# Serological and Molecular Detection of *Brucella Melitensis*, Wasit, Iraq

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## **Abstract**

The current study was carried out to identify the prevalence of *Brucella melitensis* infections in humans in Wasit province serologically using the Rose-Bengal test (RBT) and molecularly by the polymerase chain reaction (PCR). The venous blood samples collected randomly from totally 300 individuals into anticoagulant-EDTA plastic tube were tested initially by RBT, and the positive samples were confirmed later by the PCR assay. The findings revealed that 24.67% (74/300) of study population was serologically positives by RBT. Molecularly, there were 39.19% (29/74) positive individuals by the conventional PCR. Regarding the region, significant increases (P<0.0456) in positivity were reported in Al-Numaniyah (17.33%) while significant reduction was seen in Badra (8%) when compared to other study regions; Al-Kut (8%) and Al-Hai (10.67%). In relation to the study period, insignificant differences (P<0.0581) were detected between the positive values of August (8%) and September (11.33%). For sex, the positive higher values were identified significantly (P<0.0363) in males (12.21%) than females (3.45%). Among age groups, the findings of positivity were increased significantly (P<0.04) in >20-40 years old (9.8%) and > 40 years old (10.92%) than those of  $\leq 20$  years old (3.57%). In conclusion, the combination of different diagnostic methods provided more insurance and supported high reliably findings. Also, the application of molecular-PCR assays is of great value since it provides a more sensitive and specific data about the status of infection in addition to their ability in detection of organism in blood samples efficiently. In people and animals living in the same households, brucellosis allows for identification and quantification of risk factors for brucellosis transmission at the household level. However, the prevalence of infection in animal reservoirs can provide a key to its prevalence in humans; therefore, surveys of brucellosis in different domestic and wild animals are required

**Keywords**: Brucellosis, Rose-Bengal, PCR, Season, Age, Sex

## Introduction

Brucellosis as one of the ancient and most widespread zoonotic diseases affects humans as well as the animals, a bacterial disease resulted by a Gram-negative, facultative intracellular bacterium of the genus *Brucella* (Pradeepkiran et al., 2021). Currently, the scientifically communities referred for 6 separately species that having different morphological characteristics based on presence or absence the *O*15 polysaccharide (OPS) in its smooth lipopolysaccharide (SLPS) that covering the outer membranes (Coelho et al., 2019; Akoko et al., 2021). Many researchers thought that SLPS have the ability for evading innate immunity as well as for inducing the inflammation by activation the cytokines. This evasion technique increase the virulence of SLPS when compared to rough lipopolysaccharide (RLPS) strain,

unable for inhibiting the immune responses of the host and greatly impact the innate immunes (Dykman, 2020; Shatti et al., 2021). Other differences are the cleavages of DNAs through a phage specifically for *Brucella* as approximately 40 phages could causes a fully lysing in the genomes of one or more species. R/C phages cause lysing of phages in RLPS of Tb, Wb, and Iz1 differentiating of species within SLPS are susceptibly lysing with the Iz1 and have different lysing with WB (Matope et al., 2009; Tevdoradze et al., 2015; Saxena, 2021). Additionally, variations are not found between the species of *Brucella*, but exist in other types of *Brucella* that having several distinct biovars. This difference in biovar has variation in biochemically and growing feature like variation in capabilities for growing within the existence CO<sub>2</sub>, to result in H<sub>2</sub>S, to agglutinating the A and M antisera, and to growing with existence of urea (Al-Mariri, 2015; García Lobo et al., 2019).

In a number of developed countries, brucellosis has been eradicated through test and slaughter, vaccination and restriction of animal movements (Bahmani and Bahmani, 2022). However, many developing countries remain with the highest disease incidence rates such as those located within Asia (Laine et al., 2022). In Iraq, human brucellosis was confirmed firstly by Al-Zahawi in 1937 (Hussein et al., 2019); however, recent information about the annual prevalence of brucellosis remained low and need to furthermore support. Therefore, the current study was carried out to identify the prevalence of *B. melitensis* infections in humans in Wasit province (Iraq), serologically using the RBT and molecularly by the conventional PCR.

