2012; Oyeyipo et al., 2014). Also, we agreed with Mohammed and Al-Thwani (2019) who detected that the levels of GPx were decreased significantly while MDA was increased significantly in experimentally mice injected nicotine subcutaneously. In human, Abdul-Rasheed and Al-Rubayee (2013) showed that the plasma MDA level was significantly increased in smokers compared with non-smokers; however, the plasma SOD AND GPx were differed insignificantly.

Mohammed and Al-Thwani (2019) observed that injection of nicotine causes different pathological changes in lung tissues of mice such as lymphocytes infiltration, fibrosis, alveolar edema, hemorrhage, congestion of blood vessels, emphysema and damage to alveoli. Structural modification in cellular proteins due to oxidative stress was also showed in smokers (Stangenberg et al., 2015; Caliri et al., 2021); while others mentioned that the damage in tissue is limited to few protein molecules (Zong et al., 2019). As observed by other researchers, the exposing of lung tissues to great flux of hydroxyle radicals and hydrogen peroxide could reflect in absence of balance in antioxidant production (Tribble and Jones, 1990; Rahal et al., 2014). Raddam et al. (2017) mentioned that nicotine play a role in increasing the generation of hydrogen peroxide and superoxide anion which in turn elevate significantly the production of ROS and cause deleterious injury to alveolar macrophage by cause releasing of proteolytic enzymes.

Conclusion

This study showed that nicotine is a harmful material of serious systemic side effects which detected by reducing of the antioxidants and induces of furthermore MDA. Moreover studies are of great importance to detect the effect of nicotine on lung development as well as its effect on other body organs.

Conflict of interest

No.

Authors' contributions

MAS: Experimental study, collection of blood and tissue samples, and statistical analysis. NED: Tissue processing and serology. Both authors contributed equally in microscopic examination of slides, reading and approving the final copy of the manuscript.

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References

[1] Abdul-Rasheed, O. F., and Al-Rubayee, W. T. (2013). Effects of cigarette smoking on lipid peroxidation and antioxidant status in Iraqi men at Baghdad city. *International Journal of Basic and Applied Sciences*, 2(1), 47-50.

- [2] Agarwal, P., Bagewadi, A., Keluskar, V., and Vinuth, D. P. (2019). Superoxide dismutase, glutathione peroxidase, and catalase antioxidant enzymes in chronic tobacco smokers and chewers: a case–control study. *Indian Journal of Dental Research*, 30(2), 219.
- [3] Ahmadi-Motamayel, F., Falsafi, P., Goodarzi, M. T., and Poorolajal, J. (2017). Evaluation of salivary catalase, vitamin C, and alpha-amylase in smokers and non-smokers: a retrospective cohort study. *Journal of Oral Pathology and Medicine*, 46(5), 377-380.
- [4] Alhusban, A. A., Hammad, A. M., Alzaghari, L. F., Shallan, A. I., and Shnewer, K. (2023). Rapid and sensitive HPLC–MS/MS method for the quantification of dopamine, GABA, serotonin, glutamine and glutamate in rat brain regions after exposure to tobacco cigarettes. *Biomedical Chromatography*, *37*(1), e5513.
- [5] Al-Khafaji, F. M. A. (2015). Role of antioxidant therapy and cessation of smoking in treatment of infertile men. *Al-Qadisiyah Medical Journal*, *11*(20), 133-142.
- [6] Bardaweel, S. K., Gul, M., Alzweiri, M., Ishaqat, A., ALSalamat, H. A., and Bashatwah, R. M. (2018). Reactive oxygen species: The dual role in physiological and pathological conditions of the human body. *The Eurasian journal of medicine*, 50(3), 193.
- [7] Caliri, A. W., Tommasi, S., and Besaratinia, A. (2021). Relationships among smoking, oxidative stress, inflammation, macromolecular damage, and cancer. *Mutation Research/Reviews in Mutation Research*, 787, 108365.
- [8] Carter, B. D., Abnet, C. C., Feskanich, D., Freedman, N. D., Hartge, P., Lewis, C. E., and Jacobs, E. J. (2015). Smoking and mortality—beyond established causes. *New England journal of medicine*, *372*(7), 631-640.
- [9] Crooks, I., Neilson, L., Scott, K., Reynolds, L., Oke, T., Forster, M., and Proctor, C. (2018). Evaluation of flavourings potentially used in a heated tobacco product: chemical analysis, in vitro mutagenicity, genotoxicity, cytotoxicity and in vitro tumour promoting activity. *Food and Chemical Toxicology*, *118*, 940-952.
- [10] Dai, D. F., Chiao, Y. A., Martin, G. M., Marcinek, D. J., Basisty, N., Quarles, E. K., and Rabinovitch, P. S. (2017). Mitochondrial-targeted catalase: extended longevity and