# Materials and methods

# Ethical approval

The study approves by the Scientific Committee of the Department of Basic Science in the College of Density (University of Wasit, Wasit, Iraq).

## **Samples**

Totally, 300 individuals of different ages and sexes were selected randomly from different areas in Wasit province (Iraq) during August and September (2023). Each individual was subjected for draining 5 ml of venous blood into glass vacutainer, and then divided equally into an EDTA-anticoagulant plastic tube and free-anticoagulant glass gel tube. The first tube was kept frozen for later molecular testing while the second tube was centrifuged (5000 rpm / 5 minutes) and the obtained sera were tested directly by the RBT.

# RBT

As described by Al-Hassani et al. (2018), the Rose-Bengal Antigen Kit (Wuhan Ecalbio, China) was served to detect the suspected seropositive samples to brucellosis.

# Conventional PCR

According the manufacturer instructions of the G-spin Total DNA Extraction Kit (Intron, Korea), the EDTA-blood samples were thawed in water bath and processed for extraction of DNAs. All extracted DNAs were tested spectrophotometrically using the Nanodrop System (Thermo-scientific, UK) to detect the concentration and purity of each sample. Targeting the *16S rRNA* gene, one set of primers was designed [(F: 5′-AGG CCC TAG GGT TGT AAA GC-′3) and (R: 5′-GTT TAC GGC GTG GAC TAC CA-′3)] based on the GenBank-NCBI data (OR053959.1), the MasterMix tubes were prepared at a final volume of 20 μl. PCR reaction was carried out using the thermocycler system as following; 1 cycle initial denaturation (95°C/5 Minutes); 35 cycles of denaturation (95°C/30 seconds), annealing (58°C/30 seconds) and extension (72°C/30 seconds); and 1 cycle final extension (72°C/7 minutes). Electrophoresis

using the agarose-gel (1.5%) stained with Ethidium Bromide was performed at 80AM and 100 Volt for 90 minutes. According to bands of the standard Ladder Marker (100-1500 bp), the positive PCR products were identified by the ultraviolet (UV) transilluminator (Clinex, China) at approximately 378 bp.

# Statistical analysis

The *t*-test in the GraphPad Prism Software (*version 6.0.1*) was applied to estimate significant differences between the obtained results at \* (P<0.05), (Ibraheim et al., 2023).

# **Results**

The findings revealed that 24.67% (74/300) of study population was serologically positives by RBT (Figure 1). Molecularly, there were 39.19% (29/74) positive individuals by the conventional PCR (Figures 2, 3)

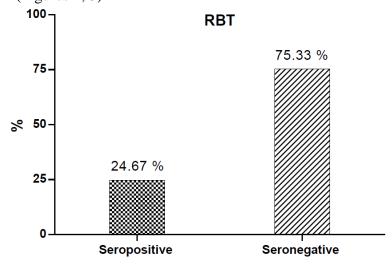


Figure (1): Total results of RBT among totally 300 serum samples

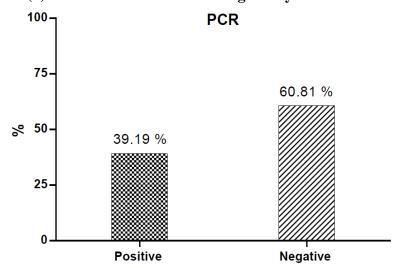


Figure (1): Total results of RBT among totally 300 serum samples



Figure (3): Agarose-gel electrophoresis of some positive PCR products at 80Am and 100 Volt for 90 minutes; Lane M represent ladder marker (100-1500 bp), while lanes 1-16 represent positive samples at 378 bp

Regarding the region, significant increases (P<0.0456) in positivity were reported in Al-Numaniyah (17.33%) while significant reduction was seen in Badra (8%) when compared to other study regions; Al-Kut (8%) and Al-Hai (10.67%), (Table 1).

Table (1): Association of positive PCR results to region factor (Total No: 29)

Factor	Group	Total No.	Positive	
	Al-Numaniyah	75	13	17.33 *
	Al-Kut	75	6	8
Region	Badra	75	2	2.67
	Al-Hai	75	8	10.67
	P-value	<u>.</u>	•	0.0456

In relation to the study period, insignificant differences (P<0.0581) were detected between the positive values of August (8%) and September (11.33%), (Table 2).

Table (2): Association of positive PCR results to period factor (Total No. 29)

	Group	Total No.	Positive	
	August	150	12	8
Period	September	150	17	11.33
	P-value			0.0581

For sex, the positive higher values were identified significantly (P<0.0363) in males (12.21%) than females (3.45%), (Table 3)

Table (3): Association of positive PCR results to sex factor (Total No: 29)

Factor	Group	Total No.	Positive (No: 29)	
	Female	87	3	3.45
Sex	Male	213	26	12.21 *
	P-value			0.0363

Among age groups, the findings of positivity were increased significantly (P<0.04) in >20-40 years old (9.8%) and > 40 years old (10.92%) than those of  $\leq$ 20 years old (3.57%), (Table 4).

Table (4): Association of positive PCR results to age factor (Total No: 29)

Factor	Group	Total No.	Positive (No: 29)	
	≤ 20	28	1	3.57
Age	> 20-40	153	15	9.8 *
	> 40	119	13	10.92 *
P-value				0.04

## **Discussion**

In Iraq, the disease remains one of the major disease problems affecting the livestock and human health. In comparison with our findings, the overall prevalence of brucellosis was 23% in Iraq (Shatti et al., 2021), 8.8% in Kyrgyzstan (Bonfoh et al., 2012), 26.7% in Turkey (Yumuk and O'Callaghan, 2012), 16.4% in Kenya (Osoro et al., 2015), 17% in Uganda (Tumwine et al., 2015), 7.2% in Portugal (Pelerito et al., 2017), 9.44% in Egypt (Abdelbaset et al., 2018), 12.8% in Saudi Arabia (Alkahtani et al., 2020) and 34.88% in Mexico (Guzmán-Bracho et al., 2020). This difference in prevalence of B. melitensis might follow diagnostic assay used in investigation (serological type, sensitivity and specificity of the instruments used, specimen type, target antigen or antibody), target population (age, breed, individual or group, herd size, sampling and data collection methods) and environment (geographical location of the study area, presence of other domestic and/or wild animals, treatment, prevention and control). Duran-Ferrer et al. (2004) showed that high levels of circulating antibodies are associated with active Brucella infection. Several studies have shown that antibody persists for years, even after successful treatment and complete recovery from acute brucellosis, in healthy animals exposed to repeated exposure to organism (Buchanan et al., 2004; Gupta et al., 2010; Almuneef and Memish, 2013).

As serological assays affected by cross-reactivity, low sensitivity and specificity in diagnosis of Brucella species, it is recommended to using a more practical, faster, safer, simpler, and accurate technique as molecular PCR that shown to be more sensitive in diagnosis of brucellosis in pure cultures (Sergueev et al., 2017), blood (Acıkgöz et al., 2018), milk and cheese (Altun et al., 2017) and naturally infected animal organs (Akhtar et al., 2017). PCRbased methods are highly applicable and effective for diagnosing acute brucellosis in early stages of disease, studying predictive biomarkers for post-treatment management, and monitoring disease progression for early detection of recurrence (Al-Hassani et al., 2018; Dadar et al., 2019). The PCR method has several advantages in the diagnosis of human brucellosis, including rapidity, safety, high sensitivity, and specificity. Also, it can be considered to complement the traditional methods adopted by serology and/or culture (Rahimi et al., 2020; Kılıç et al., 2021). Similarly, detection of Brucella by PCR has also emerged as a new, more effective diagnostic tool for different stages of diseases (acute, subacute and chronic) and different clinical specimens such as serum, urine, cerebrospinal fluid, synovial or pleural fluid, and pus (Ulu Kilic et al., 2013; Sulayman et al., 2020; Di Bonaventura et al., 2021).