- the roles in various disease models. *Progress in Molecular Biology and Translational Science*, 146, 203-241.
- [11] Etemadi, A., Poustchi, H., Chang, C. M., Calafat, A. M., Blount, B. C., Bhandari, D., and Freedman, N. D. (2023). Exposure to polycyclic aromatic hydrocarbons, volatile organic compounds, and tobacco-specific nitrosamines and incidence of esophageal cancer. *JNCI: Journal of the National Cancer Institute*, djad218.
- [12] Garg, N., Singh, R., Dixit, J., Jain, A., and Tewari, V. (2006). Levels of lipid peroxides and antioxidants in smokers and nonsmokers. *Journal of periodontal research*, 41(5), 405-410.
- [13] Gebicka, L., and Krych-Madej, J. (2019). The role of catalases in the prevention/promotion of oxidative stress. *Journal of inorganic biochemistry*, 197, 110699.
- [14] Gharban, H. A. (2022). Clinical and Serological Diagnosis of Bovine Hypodermosis in Wasit Province. *Revista Electronica de Veterinaria*, 23 (3), 457-466.
- [15] Gharban, H. A., Al-Shaeli, S. J., and Hussen, T. J. (2023). Molecular genotyping, histopathological and immunohistochemical studies of bovine papillomatosis. *Open Veterinary Journal*, *13*(1), 26-41.
- [16] Hou, W., Hu, S., Li, C., Ma, H., Wang, Q., Meng, G., and Zhang, J. (2019). Cigarette smoke induced lung barrier dysfunction, EMT, and tissue remodeling: a possible link between COPD and lung cancer. *BioMed research international*, 2019.
- [17] Jaber, S. H. (2013). An association and genetic polymorphisms of CYP2D6 gene in chronic renal failure patients in AL-Qadisiya province/Iraq. *Al-Qadisiyah Journal of Pure Science*, 18(2), 38-47.
- [18] Jain, A., and Flora, S. J. S. (2012). Dose related effects of nicotine on oxidative injury in young, adult and old rats. *Journal of environmental biology*, *33*(2), 233-238.
- [19] Joshi, B., Singh, S., Sharma, P., Mohaparata, T., and Kumar, P. (2020). Effect of Cigarette Smoking on Selected Antioxidant Enzymes and Oxidative Stress Biomarkers. *Journal of Clinical and Diagnostic Research*, 14(10).
- [20] Khudhair, S. H. (2012). Evaluation of lipid profile & BMI in smokers & non-smokers individuals. *Al-Qadisiyah Medical Journal*, 8(14), 239-250.