Changes in the prevalence of brucellosis in the areas studied are consistent with changes reported by other researchers (Al-Mossawy, 2017; Al-Hakami et al., 2019; Shatti et al., 2021). Ingestion, direct contact, and inhalation are the main routes of infection, but the relative

importance of the method and route of transmission of the pathogen varies depending on the epidemiological region, pet store, professional group, and exposed consumer (Mufinda et al., 2017; Al-Zubaidy, 2018; Gharban et al., 2023). Buzgan et al. (2010) conducted a retrospective analysis of 1,028 incidents occurred between 1998 and 2007, with explaining that these cases might be linked to consumption of unpasteurized dairy products or contact with animals, reflecting the rural nature of the area. Socioeconomic and educational factors are also independent risk factors as concluded by Araj and Azzam (1996) as the risk of brucellosis is higher among people in high-risk occupations. In contrast, Sofian et al. (2008) showed that occupation and education level are not significant risk factors. However, infection in humans may be related to various factors such as lifestyle, economic level of social life, social eating habits and animal management (Alavi et al., 2007; Bosilkovski et al., 2015). A number of studies recorded that handling of infected animals, caring of newborns, cleaning animal nests, and disposing to aborted fetuses increased the risk of infection in certain areas than other (Shareef, 2006; Al-Mossawy, 2017).

The association of brucellosis with temporal or seasonal factors is controversial because some symptoms could persist for long time. Additionally, the incubation period of brucellosis is usually 1–3 weeks, but it may take several months for symptoms of infection to appear (Seleem et al., 2010; Jaffe, 2016). However, seasonal distribution of human brucellosis may indicate that the weather of this month is most favorable to survival of *Brucella* and spreading of infection (Zhou et al., 2018). Alkahtani et al. (2020) showed that the seasonal nature of brucellosis may be associated with various risk factors including rain, sunlight, and consumption the products of infected animals in large numbers. Pal et al. (2020) revealed that brucellosis can occur at any time of a year, but the peak incidence of human brucellosis might closely relate to the month of birth and gestation in field animals due to increasing the excretion of *Brucella* at birth into the environment.

There is debate about the relationship between the prevalence of brucellosis and the sex factor. Although, various studies shown that females are more affected by brucellosis than males (Daood et al., 2020; Ibrahim et al., 2021; Shatti et al., 2021), others reported no gender association to susceptibility (Ahmed et al., 2010; Rahman et al., 2012; Al-Mossawy, 2017). In agreement with our findings, studies conducted in Uganda (Tumwine et al., 2015) and Saudi Arabia (Alkahtani et al., 2020) showed the gender-specific results and a high prevalence of disease in males. Jabary and Al-Samarraee (2015) observed that sexually mature animals are more motivated than immature animals, which might be related to the fact that sex hormones, meso-erythritol present in males (testicles and seminal glands) and erythritol found in allantoic fluid of females stimulate the growing and reproduction of *Brucella*.

Statistical analysis of the study data showed the role of age as a risk factor to brucellosis. In Iraq, Daood et al. (2020) found that the prevalence of brucellosis was highest in males in the age group (31–40 years), followed by >40 and 11–20 years old. Buzgan et al. (2010) found that the prevalence of brucellosis was highest in patients aged 14 to 34 years, which might be reflected by obtaining immunity against secondary infections as the population ages. A study conducted in Lebanon found that the incidence of brucellosis increases with age (Kalaajieh, 2000). Tumwine et al. (2015) showed that young people in school and middle-aged people engaged in professional activities are more susceptible to brucellosis. Another study found that brucellosis is more common in young people especially where raising livestock existed at an

early age (Gur et al. 2003). Alkahtani et al. (2020) observed the high incidence of brucellosis in adults due to the greater frequency of exposure of older ages to infected animals, and children at low risk because the less likely come into contact with infected animals. Rahman et al. (2012) showed that people aged 40 to 80 years are more susceptible to brucellosis. However, Sofian et al. (2008) did not show any significant difference in terms of age factors.

# Conclusion

The combination of different diagnostic methods provided more insurance and supported high reliably findings. Also, the application of molecular-PCR assays is of great value since it provides a more sensitive and specific data about the status of infection in addition to their ability in detection of organism in blood samples efficiently. In people and animals living in the same households, brucellosis allows for identification and quantification of risk factors for brucellosis transmission at the household level. However, the prevalence of infection in animal reservoirs can provide a key to its prevalence in humans; therefore, surveys of brucellosis in different domestic and wild animals are required.

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## **Conflicts of interest**

No.

#### **Funds**

No external funds were received.

# **Data availability**

All data were used within this manuscript.

## References

- [1] Abdelbaset, A. E., Abushahba, M. F., Hamed, M. I., and Rawy, M. S. (2018). Sero-diagnosis of brucellosis in sheep and humans in Assiut and El-Minya governorates, Egypt. *International journal of veterinary science and medicine*, 6, S63 S67.
- [2] Açıkgöz, Z. C., Başyiğit, T., Zeybek, H., and Durmaz, R. (2018). Comparison of two commercial DNA extraction kits and PCR master mixes for the detection of *Brucella* from blood samples and blood culture bottles. *Mikrobiyoloji bulteni*, 52(2), 135 146.
- [3] Ahmed, M. O., Elmeshri, S. E., Abuzweda, A. R., Blauo, M., Abouzeed, Y. M., Ibrahim, A., and Elrais, A. (2010). Seroprevalence of brucellosis in animals and human populations in the western mountains region in Libya, December 2006–January 2008. *Eurosurveillance*, 15(30), 19625.
- [4] Akhtar, R., Anwar, M. N., Khan, I., El Adawy, H., Aslam, A., Mustafa, G., and Naz, S. (2017). Pathological investigations of organ affinity of *Brucella* species and their cross species transmission. *Pakistan Veterinary Journal*, *37*(3), 372 374.
- [5] Akoko, J. M., Pelle, R., Lukambagire, A. S., Machuka, E. M., Nthiwa, D., Mathew, C., and Ouma, C. (2021). Molecular epidemiology of Brucella species in mixed livestock-human ecosystems in Kenya. *Scientific reports*, 11(1), 8881.
- [6] Alavi, S. M., Rafiei, A., and Nikkhooi, A. (2007). The effect of lifestyle on brucellosis among nomads in Khuzestan province of Iran. *Pakistan Journal of Medical Sciences*, 23(3), 358.

- [7] Al-Hakami, A. M., Alqahtani, A. J., Moosa, R. A., Kadasah, S. K., Gofashe, T. Y., Binzafrah, A. F., and Hamid, M. E. (2019). Seroprevalence of brucellosis among exposed agro pastoral communities in southern Saudi Arabia. *Asian Pacific Journal of Tropical Medicine*, *12*(12), 545.
- [8] Al-Hassani, M.K.A., Al-Gharban, H.A.J., and Manher, L.F. (2018). Application of Three Diagnostic Serologic Techniques to Detect of Dromedary Camel's Brucellosis. *Al-Qadisiyah journal For pure science*, 23 (2), 61-74.
- [9] Alkahtani, A. M., Assiry, M. M., Chandramoorthy, H. C., Al Hakami, A. M., and Hamid, M. E. (2020). Sero prevalence and risk factors of brucellosis among suspected febrile patients attending a referral hospital in southern Saudi Arabia (2014–2018). *BMC infectious diseases*, 20(1), 1 8.
- [10] Al-Mariri, A. (2015). Isolation of Brucella melitensis strains from Syrian bovine milk samples. *Bulgarian Journal of Veterinary Medicine*, 18(1).
- [11] Al-Mossawy, F.K.S. (2017). Isolation and detection of Brucella abortus infection and its biovars in human using PCR technique and molecular genetic sequence in Middle and Southern of Iraq. MSc thesis, College of Veterinary Medicine, University of Bahdad, Baghdad, Iraq.
- [12] Almuneef, M., and Memish, Z. A. (2013). Prevalence of *Brucella* antibodies after acute brucellosis. *Journal of chemotherapy*, *15*(2), 148 151.
- [13] Altun, S. K., Yiğin, A., Gürbilek, S. E., Gürbüz, S., Demirci, M., Keskin, O., and Tel, O. Y. (2017). An enzyme linked immunosorbent assay for *Brucella* specific antibody and real time PCR for detecting *Brucella* spp. in milk and cheese in Şanlıurfa, Turkey. *Pak Vet J*, *37*(1), 39 42.
- [14] Al-Zubaidy, I. A. (2018). Study the Immunogenisity of Enterobacterial Common Antigen Against Challenge with Brucella melitensis in Guinea Pigs. Indian Journal of Natural Sciences, 9 (50), 1-10.
- [15] Araj, G. F., and Azzam, R. A. (1996). Seroprevalence of *Brucella* antibodies among persons in high risk occupation in Lebanon. *Epidemiology and Infection*, 117(2), 281 288.
- [16] Bahmani, N., and Bahmani, A. (2022). A review of brucellosis in the Middle East and control of animal brucellosis in an Iranian experience. *Reviews and Research in Medical Microbiology*, *33*(1), e63-e69.
- [17] Bonfoh, B., Kasymbekov, J., Dürr, S., Toktobaev, N., Doherr, M. G., Schueth, T., and Schelling, E. (2012). Representative seroprevalences of brucellosis in humans and livestock in Kyrgyzstan. *EcoHealth*, *9*(2), 132 138.
- [18] Bosilkovski, M., Krteva, L., Caparoska, S., Labacevski, N., and Petrovski, M. (2015). Childhood brucellosis: review of 317 cases. *Asian Pacific journal of tropical medicine*, 8(12), 1027 1032.
- [19] Buchanan, T. M., Sulzer, C. R., Frix, M. K., and Feldman, R. A. (2004). Brucellosis in the United States, 1960 1972: an abattoir associated disease. *Medicine*, *53*(6), 415 425.
- [20] Buzgan, T., Karahocagil, M. K., Irmak, H., Baran, A. I., Karsen, H., Evirgen, O., and Akdeniz, H. (2010). Clinical manifestations and complications in 1028 cases of