- [21] Kõks, G., Fischer, K., and Kõks, S. (2018). Smoking-related general and cause-specific mortality in Estonia. *BMC public health*, *18*, 1-11.
- [22] Ledda, C., Loreto, C., Zammit, C., Marconi, A., Fago, L., Matera, S., and Rapisarda, V. (2017). Non-infective occupational risk factors for hepatocellular carcinoma: A review. *Molecular Medicine Reports*, 15(2), 511-533.
- [23] Maciejczyk, M., Mikoluc, B., Pietrucha, B., Heropolitanska-Pliszka, E., Pac, M., Motkowski, R., and Car, H. (2017). Oxidative stress, mitochondrial abnormalities and antioxidant defense in Ataxia-telangiectasia, Bloom syndrome and Nijmegen breakage syndrome. *Redox Biology*, 11, 375-383.
- [24] Mayer, B. (2014). How much nicotine kills a human? Tracing back the generally accepted lethal dose to dubious self-experiments in the nineteenth century. *Archives of toxicology*, 88(1), 5-7.
- [25] Mishra, A., Chaturvedi, P., Datta, S., Sinukumar, S., Joshi, P., and Garg, A. (2015). Harmful effects of nicotine. *Indian Journal of Medical and Paediatric Oncology*, 36(01), 24-31.
- [26] Mohammed, B. J., and Al-Thwani, A. N. (2019). Evaluation the effect of nicotine injection on the lungs of mice. *Journal of Reports in Pharmaceutical Sciences*, 8(1), 34-38.
- [27] Nandi, A., Yan, L. J., Jana, C. K., and Das, N. (2019). Role of catalase in oxidative stress-and age-associated degenerative diseases. *Oxidative medicine and cellular longevity*, 2019.
- [28] Nlemedim, O. N. (2017). Organic chemical compounds in different Brands of Cigarette Smoke (Doctoral dissertation, Texas Southern University).
- [29] Noichri, Y., Chalghoum, A., Chkioua, L., Baudin, B., Ernez, S., Ferchichi, S., and Miled, A. (2013). Low erythrocyte catalase enzyme activity is correlated with high serum total homocysteine levels in Tunisian patients with acute myocardial infarction. *Diagnostic pathology*, 8, 1-7.
- [30] Oladunjoye, Z. M., Adejumo, E. N., Ganiyu, A. O., and Quadri, J. A. (2022). Serum Glutathione Peroxidase and Superoxide Dismutase Levels in Young Adult Active

- Smokers and Non-Smokers in a Southwest Based Tertiary Institution. *Sokoto Journal of Medical Laboratory Science*, 7(4).
- [31] Oyeyipo, I. P., Raji, Y., and Bolarinwa, A. F. (2014). Nicotine alters serum antioxidant profile in male albino rats. *North American Journal of Medical Sciences*, *6*(4), 168.
- [32] Panda, B., and Albano, G. (2021). Synthetic Methods for the Preparation of Conformationally Restricted Analogues of Nicotine. *Molecules*, 26(24), 7544.
- [33] Prasad, N., Ramteke, P., Dholia, N., and Yadav, U. C. (2018). Therapeutic interventions to block oxidative stress-associated pathologies. In *Immunity and inflammation in health and disease* (pp. 341-362). Academic Press.
- [34] Raddam, Q. N., Zeidan, M. M., Asaad, N. K., and Abdulrahman, M. A. (2017). Smoking effects on blood antioxidants level: lactate dehydrogenase, catalase, superoxide dismutase and glutathione peroxidase in university students. *Journal of Clinical and Experimental Pathology*, 7(6), 2161-0681.
- [35] Rahal, A., Kumar, A., Singh, V., Yadav, B., Tiwari, R., Chakraborty, S., and Dhama, K. (2014). Oxidative stress, prooxidants, and antioxidants: the interplay. *BioMed research international*, 2014.
- [36] Reejamol, M. K., and Swaminathan, M. (2013). Estimation of lipid peroxides and antioxidants in smokers and non-smokers with periodontitis. *King Saud University Journal of Dental Sciences*, 4(2), 53-56.
- [37] Rickert, W. S., Trivedi, A. H., Momin, R. A., Wagstaff, W. G., and Lauterbach, J. H. (2011). Mutagenic, cytotoxic, and genotoxic properties of tobacco smoke produced by cigarillos available on the Canadian market. *Regulatory Toxicology and Pharmacology*, 61(2), 199-209.
- [38] Rowell, T. R., and Tarran, R. (2015). Will chronic e-cigarette use cause lung disease?. American Journal of Physiology-Lung Cellular and Molecular Physiology, 309(12), L1398-L1409.
- [39] Shah, A. A., Khand, F., and Khand, T. U. (2015). Effect of smoking on serum xanthine oxidase, malondialdehyde, ascorbic acid and α-tocopherol levels in healthy male subjects. *Pakistan journal of medical sciences*, 31(1), 146.