- brucellosis: a retrospective evaluation and review of the literature. *International journal of infectious diseases*, 14(6), e469 e478.
- [21] Coelho, A. C., Coelho, A., Quintas, H., Fernandes, C., Saavedra, M. J., and Simões, J. (2019). Pathogenesis of Brucella. *Brucellosis in Goats and Sheep: an endemic and re-emerging old zoonosis in the 21st century*, 99-126.
- [22] Dadar, M., Shahali, Y., and Wareth, G. (2019). Molecular diagnosis of acute and chronic brucellosis in humans. *Microbial technology for the welfare of society*, 223-245.
- [23] Daood, I. I., Zajmi, A., Nouri, H. S., and Al Jubory, D. I. H. (2020). Seroprevalence of Brucellosis from the city Mosul Iraq. *International Journal of Psychosocial Rehabilitation*, 24(2), 3690 3696.
- [24] Di Bonaventura, G., Angeletti, S., Ianni, A., Petitti, T., and Gherardi, G. (2021). Microbiological laboratory diagnosis of human brucellosis: An overview. *Pathogens*, *10*(12), 1623.
- [25] Duran Ferrer, M., Leon, L., Nielsen, K., Caporale, V., Mendoza, J., Osuna, A., and Garrido, F. (2004). Antibody response and antigen specific gamma interferon profiles of vaccinated and unvaccinated pregnant sheep experimentally infected with *Brucella melitensis*. *Veterinary microbiology*, *100*(3 4), 219 231.
- [26] Dykman, L. A. (2020). Gold nanoparticles for preparation of antibodies and vaccines against infectious diseases. *Expert review of vaccines*, 19(5), 465-477.
- [27] García Lobo, J. M., Ortiz, Y., Gonzalez-Riancho, C., Seoane, A., Arellano-Reynoso, B., and Sangari, F. J. (2019). Polymorphisms in Brucella carbonic anhydrase II mediate CO2 dependence and fitness in vivo. *Frontiers in Microbiology*, *10*, 2751.
- [28] Gharban, H. A., Al-Ghuraibawi, H. N., Al-Rubaye, Z. A., Jahlol, H. A., Al-Zergany, A. A., and Al-Abedi, G. J. (2023). Most Clinically Detected Viral Diseases in Field Animals of Wasit Province, Iraq. *Annals of the Romanian Society for Cell Biology*, 27(01), 154-168.
- [29] Gupta, V. K., Kumari, R., Vohra, J., Singh, S. V., and Vihan, V. S. (2010). Comparative evaluation of recombinant BP26 protein for serological diagnosis of *Brucella melitensis* infection in goats. *Small Ruminant Research*, *93*(2 3), 119 125.
- [30] Gur, A., Geyik, M. F., Dikici, B., Nas, K., Çevik, R., Saraç, J., and Hosoglu, S. (2003). Complications of brucellosis in different age groups: a study of 283 cases in southeastern Anatolia of Turkey. *Yonsei medical journal*, 44(1), 33 44.
- [31] Guzmán Bracho, C., Salgado Jiménez, B., Beltrán Parra, L. G., Hernández Monroy, I., Vargas Pino, F., Rodríguez, D., and Díaz Quiñónez, J. A. (2020). Evaluation of serological diagnostic tests of human brucellosis for prevention and control in Mexico. *European Journal of Clinical Microbiology and Infectious Diseases*, 39(3), 575 581.
- [32] Hussein, A. A., Alzubaidi, A. F. A., and Musa, I. (2019). Incidence of brucellosis in Iraq during 2017. *Biochemical and Cellular Archives*, 19(1).
- [33] Ibraheim, H. K., Fayez, R. A., Jasim, A. S., and Gharban, H. A. (2023). Role of nuc gene in Staphylococcus aureus to phagocytic activity in different cattle infections. *Open Veterinary Journal*, *13*(8), 1021-1026.

- [34] Ibrahim, M., Schelling, E., Zinsstag, J., Hattendorf, J., Andargie, E., and Tschopp, R. (2021). Sero prevalence of brucellosis, Q fever and rift valley fever in humans and livestock in Somali region, Ethiopia. *PLOS Neglected Tropical Diseases*, *15*(1), e0008100.
- [35] Jabary, O.M. and Al Samarraee, I.A. (2015). Detection of *Brucella* antibodies of sheep and goats by using two serological tests in Al Sulaimanya governorate. *The Iraqi Journal of Veterinary Medicine (ISSN P: 1609 5693 ISSN E: 2410 7409)*, 39(2), 32 37.
- [36] Jaff, D. (2016). Brucellosis in Iraqi Kurdistan: an overview. *Journal of Entomology and Zoology Studies*, 4(4), 1113 1115.
- [37] Kalaajieh, W. K. (2000). Epidemiology of human brucellosis in Lebanon in 1997. *Médecine et maladies infectieuses*, 30(1), 43 46.
- [38] Kılıç, S., Çelebi, B., and Turan, M. (2021). *Brucella melitensis* and *Brucella abortus* genotyping via real time PCR targeting 21 variable genome loci. *Journal of Microbiological Methods*, 180, 106125.
- [39] Laine, C. G., Scott, H. M., and Arenas-Gamboa, A. M. (2022). Human brucellosis: Widespread information deficiency hinders an understanding of global disease frequency. *PLOS Neglected Tropical Diseases*, *16*(5), e0010404.
- [40] Matope, G., Bhebhe, E., Muma, J. B., Skjerve, E., and Djønne, B. (2009). Characterization of some Brucella species from Zimbabwe by biochemical profiling and AMOS-PCR. *BMC research notes*, 2, 1-6.
- [41] Mufinda, F. C., Boinas, F., and Nunes, C. (2017). Prevalence and factors associated with human brucellosis in livestock professionals. *Revista de saude publica*, *51*, 57.
- [42] Osoro, E. M., Munyua, P., Omulo, S., Ogola, E., Ade, F., Mbatha, P., and Guerra, M. (2015). Strong association between human and animal *Brucella* seropositivity in a linked study in Kenya, 2012–2013. *The American journal of tropical medicine and hygiene*, 93(2), 224 231.
- [43] Pal, M., Kerorsa, G. B., Desalegn, C., and Kandi, V. (2020). Human and Animal Brucellosis: A Comprehensive Review of Biology, Pathogenesis, Epidemiology, Risk Factors, Clinical Signs, Laboratory Diagnosis. *American Journal of Infectious Diseases*, 8(4), 118-126.
- [44] Pelerito, A., Cordeiro, R., Matos, R., Santos, M. A., Soeiro, S., Santos, J., and Núncio, S. (2017). Human brucellosis in Portugal—Retrospective analysis of suspected clinical cases of infection from 2009 to 2016. *PLoS One*, *12*(7), e0179667.
- [45] Pradeepkiran, J. A., Bhaskar, M., Shrikanya, K. V. L., Krishna, P. G., Reddy, M. H., Venkatrayulu, C., and Sainath, S. B. (2021). Introduction to brucellosis. In *Brucella Melitensis*. Academic Press. Pp. 1-23.
- [46] Rahimi, H., Tukmechi, A., and Rashidian, E. (2020). Use of touch down polymerase chain reaction to enhance the sensitivity of *Brucella melitensis* detection in raw milk. *Animal Biotechnology*, 1 6.
- [47] Rahman, A. A., Dirk, B., Fretin, D., Saegerman, C., Ahmed, M. U., Muhammad, N., and Abatih, E. (2012). Seroprevalence and risk factors for brucellosis in a high risk group of individuals in Bangladesh. *Foodborne Pathogens and Disease*, *9*(3), 190 197.