- [40] Sies, H. (1997). Oxidative stress: oxidants and antioxidants. *Experimental Physiology: Translation and Integration*, 82(2), 291-295.
- [41] Silambarasan, T., Manivannan, J., Krishna Priya, M., Suganya, N., Chatterjee, S., and Raja, B. (2014). Sinapic acid prevents hypertension and cardiovascular remodeling in pharmacological model of nitric oxide inhibited rats. *PloS one*, *9*(12), e115682.
- [42] Singh, R., Mahdi, A. A., Singh, R. K., Gierke, C. L., and Cornelissen, G. (2018). Effect of gender, age, diet and smoking status on the circadian rhythm of ascorbic acid (vitamin C) of healthy Indians. *Journal of Applied Biomedicine*, *16*(3), 180-185.
- [43] Sinha, D. N., Suliankatchi, R. A., Gupta, P. C., Thamarangsi, T., Agarwal, N., Parascandola, M., and Mehrotra, R. (2018). Global burden of all-cause and cause-specific mortality due to smokeless tobacco use: systematic review and meta-analysis. *Tobacco control*, 27(1), 35-42.
- [44] Smart, D. J., Helbling, F. R., Verardo, M., McHugh, D., and Vanscheeuwijck, P. (2019). Mode-of-action analysis of the effects induced by nicotine in the in vitro micronucleus assay. *Environmental and Molecular Mutagenesis*, 60(9), 778-791.
- [45] Stangenberg, S., Nguyen, L. T., Chen, H., Al-Odat, I., Killingsworth, M. C., Gosnell, M. E., and Saad, S. (2015). Oxidative stress, mitochondrial perturbations and fetal programming of renal disease induced by maternal smoking. *The international journal of biochemistry and cell biology*, 64, 81-90.
- [46] Tega, Y., Yamazaki, Y., Akanuma, S. I., Kubo, Y., and Hosoya, K. I. (2018). Impact of nicotine transport across the blood–brain barrier: carrier-mediated transport of nicotine and interaction with central nervous system drugs. *Biological and Pharmaceutical Bulletin*, 41(9), 1330-1336.
- [47] Tribble, D. L., and Jones, D. P. (1990). Oxygen dependence of oxidative stress: rate of NADPH supply for maintaining the GSH pool during hypoxia. *Biochemical pharmacology*, 39(4), 729-736.
- [48] WHO (World Health Organization), (2015). WHO global report on trends in prevalence of tobacco smoking 2015. World Health Organization.

- [49] Wylie, S., and Li, H. (2022). Historical and scientific evidence for the origin and cultural importance to Australia's first-nations peoples of the laboratory accession of Nicotiana benthamiana, a model for plant virology. *Viruses*, *14*(4), 771.
- [50] Zhang, C., Wang, X., Du, J., Gu, Z., and Zhao, Y. (2021). Reactive oxygen species-regulating strategies based on nanomaterials for disease treatment. *Advanced Science*, 8(3), 2002797.
- [51] Zong, D., Liu, X., Li, J., Ouyang, R., and Chen, P. (2019). The role of cigarette smoke-induced epigenetic alterations in inflammation. *Epigenetics and chromatin*, 12(1), 1-25.