- [48] Saxena, H. M. (2021). Bacteriophage and its Potential for Therapeutic Use in Brucellosis among Cattles. Research and Reviews: Journal of Veterinary Science and Technology. 2021; 10 (2): 9–17p. *Bacteriophage and its Potential for Therapeutic Hari Mohan Saxena STM Journals*, 2.
- [49] Seleem, M. N., Boyle, S. M., and Sriranganathan, N. (2010). Brucellosis: a reemerging zoonosis. *Veterinary microbiology*, 140(3 4), 392 398.
- [50] Sergueev, K. V., Filippov, A. A., and Nikolich, M. P. (2017). Highly sensitive bacteriophage based detection of *Brucella abortus* in mixed culture and spiked blood. *Viruses*, *9*(6), 144.
- [51] Shareef, J. M. (2006). A review of serological investigations of brucellosis among farm animals and humans in northern provinces of Iraq (1974–2004). *Journal of Veterinary Medicine, Series B*, 53, 38 40.
- [52] Shatti, A. A., Al-Rammah, H. S., and Zakair, K. Y. (2021). Serological detection of Brucella mellitensis in human and goats. *Biochemical and Cellular Archives*, 21(1), 777-782.
- [53] Sofian, M., Aghakhani, A., Velayati, A. A., Banifazl, M., Eslamifar, A., and Ramezani, A. (2008). Risk factors for human brucellosis in Iran: a case–control study. *International journal of infectious diseases*, 12(2), 157 161.
- [54] Sulayman, S. M. A., Bora, R. S., Sabir, J. S., and Ahmed, M. M. M. (2020). Brucellosis: current status of the disease and future perspectives. *Postępy Mikrobiologii-Advancements of Microbiology*, 59(4), 337-344.
- [55] Tevdoradze, E., Farlow, J., Kotorashvili, A., Skhirtladze, N., Antadze, I., Gunia, S., and Kutateladze, M. (2015). Whole genome sequence comparison of ten diagnostic brucellaphages propagated on two Brucella abortus hosts. *Virology journal*, *12*(1), 1-11.
- [56] Tumwine, G., Matovu, E., Kabasa, J. D., Owiny, D. O., and Majalija, S. (2015). Human brucellosis: sero prevalence and associated risk factors in agro pastoral communities of Kiboga District, Central Uganda. *BMC public health*, 15(1), 1 8.
- [57] Ulu Kilic, A., Metan, G., and Alp, E. (2013). Clinical presentations and diagnosis of brucellosis. *Recent patents on anti-infective drug discovery*, 8(1), 34-41.
- [58] Yumuk, Z., and O'Callaghan, D. (2012). Brucellosis in Turkey—an overview. *International Journal of Infectious Diseases*, 16(4), e228 e235.
- [59] Zhou, L., Fan, M., Hou, Q., Jin, Z., and Sun, X. (2018). Transmission dynamics and optimal control of brucellosis in Inner Mongolia of China. *Mathematical Biosciences and Engineering*, 15(2), 543.